UNIVERSIDADE FEDERAL DO PARANÁ

CAROLINA MELCHIOR DO PRADO

ASPECTOS EPIDEMIOLÓGICOS E MICROBIOLÓGICOS DA ESPOROTRICOSE TRANSMITIDA POR GATOS NA REGIÃO DA TRÍPLICE FRONTEIRA DO PARANÁ

CURITIBA

2025

CAROLINA MELCHIOR DO PRADO

ASPECTOS EPIDEMIOLÓGICOS E MICROBIOLÓGICOS DA ESPOROTRICOSE TRANSMITIDA POR GATOS NA REGIÃO DA TRÍPLICE FRONTEIRA DO PARANÁ

Tese apresentada ao Programa de Pós-Graduação em Microbiologia, Parasitologia e Patologia, Setor de Ciências Biológicas, Universidade Federal do Paraná, como requisito parcial para obtenção do título de Doutor em Microbiologia.

Orientador: Prof. Dr. Flávio de Queiroz Telles Filho

CURITIBA 2025

DADOS INTERNACIONAIS DE CATALOGAÇÃO NA PUBLICAÇÃO (CIP) UNIVERSIDADE FEDERAL DO PARANÁ SISTEMA DE BIBLIOTECAS – BIBLIOTECA DE CIÊNCIAS BIOLÓGICAS

Prado, Carolina Melchior do, 1993-Aspectos epidemiológicos e microbiológicos da esporotricose transmitida por gatos na região da tríplice fronteira do Paraná / Carolina Melchior do Prado. – Curitiba, 2025. 1 recurso on-line : PDF.
Tese (Doutorado) – Universidade Federal do Paraná, Setor de Ciências Biológicas, Programa de Pós-Graduação em Microbiologia, Parasitologia e Patologia. Orientador: Prof. Dr. Flávio de Queiroz Telles Filho.
1. Esporotricose. 2. Farmacorresistência fúngica. 3. Zoonoses.
4. Epidemiologia. I. Telles Filho, Flávio Queiroz. II. Universidade Federal do Paraná. Setor de Ciências Biológicas. Programa de Pós-Graduação em Microbiologia, Parasitologia e Patologia. III. Título.

Bibliotecária: Giana Mara Seniski Silva. CRB-9/1406



MINISTÉRIO DA EDUCAÇÃO SETOR DE CIÊNCIAS BIOLÓGICAS UNIVERSIDADE FEDERAL DO PARANÁ PRÓ-REITORIA DE PÓS-GRADUAÇÃO PROGRAMA DE PÓS-GRADUAÇÃO MICROBIOLOGIA, PARASITOLOGIA E PATOLOGIA - 40001016044P0

TERMO DE APROVAÇÃO

Os membros da Banca Examinadora designada pelo Colegiado do Programa de Pós-Graduação MICROBIOLOGIA, PARASITOLOGIA E PATOLOGIA da Universidade Federal do Paraná foram convocados para realizar a arguição da tese de Doutorado de CAROLINA MELCHIOR DO PRADO, intitulada: Aspectos epidemiológicos e microbiológicos da esporotricose transmitida por gatos na Região da Tríplice Fronteira do Paraná., sob orientação do Prof. Dr. FLÁVIO DE QUEIROZ TELLES FILHO, que após terem inquirido a aluna e realizada a avaliação do trabalho, são de parecer pela sua APROVAÇÃO no rito de defesa.

A outorga do título de doutora está sujeita à homologação pelo colegiado, ao atendimento de todas as indicações e correções solicitadas pela banca e ao pleno atendimento das demandas regimentais do Programa de Pós-Graduação.

Curitiba, 27 de Março de 2025.

Assinatura Eletrônica 12/04/2025 01:26:51.0 FLÁVIO DE QUEIROZ TELLES FILHO Presidente da Banca Examinadora

Assinatura Eletrônica 07/04/2025 15:02:26.0 VANIA APARECIDA VICENTE Avaliador Interno (UNIVERSIDADE FEDERAL DO PARANÁ) Assinatura Eletrônica 22/04/2025 09:12:26.0 MARCONI RODRIGUES DE FARIAS Avaliador Externo (PONTIFICA UNIVERSIDADE CATÓLICA DO PARANA)

Assinatura Eletrônica 03/04/2025 10:32:02.0 EELCO FJ MEIJER Avaliador Externo (CANISIUS-WILHELMINA HOSPITAL) Assinatura Eletrônica 03/04/2025 11:30:16.0 REGIELLY CAROLINE RAIMUNDO COGNIALLI Avaliador Externo (COMPLEXO HOSPITAL DE CLÍNICAS - CHC-UFPR)

I dedicate this work to my feline patients, the victims of sporotrichosis, who suffer from this disease every day. It is for them that my fight continues in the search for better treatments and prevention solutions.

ACKNOWLEDGMENTS

To Dr. Flávio, my dear advisor, my eternal professor, who ignited the flame of mycology within me and nurtured my growth in it. Your ethics and vision of what science should be, your consideration for patients, and your mastery in teaching us, your students, will be eternal inspirations for me wherever I go. Being a "Flaviana" and able to show the world that I was a student of one of the greatest references in Medical Mycology in Latin America will always be my pride and honor.

To my dear dream team at the Radboudumc-CWZ Center of Expertise for Mycology, Dr. Eelco, Dr. Theun, and Bram, first for welcoming me so kindly into my now beloved Netherlands and for guiding me on the path of better science. You have transformed me into a better researcher, and I will always remember how I arrived with so many questions in my head and how I left with even more, hahaha — though better and more interesting questions. I look forward with hope and great enthusiasm to the future of our collaboration, but in any case, it will always be an honor and a great privilege to have been able to work and learn with you.

To Dr. Luciana and Dr. Carlos from the Zoonoses Control Center of Foz do Iguaçu, there are no words that can adequately express my gratitude to you, my colleagues, who shared the (often exhausting and challenging) routine of searching for sporotrichosis cases in Foz. You helped and saved so many patients through proper treatment and guidance for their owners. You challenged the system and helped improve it. Your commitment to Public Health and patient well-being is a great inspiration. All the merit of this research we conducted over three years belongs to us, and I feel very grateful to share this victory with people I admire so much.

To my dear and dedicated Paraguayan team, my dear friends and colleagues: José Pereira from the Ministry of Public Health and Social Welfare of Paraguay, who opened the doors of this wonderful country to us and was always a problem-solver in the development of our research. To my dear Gabi, Nancy, and Leti from the National University of the East, who fought alongside me and never spared any effort in the (always challenging) search for human and feline cases. If today human and feline sporotrichosis is being discussed on the Paraguayan border, be assured that the merit is yours.

To the Argentine researchers from the National Institute of Tropical Medicine ANLIS -"Carlos G. Malbrán," for allowing my collaboration in the incredible project they carry out on the Argentine side of the triple border.

To the Brazilian, Paraguayan, and Argentine veterinarians from the private sector, who on many occasions helped me not only in the search and support for correct diagnosis but also in the proper care and treatment of patients.

To Professor Vânia, who always opened the doors of the laboratory to me, taught me so much, and continues to be an example for the mycologist I am becoming. I will never forget the day you messaged me about the scholarship abroad and how you helped me through the entire process. If today I aspire to greater things, it is certainly thanks to you.

To Dr. Jacques for his guidance, friendship, trust, and valuable teachings.

To my dear colleagues and friends from the Mycology Laboratory at UFPR — Marlon, Gui, Izadora, Bruno, Emanuel, Bruna, Germana, and Jason — who taught me everything from preparing media to sequencing calmodulin and really everything in between (which is no small thing). I also thank the dear Regi, Lili, Marison, and Adri from the Mycology Laboratory at CHC-UFPR for teaching me much of what I know about fungal identification, including *Sporothrix*.

To Professor Juliana, who helped me at a critical moment in my life and certainly played a crucial role in the future of my journey as a researcher. Your patience and kindness will always be remembered wherever I go, as will my eternal gratitude.

To Professors Walfrido and Rafaella for opening the doors of UNILA to me and truly enabling this research to take place.

To my Dutch friends and colleagues Dirk, Henriëtte, Sonia, Winny, and Tess, who made me feel at home. To my dear Jean, Andréia, and Flávia, for the moments of laugh in recent months. To my partners at the CDC, Amanda, Alisson, and Dallas for the interesting talks.

To my family, especially my mother Solange and my father Geraldo, who always encouraged me to study and improve myself, who always gave me the freedom to choose my path, supported my dreams, and provided me with the security of having a place to return to.

To my feline and canine patients, who motivate me to continue the fight.

Finally, my heartfelt thanks to Dr. Herinne for her vital help in difficult times.

Hou je stevig vast aan de mooie dingen in het leven

RESUMO

Sporothrix brasiliensis, um patógeno fúngico emergente na América Latina, é o agente da esporotricose transmitida por gatos, uma epi-zoonose que afeta humanos, gatos e cães. Este estudo apresenta os primeiros casos autóctones de esporotricose felina em Ciudad del Este, Paraguai. Além disso, descreve-se o primeiro caso autóctone de esporotricose humana causada por S. brasiliensis no Paraguai, transmitida por um gato paraguaio infectado. O diagnóstico e o tratamento tardios ressaltam a necessidade de maior conscientização sobre a doença. Também descreve-se o surgimento de casos urbanos de esporotricose felina em Puerto Iguazú, Argentina. Para melhor compreender a resistência antifúngica, analisou-se 108 isolados de S. brasiliensis de casos felinos na região da tríplice fronteira (2021-2023). Os testes de susceptibilidade antifúngica (AFST) contra nove antifúngicos mostraram baixos níveis de concentração inibitória mínima (MIC) para a maioria dos agentes, exceto por algumas cepas que apresentaram valores anormais (non-wild *type*) de MIC para terbinafina. Notavelmente, a esporotricose cutânea disseminada foi a apresentação clínica predominante (61%) e a única forma associada a alta mortalidade (61%). Nossos achados indicam que os diferentes desfechos clínicos não são causados por resistência pré-existente ao itraconazol. Dado o avanço geográfico da S. brasiliensis, uma abordagem de Saúde Única (One Health) é essencial para controlar essa zoonose emergente por meio de vigilância, educação pública e políticas de saúde direcionadas.

Palavras-chave: *Sporothrix brasiliensis*; Esporotricose Felina; Resistência Antifúngica; One Health; Zoonose.

ABSTRACT

Sporothrix brasiliensis, an emerging fungal pathogen in Latin America, is the causative agent of cat-transmitted sporotrichosis, an epi-zoonosis affecting humans, cats, and dogs. This study presents the first autochthonous cases of feline sporotrichosis in Ciudad del Este, Paraguay. Additionally, it describes the first autochthonous case of human sporotrichosis caused by S. brasiliensis in Paraguay, transmitted by an infected Paraguayan cat. Late diagnosis and treatment highlight the need for greater awareness of the disease. The emergence of urban cases of feline sporotrichosis in Puerto Iguazú, Argentina, is also described. To better understand antifungal resistance, 108 S. brasiliensis isolates from feline cases in the triple border region (2021-2023) were analyzed. Antifungal susceptibility testing (AFST) against nine antifungals showed low minimum inhibitory concentration (MIC) levels for most agents, except for some strains that exhibited abnormal (non-wild *type*) MIC values for terbinafine. Notably, disseminated cutaneous sporotrichosis was the predominant clinical presentation (61%) and the only form associated with high mortality (61%). Our findings indicate that different clinical outcomes are not caused by pre-existing resistance to itraconazole. Given the geographic spread of S. brasiliensis, a One Health approach is essential to control this emerging zoonosis through surveillance, public education, and targeted health policies.

Keywords: Sporothrix brasiliensis; Feline Sporotrichosis; Antifungal Resistance; One

Health; Zoonosis.

LIST OF FIGURES

CHAPTER 1

- Figure 1. Lesions in the cranial region and a small nasal deformity (A), ulcerated lesion in the left hind limb (B) and ulcerated lesions in the initial and medial portion of the tail (C) found after physical examination of animal 1. Nasal deformity and ulcerated lesion (D) found after physical examination of the animal 2.
- Figure 3. (A) Cigar-shaped yeast structures (black arrow) observed in feline samples (Gram-stain, 1000×). (B) Sporothrix colony after growth on Sabouraud agar with chloramphenicol. (C) Micromorphology of Sporothrix spp. colonies (black arrow) (lactophenol cotton blue ×400)......24

CHAPTER 2

- Figure 1. Clinical image across different time points. First consult on April 2 (Figure 1A, 1B), evolution to 13 swollen and painful nodules on May 3 (Figure 1C), consult with infectious disease specialist on May 11, (Figure 1D), itraconazole treatment for 6 days on May 16 (Figure 1E), for 18 days on May 28 (Figure 1F), for 24 days on June 3 (Figure 1G), for 50 days on June 29 (Figure 1H), for 59 days on July 8 (Figure 1I), for 89 days on August 7 (Figure 1J), and for 112 days on August 30 (Figure 1K).

CHAPTER 3

Figure 1. Map of Puerto Iguazú, Misiones, Argentina, located at the tri-border with Brazil and Paraguay, showing the spatial distribution of positive cases of feline sporotrichosis confirmed (n=9; pink) and ruled out cases (n=12; yellow) in the different neighborhoods studied. For those areas where multiple cases (positive and negative) overlap in location, the number of individual cases is indicated. The cases of where intrahousehold transmission occurred between two cats (FSCMi24 -FSCMi25 and FSCMi41 - FSCMi43) are indicated by an *. In the spatial analysis, the distance between households with feline sporotrichosis using cases was measured nearest-neighbor calculation in ArcGIS Pro, which calculates the shortest distance

CHAPTER 4

LIST OF TABLES

CHAPTER 2

CHAPTER 3

CHAPTER 4

Table 1. Prevalence of sporotrichosis per district63
Table 2. Characteristics of 108 cats with sporotrichosis 63
Table 3. Outcome of 108 cats with sporotrichosis based on clinical form64
Table 4. Treatment regime and duration of 50 clinical cured cats with sporotrichosis
based on clinical form64
Table 5. Treatment regime and duration of 40 dead cats with sporotrichosis 65
Table 6. Distribution of minimum inhibitory concentration (MIC) values against 108
clinical isolates of Sporothrix brasiliensis according to CLSI M38 and
M27 guidelines. M lines refer to mycelial phase and Y lines refer to
yeast phase. MICs in μg/mL67

SUMMARY

INTRODUCTION	16
1 CHAPTER 1 - FIRST CASES OF FELINE SPOROTRICHOSIS CAUSED BY	
SPOROTHRIX BRASILIENSIS IN PARAGUAY	18
1.1 ABSTRACT	19
1.2 INTRODUCTION	19
1.3 CASE PRESENTATION	20
1.4 DISCUSSION	27
1.5 REFERENCES	32
2 CHAPTER 2 - THE FIRST AUTOCHTHONOUS HUMAN CASE OF	
SPOROTRICHOSIS BY SPOROTHRIX BRASILIENSIS IN PARAGUAY	37
2.1 ABSTRACT	38
2.2 INTRODUCTION	38
2.3 CASE REPORT	38
2.4 DISCUSSION	43
2.5 REFERENCES	44
3 CHAPTER 3 - EMERGENCE OF FELINE SPOROTRICHOSIS IN PUERTO	
IGUAZÚ (ARGENTINA) NEAR THE ARGENTINA-BRAZIL BORDER	47
3.1 ABSTRACT	48
3.2 TEXT	48
3.3 REFERENCES	54
4 CHAPTER 4 - ABSENCE OF INITIAL ELEVATED ITRACONAZOLE MINIMU	JM
INHIBITORY CONCENTRATIONS IN SPOROTHRIX BRASILIENSIS FROM	
FELINES WITH TREATMENT FAILURE	
4.1 ABSTRACT	
4.2 TEXT	58
4.3 MATERIALS AND METHODS	59
4.3.1 Isolate and data collection	59
4.3.2 Diagnosis	59
4.3.3 Short tandem repeat genotyping	60
4.3.4 Antifungal susceptibility testing (AFST)	60
4.3.5 Ethics	61
4.4 RESULTS	61

4.4.1 Clinical epidemiology	62
4.4.2 Phylogenetic analysis	65
4.4.3 Minimum inhibitory concentration investigation	66
4.5 DISCUSSION	68
4.6 REFERENCES	79
CONCLUSION	87
REFERENCES	. 88

INTRODUCTION

Sporotrichosis is a neglected disease that primarily affects individuals with low economic and social status, living in precarious conditions and regions lacking basic sanitation. Domestic cats are the most affected victims of the disease, often abandoned or euthanized due to a lack of proper diagnosis and treatment. This factor is a direct consequence of the population's and veterinarians' lack of awareness about the disease. Additionally, an expanding epizootic outbreak continues to affect human and feline patients in various Brazilian regions, with the potential to reach neighboring countries. Therefore, understanding the disease in border regions is crucial for developing a control and prevention program and for constructing specific strategies for these locations.

This thesis addresses, in four chapters, an investigation of cases of Cattransmitted sporotrichosis (CTS) caused by *Sporothrix brasiliensis* in the triple border region (TBR) between Brazil, Paraguay, and Argentina.

- The first chapter reports the first cases of feline sporotrichosis in Paraguay, where resident cats from Ciudad del Este presented with severe ulcerative lesions, respiratory symptoms, and nasal deformities. Molecular analysis confirmed the presence of *S. brasiliensis*.
- The second chapter describes the first autochthonous case of human sporotrichosis in Paraguay. A 39-year-old woman from Presidente Franco developed progressive skin lesions after being scratched by her cat. The diagnosis was initially mistaken for a bacterial infection, highlighting the need for greater awareness of the disease.
- The third chapter addresses a significant outbreak on the Argentine side, in the city of Puerto Iguazú, Misiones. Surveillance between August 2023 and February 2024 detected *S. brasiliensis* in 42.8% of sampled cats, suggesting a transmission dynamic similar to that observed in the Brazilian epidemic.
- The fourth chapter focuses on the analysis of 108 S. brasiliensis isolates from cats before antifungal treatment. Antifungal susceptibility tests indicated that most strains had low minimum inhibitory concentration (MIC) levels for itraconazole, suggesting that treatment failures were not due to pre-existing drug resistance. Clinical outcomes of the cats were also analyzed, revealing that those with disseminated cutaneous sporotrichosis had a 61% mortality

rate, while those with fixed cutaneous or extracutaneous forms did not die from the disease. This suggests that disease severity and progression significantly influence treatment outcomes. The study highlights the rapid spread of *Sporothrix brasiliensis* in the TBR, with cases initially concentrated in the eastern region of Foz do Iguaçu, Brazil, and later spreading to areas near the international bridges. Regarding genetic analysis, genotyping through Short Tandem Repeat (STR) revealed that all isolates from the TBR are closely related to the Rio de Janeiro (RdJ) clade, a dominant *S. brasiliensis* genotype group. This suggests that the pathogen may have been introduced into the region through the movement of infected cats.

1 CHAPTER 1 - First Cases of Feline Sporotrichosis Caused by Sporothrix brasiliensis in Paraguay

Authors: Carolina Melchior do Prado¹, Emanuel Razzolini², Gabriela Santacruz³, Leticia Ojeda³, Marlon Roger Geraldo¹, Nancy Segovia³, José Pereira Brunelli⁴, Vânia Aparecida Vicente^{1,2}, Walfrido Kühl Svoboda⁵ and Flávio Queiroz-Telles^{6,*}

1 Postgraduate Program in Microbiology, Parasitology and Pathology, Biological Sciences, Department of Basic Pathology, Federal University of Parana, Curitiba 81531-980, Brazil

2 Postgraduate Program in Bioprocess and Biotechnology Engineering, Technology Sector, Department of Biotechnology, Federal University of Parana, Curitiba 81531-990, Brazil

3 Regional Epidemiological Laboratory, Faculty of Health Sciences, National University of the East, Minga Guazú 7420, Paraguay

4 Ministry of Public Health and Social Welfare, Asuncion 2160, Paraguay

5 Department of Public Health, Federal University of Latin American Integration, Foz do Iguaçu 85870-650, Brazil

6 Department of Public Health, Hospital de Clínicas, Federal University of Paraná, Curitiba 80060-900, Brazil

*Author to whom correspondence should be addressed.

J. Fungi 2023, 9(10), 972; https://doi.org/10.3390/jof9100972

Submission received: 28 August 2023 / Revised: 23 September 2023 / Accepted: 26 September 2023 / Published: 27 September 2023

(This article belongs to the Special Issue Fungal Infections of Implantation (Subcutaneous Mycoses), 2nd Edition)

1.1 Abstract

Sporothrix brasiliensis is an emerging fungal pathogen causing cat-transmitted sporotrichosis, an epi-zoonosis affecting humans, cats and dogs in Brazil and now spreading to neighboring South American countries. Here, we report the first two autochthonous cases of cat-transmitted sporotrichosis in Paraguay. The first case was a four-year-old male cat showing several ulcerative lesions, nasal deformity and respiratory symptoms. The second case was a one-year-old male cat showing a single ulcerated lesion, respiratory symptoms and nasal deformity. Both cases were admitted to a veterinary clinic in Ciudad del Este, Paraguay. Isolates were recovered from swabs of the two cases. Using molecular methods, the isolates were identified as *S. brasiliensis*.

1.2 Introduction

Sporotrichosis, caused by fungi of the genus *Sporothrix*, is the most prevalent and globally distributed of the implantation mycoses [1]. Transmission is usually by transcutaneous or transmucosal traumatic inoculation of the fungus present in soil, plants and decomposing organic matter (sapronotic), and through enzootic (cat-to-cat/dog) and zoonotic (cat-to-human) routes. *Sporothrix schenckii* and *Sporothrix brasiliensis* are the most common etiological agents in human and animal diseases in Brazil [2,3].

Cat-transmitted sporotrichosis (CTS) is a neglected disease. Since the 1990s, this important zoonosis has emerged as a public health problem in Brazil, where it has been responsible for thousands of associated human and feline cases, which almost exclusively involve infections with *S. brasiliensis* [4]. Cases have quickly spread throughout Brazil and to other countries in South America [5]. In the enzootic (cat-to-cat, cat-to-dog) and zoonotic (cat-to-human) routes, transmission can occur through bites, scratches, contact with exudate from skin lesions and probably through respiratory droplets during cat sneezing [6,7].

In cats, clinical manifestations may include single or multiple ulcerative lesions on the skin, associated with enlarged lymph nodes and respiratory signs (mainly sneezing). Most commonly, cats can present a disseminated disease with multiple cutaneous lesions and mucosal involvement, especially the nasal mucosa. Conjunctival, oral and genital mucosa may also be affected [8]. Culture isolation is the gold standard technique for the diagnosis of sporotrichosis in animals. But since infected cats display a high fungal burden, the direct examination of yeast cells isolated from skin lesions is frequently used by veterinarians and microbiologists in clinical practice [6]. Molecular identification methods based on the sequence analysis of the calmodulin gene are required to identify the *Sporothrix* species [9].

In Paraguay, the only reported CTS cases so far occurred in 2017 among relatives who moved from Brazil with an infected cat. Laboratory tests performed identified the etiological agent as Sporothrix spp. (without molecular identification). This was the first record of a travel-associated case of Sporothrix outside Brazil or Argentina, documenting the potential for spread to new areas through the transport of infected cats [10]. Other cases supporting CTS's potential to spread were observed in the U.S. and U.K. In 2020, a Brazilian woman was diagnosed with CTS in Boston, U.S., after being bitten by a sick cat in Minas Gerais, Brazil [11]. More recently, researchers reported the first human and feline S. brasiliensis cases in the U.K., both involving people who had moved from Brazil [12]. In 2019, the World Health Organization published an international alert on the potential risk of transmission of infection by S. brasiliensis in neighboring countries [13]. Besides the spread of CTS in Brazil, sporotrichosis cases caused by S. brasiliensis have been observed in cats in other Latin American countries without previous exposure to Brazil [14,15]. In Argentina, for example, S. brasiliensis has already been described in cats from the provinces of Buenos Aires and Santa Cruz, located in the Argentinean Patagonian region. In Misiones, a province bordering Brazil, there was a single description of S. brasiliensis isolated from a human in 1986 [15]. In 2022, researchers from Chile described three cases of domestic and feral cats with S. brasiliensis in the Magallanes region of southern Chile. Those cases were the first isolation reports of S. brasiliensis in cats from Chile. Cases 1 and 2 came from the Río Verde County, a small rural village, and case 3 came from the city of Punta Arenas [14]. Here, we report the first autochthonous feline cases of CTS by S. brasiliensis in Paraguay (PY).

1.3 Case Presentation

This study was approved by the Committee for Ethics in Research of the Federal University of Parana (number—CAAE 52726021.8.0000.0102) and by the Animal Use Ethics Committee of the Agricultural Sciences Campus of the Federal University of Parana.

Ciudad del Este (CDE) (25°29'36" S and 54°40'18" W) is a city and district of PY, located in the extreme east of the country on the banks of the Paraná River. More specifically, it is situated on the Triple Border with Argentina and Brazil and has approximately 300,000 inhabitants. It is the capital of the Alto Paraná department, located 327 km from the country's capital—Asunción.

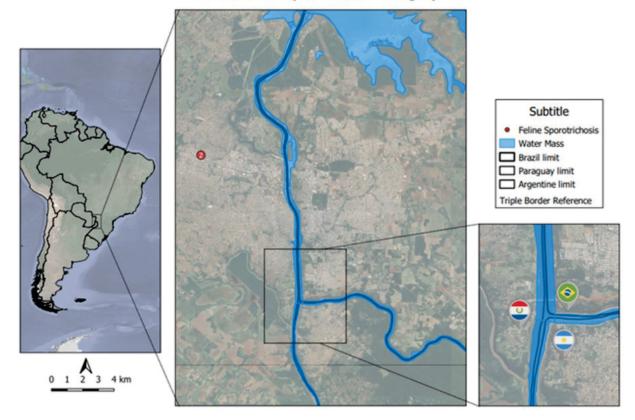
The first case observed in this city was a four-year-old mixed-breed male cat admitted on June 22 to a veterinary clinic in CDE, with numerous ulcerated lesions and respiratory symptoms. The animal had a history of respiratory signs (sneezing, nasal secretion and respiratory difficulty) for about twenty-one days and ulcerated lesions for about two months (beginning with lesions on the tail and later involving lesions on the face and paws). After a physical examination, four lesions in the cranial region (Figure 1A), a small nasal deformity (Figure 1A), one lesion in the left hind limb (Figure 1B) and four ulcerated lesions in the initial and medial portion of the tail (which had been amputated due to tissue necrosis) were found (Figure 1C).



Figure 1. Lesions in the cranial region and a small nasal deformity (A), ulcerated lesion in the left hind limb (B) and ulcerated lesions in the initial and medial portion of the tail (C) found after physical examination of animal 1. Nasal deformity and ulcerated lesion (D) found after physical examination of the animal 2.

Two months later (on August 22), a second case was admitted to the same veterinary clinic in CDE. It was a one-year-old mixed-breed male cat showing a single ulcerated lesion in the nose, respiratory symptoms (sneezing, nasal secretion and respiratory difficulty) and nasal deformation. The onset of clinical signs occurred eight days before the presentation at the clinic. After a physical examination, the only ulcerated lesion was confirmed, together with a significant nasal deformity (Figure 1D). This patient, who presented symptoms two months after the diagnosis of the first case, was from the same residence as the previous one.

In both cases, feline sporotrichosis was determined as the main diagnostic suspicion. Relevant clinical and epidemiological data were collected through clinical-epidemiological questionnaires applied to the owners (Figure A1 and Figure A2). The animals had a history of climbing and scratching trees and burying their feces outside. They were neutered and had no history of other previous illnesses. Both had had only their first rabies vaccine and access to the yard, the street and other residences. The owners reported that both animals had always lived in CDE and had no previous travel history. The geographic coordinates of the cats' residence were established using Google Earth® software. Cartographic bases from the Brazilian Institute of Geography and Statistics (IBGE) were used for georeferencing the coordinates. Quantus Gis software (QGIS) was used to assemble the map (Figure 2). The animals had not received any previous treatment with antibiotics and antifungals.



Case of Feline Sporotrichosis - Paraguay

Figure 2. Spatial distribution of the two animals with a positive diagnosis for sporotrichosis in Ciudad del Este, Paraguay.

In the first case, the patient was referred for euthanasia due to his poorly clinical status. In the second case, 100 mg/day itraconazole and 30 mg/day potassium iodide therapy were applied for about eight months until the clinical signs disappeared and were maintained for another two months to minimize the risk of recurrence.

For both cases, swab samples of the lesions and slides with an imprint of the lesions were collected to perform the definitive diagnosis. Mycological examination of the slide samples was performed based on direct Gram-stain microscopy, which enabled visualizing cigar-shaped yeast structures in the feline samples (Figure 3A). Considering that culture is the gold standard for the diagnosis of sporotrichosis [6], swabs collected from the wounds were placed in Stuart's medium and sent to the laboratory for mycological cultures. The samples were cultivated on Sabouraud dextrose agar (SDA) (KASVI, ttps://www.kasvi.com.br, accessed on 9 June 2022) containing chloramphenicol and incubated at 25 °C for the growth of colonies for up to 10 days.

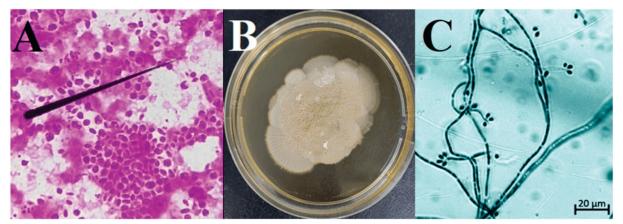


Figure 3. (A) Cigar-shaped yeast structures (black arrow) observed in feline samples (Gram-stain, 1000×). (B) *Sporothrix* colony after growth on Sabouraud agar with chloramphenicol. (C) Micromorphology of *Sporothrix* spp. colonies (black arrow) (lactophenol cotton blue ×400).

Macroscopic and microscopic characteristics of the colonies were observed and confirmed the fungi of the genus *Sporothrix* identity. The clinical samples showed growth of a white, creamy colony at 25 °C (Figure 3B). For the identification of the mycelial phase, monosporic cultures were grown on SDA at room temperature. We used the slide culture method to characterize microscopic filamentous features of the *Sporothrix* spp. isolates. We inoculated mycelia fragments into 1 × 1 cm SDA blocks, which were then mounted on a slide, covered with a coverslip and incubated for 14–21 days at room temperature. The mycelial preparation was stained with lactophenol blue and visualized with a Zeiss AxioObserver Z1 microscope equipped with a 40× objective. The microscopical analysis showed hyaline hyphae and septate with rounded unicellular conidia (Figure 3C).

Molecular analysis of the two isolates was based on calmodulin (CAL) gene sequences. For molecular identification, colonies cultivated on Sabouraud glucose agar (SGA) were used for DNA extraction. The fungi mycelium was transferred to a 1.5 mL tube containing 100 mg of a silica celite mixture (2:1, w/w) and 300 µL CTAB buffer [CTAB 2% (w/v), NaCl 1.4 M, Tris-HCl 100 mM, pH 8.0; EDTA 20 mM]. Fungi cells were disrupted with a pestle for 5 min and incubated for 40 min at 65 °C. Then, 500 µL 24:1 chloroform: isoamyl alcohol (CIA) was added and the solution was centrifuged for 10 min at 20,500× g. The supernatant was collected in a new tube with 2 vols of ice-cold 96% ethanol. The DNA was precipitated for 2 h at -20 °C and then centrifuged for 10 min at 20,500× g, washed with cold 70% ethanol, dried at room temperature and resuspended in 100 µL in ultrapure water [16]. DNA integrity was assessed by running a 5 µL aliquot of the extraction on a 0.8% agar agarose-gel electrophoresis stained with 0.5 μ g/ mL ethidium bromide. To estimate the DNA yield, we used a NanoDrop 1000c Instrument (Thermo Fisher Scientific, Waltham, MA, USA). Amplification of the CAL was performed using CL1 and CL2A primers [17]. PCR reaction mixtures were made with a total volume of 12.5 µL (1× PCR buffer, 2.0 mM MgCl2, 25 µM deoxynucleotide triphosphates (dNTPs)), 0.5 µM of each forward and reverse primer, 1 U of Tag DNA polymerase (Ludwig Biotec, Bela Vista, Brazil) and 10 ng of genomic DNA. The PCR parameters for amplification were 94 °C for 3 min, followed by 35 cycles consisting of 94 °C for 35 s, 58 °C for 30 s and 72 °C for 1 min, and a delay at 72 °C for 1 min, performed in an ABI Prism 2720 thermocycler (Applied Biosystems, Foster City, CA, USA). Amplicons were sequenced with BigDye Terminator cycle sequencing kit v. 3.1 (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's instructions, using the same primers of the PCR, and the amplification conditions were as follows: 95 °C for 1 min, followed by 30 cycles consisting of 95 °C for 10 s, 50 °C for 5 s and 60 °C. The consensus sequence of the

CAL regions was visually inspected using MEGA v.7 software [18] and compared to the GenBank Blast (NCBI). The sequences of CAL were aligned with reference strains (Table A1) using the online MUSCLE interface and the substitution model was selected for each genus using the MEGA software (GTR + G). The chosen index was the one that presented the lowest AIC. The phylogeny was constructed based on the CAL gene. Maximum likelihood analysis was performed in the RaxML-HPC2 [19] implemented in the CIPRES server using the 1000 bootstrap to reach split frequencies of \leq 0.01. The resulting trees were plotted in FigTree v.1.4.2 [20]. The results showed that the clinical strains were *Sporothrix brasiliensis* (Figure **3**4).

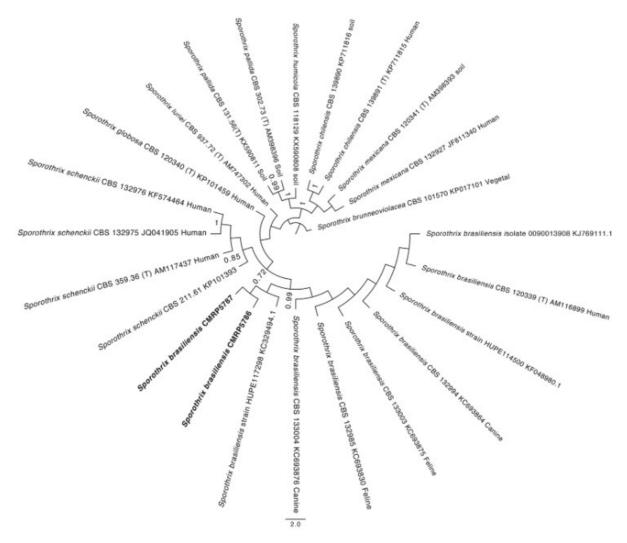


Figure 4. Phylogenetic tree of *Sporothrix brasiliensis*, obtained by ML analysis based on partial sequences of the CAL. The isolates CMRP5786 and CMRP5787 represent the isolates of Paraguay. Reference strains used can be seen in Table A1.

1.4 Discussion

These are the first feline cases of CTS in PY. Human sporotrichosis is a deep mycosis, relatively common in PY. In PY, this disease is still mostly related to the classic form of transmission, the sapronotic route, and has the greatest effect on certain professions such as carpenters, gardeners and farmers [10].

Ciudad del Este is situated near the Iguazu Falls at the busiest frontier in Brazil and in the context of the triple international border between Brazil, Paraguay and Argentina. The flow of people, animals, plants and microorganisms, sometimes without proper inspection and monitoring, is constant. And in terms of the flow of people, Foz do Iguaçu, as an important tourist destination, presents other peculiarities, occupying the third place in the preference of foreign tourists arriving in the country. Therefore, it is necessary to reflect on how these international borders can contribute to the dissemination of microorganisms such as *S. brasiliensis* in view of the lack of specific public health policies in this region [21].

In recent decades, the domestic cat (*Felis catus*) has played an important role in the epidemiology of sporotrichosis due to its increased susceptibility to this disease and its role as a vehicle of transmission to humans through bites, scratches, sneezing and contact with secretions from infected animals [8]. Sporotrichosis in cats is generally systemic, showing disseminated lesions with a high burden of infectious yeast-like cells and which, without treatment, cause death [22]. Infected animals show disseminated cutaneous lesions characterized by ulcers and nodules with exudate, especially on the face. Upon licking or touching with the extremities, the agent is transferred to the paws or the mouth, facilitating contagion. This behavior, together with the high burden of mycotic elements in the wounds, facilitates contagion among cats and other animals such as dogs and humans [23].

Although the first animal was euthanized, it is important to note that sporotrichosis is treatable. The drugs indicated for sporotrichosis treatment are itraconazole, potassium iodide, terbinafine and amphotericin B. Itraconazole is the first choice for treatment, at a dose of 100 mg/day until the complete resolution of the lesions [24]. Treatment duration varies in the literature and is determined by the patient's clinical response. Furthermore, it is important to point out that the capacity for transmission by the infected cat reduces exponentially with the implementation of the treatment [15].

Furthermore, the animal's history of access to the streets should always be considered during the period of treatment, especially in endemic regions [23]. Therefore, it is worth highlighting the importance of considering sporotrichosis as a differential diagnosis for animals showing skin lesions due to the potential zoonotic risk of *S. brasiliensis* transmission [25].

Animals under suspicion of sporotrichosis should be isolated from other animals. Decontamination of the environment can be carried out with sodium hypochlorite (bleach) [8]. Injuries resulting from the typical behavior of cats in distress should be avoided through the application of correct animal restraint techniques and the use of personal protective equipment, such as procedure gloves, aprons and goggles [26]. This is especially important as this fungal species has a high transmission potential due to contact with secretions and exudates from the lesions of sick cats during treatment and handling procedures. Feline management methods based on the "Cat-Friendly Practices" philosophy can help prevent injuries, as they aim to keep the patient calm so that their handling is peaceful and safe [27].

Author Contributions

Conceptualization, C.M.d.P., V.A.V., W.K.S. and F.Q.-T.; writing—original draft preparation, C.M.d.P. and F.Q.-T.; writing—review and editing, C.M.d.P., V.A.V., W.K.S. and F.Q.-T.; investigation and data curation, C.M.d.P., E.R., G.S., L.O., M.R.G., N.S. and J.P.B. All authors have read and agreed to the published version of the manuscript.

Funding

The work of F. Queiroz-Telles received fellowships from CNPq (grant number 312811/2018–7), Brasilia, Brazil. The work was supported by the Araucaria Foundation (http://www.fappr.pr.gov.br/, accessed on 31 June 2021) (NAPI grant number 113/2020).

Institutional Review Board Statement

The study was conducted in accordance with the Declaration of Helsinki, and approved by the Committee for Ethics in Research of the Federal University of Parana (number—CAAE 52726021.8.0000.0102, 28 December 2021) and by the Animal Use Ethics Committee of the Agricultural Sciences Campus of the Federal University of Parana (protocol code 028/2021, 7 June 2021).

Informed Consent Statement

Informed consent was obtained from all subjects involved in the study.

Data Availability Statement

The data presented in this study are openly available in NCBI, accession number OR501574 and OR501573.

Acknowledgments

The authors acknowledge Saddy veterinary for clinical histories, sample submission and care for these cats. We also thank Amanda Santos and Lindsay Parnell for their suggestions and English review.

Conflicts of Interest

The authors declare no conflict of interest.

Appendix A

UNIVERSIDAD FEDERAL DE PARANÀ Prote. Dr. Fravo de Queroz Teles Filio – queroz teles guiol com tr Investguadra responsable. Cavoiria Mechine do Prazo – c.mecition mulgignali.com Laboration de Micologia R.M. Brade Camago, 26. Curitas, FR CDP: 80.060-240 / Tel: (41).380-7241, (49) 98619-6604 ESTUDIO REGIONAL SOBRE ESPORTATICOSIS FELINA – CUESTIONARIO CLÍNICO FECHA DE RECOGIDA: / / 2022. IDENTIFICACIÓN DEL ANIMAL: L SOBRE EL ANIMAL ANALIZADO:	UNIVERSIDAD FEDERAL DE DARANA Prote. Dr. Fisiko Se Quento: Teles Filito – quento: Ieles Quei don tor Investigadora telescolaria de la cesporta de la cespo				
1. Nombre:2. Género: () F () M 3. Edad:4. Raza:					
	reporte 7. ¿Bajo tratamiento antifúngico? () si, () no, medicamento				
5. Propósito: () compañía, () control de roedores () otro:	utilizado:, inicio del tratamiento:/_/2, () mejora, () empeora.				
6. ¿Se ha mudado? () sí, () no, ¿cuándo y cuál es el origen?	8. Tratamiento previo: () si, () no, medicación:, fin				
7. Lugar de captura (si es errante):	del tratamiento://2 9. Curación clínica: () si, () no. 10. Transmisión por: () gato doméstico,				
8. ¿Fotos? () Sí () No. 9. ¿Cuál es el estilo de vida del animal? () sedentario. () pelear con otros	() gato externo, () sin origen, () otro:, () estomudar, () lamer, () morder, () arañar.				
animales, () trepa o araña árboles, () araña o morder otros animales, () araña o morder otros humanos,	III. SOBRE CONTACTOS CON ANIMALES Y/O HUMANOS:				
() hábito de lamer, () entierran sua excrementos fuera de la casa, () usam caja de arena. 10. Historia	1. Casos confirmados de esporotricosis en el hogar: () si, otro animal, () si, humano () no. 2.				
clínica: () castrado, () FIV, () FeLV, () Rinotraqueítis, () Calicivirosis, () Panleucopenia, ()	Humano confirmado: () con lesión, clasificación de la lesión: () cutánea fija, () cutánea diseminada,				
enfermedades parasitarias, () Leishmaniasis, () enfermedades metabólicas, () Cánoer, () trasplante	() linfocutánea, () afección respiratoria, () oftálmica, tiempo de evolución:				
de sangre, () tratamiento por más de dos semanas con esteroides. 11. ¿Vacunación? () nunca	Transmisión por: () gato doméstico, () gato externo, () sin origen, () otro:,				
vacunado, () Primera vacunación solamente, () vacunación ocasional con más de 2 años de diferencia,	() secreción nasal, () contacto con secreción, () mordedura, () rasguño, () trauma por planta. Género:				
() vacunación anual o frecuente (cada 1-2 años), () desconocido. 12. ¿Acceso? () sin acceso externo,	() F. () M. Edad:, Raza:, Ocupación:,				
() el patio trasero, () la calle, () otras residencias, () errante. 13. Lugar de descanso: () dentro de la	3. Realizado tratamiento: () si, () no , medicación:4. Cura clínica: ()				
casa, () fuera de la casa (patio, área), () en la calle, () en otra residencia, () no pudo informar,	si, () no. 5. Alguien en el hogar tuvo: () leishmaniasis, () esquistosomiasis, () diarrea, () fiebre				
14. Alimentación: () comida de gato, () comida casera, () palomas, () ratones, () no sé, () otro:	amarilla, () dengue, () hepatitis, () leptospirosis, () malaria, () otro:,() ninguno.				
II. SOBRE LA ENFERMEDAD:	IV. DATOS DE LABORATORIO:				
1. ¿Es un caso asintomático? () si, () no. 2. Inicio del cuadro clínico:	1. Tipo de muestra:				
3. Signos de la piel: () 1 lesión, () 2, () 3, () 4, () 5, () 6 o más, () mucosa nasal () mucosa oral,	2. ¿Resultado de laboratorio? Micológico directo:				
() mucosa ocular () hocico, () orejas, () boca () ojos () en las patas () por el cuerpo 4. Signos	Cultura:				
extracutáneos: () estomudos, () tos, () secreción nasal, () deformidad nasal () dificultad para respirar	Secuenciación:				
() ganglios linfáticos inflamados () compromiso aparente de hueso o cartílago, () pérdida de peso, ()					

Figure A1. Clinical–epidemiological questionnaire applied to the owners.

UNIVERSIDAD FEDERAL DE PARANÁ Prote. Dr. Flavo de Gueiroz Telles Filino - gueiroz telles@uol.com.br Investigadora regornable: Carolina Microiro do Prado - mecinicia multiginali.com Lucionationo de Miccologia / Rua Padre Camargo, 260. Curtos, PR CES 80062407 / Tre. Lucionationa - CUESTIONARIO AMBIENTAL FECHA DE RECOGIDA:// 2022. IDENTIFICACIÓN DEL ANIMAL: L DATOS DEL RESPONSABLE DEL ANIMAL (SI LOS HUBIERA):	UNIVERSIDAD FEDERAL DE PARANA Prote. Dr. Flako de Queiroz Teles Pilio - queiroz lelesquoi.com tor Investgadora responsable. Carolina Mentorio de Padro - c. mentor metigomal.com Lacoratorio de Miciologia / Rua Padre Camago, 380. Curros, PR DE 50.006-2001 (r. (4) 5906 - 1506. (1, (4) 5906 - 1506. acceso: () sí, () no, () no supo responder. 12. Otros animales en la residencia: () sí, () no, en caso afirmativo, cuáles y cuántos:13. Presencia de bosque cerca de la residencia: () sí, () no, sin sí, el gato tiene acceso: () sí, () no, () no supo responder.
1. Nombre: 2. Teléfono:	14. Presencia de fauna silvestre: () sí, () no. 15. Distancia entre casas vecinas: () sin espaciamiento,
3. Dirección/Municipalidad:	() espaciamiento medio, () espaciamiento grande. 16. El gato tiene acceso a otras residencias: () sí,
4. Coordenada geográfica:	() no, () no supo responder. 17. Presencia de ratas: () sí, () no. 18. Presencia de animales callejeros:
II. ACERCA DEL MEDIO AMBIENTE:	() perros, () gatos, () otros:
1. Tipo: () casa de mampostería, () casa de madera, () casa de mampostería/madera, () apartamento,	III. SANEAMIENTO:
() "choza", () otro:, 2. Electricidad: () en el hogar, () en la vía pública.	1. Alcantarillado: () sistema de alcantarillado () pozo negro, () zanja, () arroyos/ríos, () al aire libre,
3. Calle: () pavimentado, () piso de tierra, () pozo/boca de lobo/canal. 4. ¿Servicios públicos en el	() no supo responder. 2. Hay puntos de fugas de alcantarillado cerca de la residencia: () sí, () no,
barrio?()guardenía,()colegio,()centro de salud,()otro:5.¿Tienes	() no supo responder. 2. Fuente de agua: () cisterna, () agua tratada, () pozo artesiano, () río. 3.
un patio trasero? () sí, () no, () suelo expuesto, () oésped, () jardín/macetas, () huerto, () árboles,	Lluvia: () inundación, () retorno de aguas residuales, () basura en los sistemas de drenaje, () otro:
() presencia de escombros, () materia orgánica acumulada. 6. El gato tiene acceso a: () suelo/tierra	4. ¿Vive cerca de un arroyo o río? () sí, () no, () en caso afirmativo, ¿hay
expuesta, () césped, () jardín/macetas, () huerta, () árboles, () presencia de escombros, () materia	vegetación en las orillas? () Sí () No. 5. Limpieza urbana: () no hay recolección de basura, ()
orgánica acumulada, () no supo responder. 7. Que clase de arbol hay en el traspatio: () mango, ()	recolección de basura, frecuencia: Si la basura no se recoge, cuál es el destino: ()
aguacate, () angico, () guayaba, () seriguela, () llorona, () limon, () naranja, () arete indio, () ficus,	basura quemada, () basura enterrada, () basura llevada al vertedero, () otro:
() chisme, () ipê de colores, () araucaria, () jabuticabeira, () plátano, () otros:	() recogida de reciclables, frecuencia:, () barrido, () poda de árboles, () recogida
8. Hay señales de raspaduras en los árboles: () sí, () no,	de animales muertos, () recogida de escombros. 6. Disposición de escombros: () recogidos por la
en caso afirmativo, en qué tipo de árbol:9. Zona: () urbana, () rural, () semirrural,	alcaldía, () tirados al suelo, () recogidos en basurero, () recogidos por carretones, () centro de reciclaje,
() periferia/favela. 10. Cría de mascotas en casa con fines comerciales: () si, () no., en caso afirmativo	() no pudo informar.
cuales y cuantos:, a que distancia de la casa?, tiene acceso	IV. OTROS COMENTARIOS RELEVANTES:
el gato: () si, () no, () no supo responder. 11. Cría de animales de producción: () si, () no, ()	
gallinero, () pocilga, () cunicultura, () bovinos, () equinos, () pequeños rumiantes, ()	
otros:, ¿cuál es la distancia de residencia?, el gato tiene	

Figure A2. Clinical–epidemiological questionnaire applied to the owners.

Species	Isolate code	CBS code	Geographic origin	Source	GenBank	Referen
						се
Sporothrix_	Ss159	CBS_132976	Japan	Human	KF574464	[28]
schenckii						
Sporothrix_	Ss141	CBS_132975	Brazil	Human	JQ041905	[29]
schenckii						
Sporothrix_	-	CBS_359.36_(T)	NK	Human	AM117437	[30]
schenckii						
Sporothrix_	-	CBS_211.61	South Africa	Not Know	KP101393	[31]
schenckii						
Sporothrix_	-	CBS_302.73_(T)	United Kingdom	Soil	AM398396	[32]
pallida						
Sporothrix_	-	CBS_131.56(T)	Japan	Soil	KX590811	[33]
pallida						
Sporothrix_	Ss 132	CBS_132927	Brazil	Human	JF811340	[34]
mexicana						
Sporothrix_	FMR 9108	CBS_120341_(T)	Mexico	Soil	AM398393	[32]
mexicana						
Sporothrix_	ATCC 18616	CBS_937.72_(T)	South Africa	Human	AM747302	[35]
luriei						
Sporothrix_	-	CBS_118129	South Africa	Soil	KX590808	[33]
humicola						
Sporothrix_	-	CBS_120340_(T)	Spain	Human	KP101459	[31]
globosa						
Sporothrix_	Ss469	CBS_139891_(T)	Chile	Human	KP711815	[36]
chilensis						
Sporothrix_	Ss470	CBS_139890	Chile	Soil	KP711816	[36]
chilensis						
Sporothrix_	-	CBS_101570	USA	Vegetal	KP017101	[31]
brunneoviol						
acea						
Sporothrix_	Ss227	CBS_133004	Brazil	Canine	KC693876	[37]
brasiliensis						
Sporothrix_	Ss226	CBS_133003	Brazil	Feline	KC693875	[37]
brasiliensis						
Sporothrix_	Ss151	CBS_132994	Brazil	Canine	KC693864	[37]
brasiliensis						
Sporothrix	0090013908	-	Brazil	Human	KJ769111	[38]

 Table A1. Reference strains used for CAL gene alignment.

brasiliensis						
Sporothrix	Ss05	CBS 132985	Brazil	Feline	KC693830	[37]
brasiliensis						
Sporothrix	HUPE11729	-	Brazil	Human	KC329494	[39]
brasiliensis	8					
Sporothrix	HUPE11450	-	Brazil	Human	KF048980	[40]
brasiliensis	0					
Sporothrix	IPEC 16490	CBS 120339 (T)	Brazil	Human	AM116899	[32]
brasiliensis						
Sporothrix	CMRP5787	-	Brazil	Feline	OR501574	This
brasiliensis						study
Sporothrix	CMPRP5786	-	Brazil	Feline	OR501573	This
brasiliensis						study

1.5 References

[1] Gonçalves, S.S.; da Cruz Bahiense Rocha, I.; Rediguieri, B.C.; de Carvalho, J.A.;
Maifrede, S.B.; Kruschewsky, W.L.L.; Falqueto, A.; Rodrigues, A.M. Human and
Feline Sporotrichosis in a Reference Center of Southeastern Brazil: Genetic
Differentiation, Diversity, and Antifungal Susceptibility of *Sporothrix* Species. J. Fungi
2023, 9, 831.

[2] Rossow, J.A.; Queiroz-Telles, F.; Caceres, D.H.; Beer, K.D.; Jackson, B.R.;
Pereira, J.G.; Gremião, I.D.F.; Pereira, S.A. A One Health Approach to Combatting *Sporothrix brasiliensis*: Narrative Review of an Emerging Zoonotic Fungal Pathogen in South America. J. Fungi 2020, 6, 247.

[3] Cognialli, R.C.R.; Cáceres, D.H.; Bastos, F.d.A.G.D.; Cavassin, F.B.; Lustosa,
B.P.R.; Vicente, V.A.; Breda, G.L.; Santos-Weiss, I.; Queiroz-Telles, F. Rising
Incidence of *Sporothrix brasiliensis* Infections, Curitiba, Brazil, 2011–2022. Emerg
Infect. Dis. 2023, 29, 1330–1339.

[4] Gómez-Gaviria, M.; Martínez-Álvarez, J.A.; Mora-Montes, H.M. Current Progress in *Sporothrix brasiliensis* Basic Aspects. J. Fungi 2023, 9, 533.

[5] Schechtman, R.C.; Falcão, E.M.M.; Carard, M.; García, M.S.C.; Mercado, D.S.;
Hay, R.J. Sporotrichosis: Hyperendemic by Zoonotic Transmission, with Atypical
Presentations, Hypersensitivity Reactions and Greater Severity. An. Bras. Dermatol.
2022, 97, 1–13.

[6] Rodrigues, A.M.; Della Terra, P.P.; Gremião, I.D.; Pereira, S.A.; Orofino-Costa,R.; de Camargo, Z.P. The Threat of Emerging and Re-Emerging Pathogenic*Sporothrix* Species. Mycopathologia 2020, 185, 813–842.

[7] Bastos, F.; Farias, M.; Monti, F.; Cognialli, R.; Lopuch, L.; Gabriel, A.; Vicente, V.; Razzolini, E.; Wu, K.; Queiroz-Telles, F. Spread of *Sporothrix brasiliensis* from the Sneeze of Infected Cats: A Potential Novel Route of Transmission. Med. Mycol. 2022, 60, 272.

[8] Gremião ID, F.; Martins da Silva da Rocha, E.; Montenegro, H.; Carneiro, A.J.B.; Xavier, M.O.; de Farias, M.R.; Lopes-Bezerra, L.M. Guideline for the Management of Feline Sporotrichosis Caused by *Sporothrix brasiliensis* and Literature Revision. Braz. J. Microbiol. 2021, 52, 107–124.

[9] Rodrigues, A.M.; Gonçalves, S.S.; de Carvalho, J.A.; Borba-Santos, L.P.;

Rozental, S.; Camargo, Z.P. de Current Progress on Epidemiology, Diagnosis, and Treatment of Sporotrichosis and Their Future Trends. J. Fungi 2022, 8, 776.

[10] García Duarte, J.M.; Wattiez Acosta, V.R.; Fornerón Viera PM, L.; Aldama
Caballero, A.; Gorostiaga Matiauda, G.A.; de Oddone, V.B.R.; Pereira Brunelli, J.G.
Esporotricosis Trasmitida Por Gato Doméstico. Reporte de Un Caso Familiar.
Revista del Nacional (Itauguá) 2017, 9, 67–76.

[11] Kaadan, M.I.; Dennis, M.; Desai, N.; Yadavalli, G.; Lederer, P. One HealthEducation for Future Physicians: A Case Report of Cat-Transmitted Sporotrichosis.Open Forum Infect. Dis. 2020, 7, ofaa049.

[12] Barnacle, J.R.; Chow, Y.J.; Borman, A.M.; Wyllie, S.; Dominguez, V.; Russell, K.;
Roberts, H.; Armstrong-James, D.; Whittington, A.M. The First Three Reported
Cases of *Sporothrix brasiliensis* Cat-Transmitted Sporotrichosis Outside South
America. Med. Mycol. Case Rep. 2023, 39, 14–17.

[13] World Health Organization (WHO). *Sporothrix brasiliensis*, an Emerging Fungal Pathogen, Notable for Its Zoonotic Transmission and Epidemic Potential for Human and Animal Health in the Americas; WHO: Geneva, Switzerland, 2019.

[14] Thomson, P.; González, C.; Blank, O.; Ramírez, V.; del Río, C.; Santibáñez, S.;
Pena, P. Sporotrichosis Outbreak Due to *Sporothrix brasiliensis* in Domestic Cats in
Magallanes, Chile: A One-Health-Approach Study. J. Fungi 2023, 9, 226.

[15] Etchecopaz, A.; Toscanini, M.A.; Gisbert, A.; Mas, J.; Scarpa, M.; Iovannitti,

C.A.; Bendezú, K.; Nusblat, A.D.; Iachini, R.; Cuestas, M.L. Sporothrix brasiliensis: A

Review of an Emerging South American Fungal Pathogen, Its Related Disease, Presentation and Spread in Argentina. J. Fungi 2021, 7, 170.

[16] Voidaleski, M.F.; Queiroz-Telles, F.; Itikawa, H.T.; Müller, G.G.; Lima, B.J.F.S.; Trevisoli, L.E.; Cognialli, R.C.R.; Crispim, R.C.L.; Vicente, V.A. An Atypical Etiology of Fungal Keratitis Caused by Roussoella neopustulans. J. Fungi 2022, 8, 507. [17] Rodrigues, A.M.; de Hoog, G.S.; de Camargo, Z.P. Molecular Diagnosis of Pathogenic Sporothrix Species. PLoS Negl. Trop. Dis. 2015, 9, e0004190. [18] Kumar, S.; Stecher, G.; Tamura, K. MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. Mol. Biol. Evol. 2016, 33, 1870–1874. [19] Stamatakis, A. RAxML Version 8: A Tool for Phylogenetic Analysis and Post-Analysis of Large Phylogenies. Bioinformatics 2014, 30, 1312–1313. [20] Rambaut, A. FigTree v1.4.2, a Graphical Viewer of Phylogenetic Trees. 2014. Available online: http://tree.bio.ed.ac.uk/software/figtree/ (accessed on 10 May 2023). [21] Prado, C.M.; Svoboda, W.K.; Chiyo, L.; Queiroz-Telles, F. Fundamentos de Saúde Única (One Health) e Planejamento Estratégico Situacional para Implementação de Política Pública de Saúde para Prevenção e Controle da Esporotricose na Região da Tríplice Fronteira (Brasil, Paraguai, Argentina). In Saúde Pública na Região da Fronteira Brasil-Paraguai-Argentina; Pedro & João Editores: São Carlos, Brasil, 2022; Volume 1, Chapter 5, p. 101.

[22] Lecca, L.O.; Paiva, M.T.; de Oliveira, C.S.F.; Morais, M.H.F.; de Azevedo, M.I.;
Bastos, C.D.V.E.; Keller, K.M.; Ecco, R.; Alves, M.R.S.; Pais, G.C.T.; et al.
Associated Factors and Spatial Patterns of the Epidemic Sporotrichosis in a High
Density Human Populated Area: A Cross-Sectional Study from 2016 to 2018. Prev.
Vet. Med. 2020, 176, 104939.

[23] Maschio-Lima, T.; Marques, M.D.R.; Lemes, T.H.; Brizzotti-Mazuchi, N.S.; Caetano, M.H.; de Almeida, B.G.; Bianco, L.M.; Monteiro, R.C.; Rodrigues, A.M.; de Camargo, Z.P.; et al. Clinical and Epidemiological Aspects of Feline Sporotrichosis Caused by *Sporothrix brasiliensis* and in Vitro Antifungal Susceptibility. Vet. Res. Commun. 2021, 45, 171–179.

[24] Rabello, V.B.S.; Almeida, M.A.; Bernardes-Engemann, A.R.; Almeida-Paes, R.;
de Macedo, P.M.; Zancopé-Oliveira, R.M. The Historical Burden of Sporotrichosis in
Brazil: A Systematic Review of Cases Reported from 1907 to 2020. Braz. J.
Microbiol. 2022, 53, 231–244.

[25] Silva, C.E.; Valeriano, C.A.T.; Ferraz, C.E.; Neves, R.P.; Oliveira, M.M.E.; Silva, J.C.A.L.; Magalhães, V.; Lima-Neto, R.G. Epidemiological Features and Geographical Expansion of Sporotrichosis in the State of Pernambuco, Northeastern Brazil. Future Microbiol. 2021, 16, 1371–1379.

[26] Lloret, A.; Hartmann, K.; Pennisi, M.G.; Ferrer, L.; Addie, D.; Belák, S.; Boucraut-Baralon, C.; Egberink, H.; Frymus, T.; Gruffydd-Jones, T.; et al. Sporotrichosis in Cats: ABCD Guidelines on Prevention and Management. J. Feline Med. Surg. 2013, 15, 619–623.

[27] Taylor, S.; St Denis, K.; Collins, S.; Dowgray, N.; Ellis, S.L.H.; Heath, S.; Rodan,
I.; Ryan, L. 2022 ISFM/AAFP Cat Friendly Veterinary Environment Guidelines. J
Feline Med. Surg. 2022, 24, 1133–1163.

[28] Sasaki, A.A.; Fernandes, G.F.; Rodrigues, A.M.; Lima, F.M.; Marini, M.M.;
Feitosa, L.D.S.; De Melo Teixeira, M.; Felipe, M.S.S.; Da Silveira, J.F.; De Camargo,
Z.P. Chromosomal Polymorphism in the *Sporothrix schenckii* Complex. PLoS ONE
2014, 9, e86819.

[29] Fernandes, G.F.; dos Santos, P.O.; Rodrigues, A.M.; Sasaki, A.A.; Burger, E.; de Camargo, Z.P. Characterization of Virulence Profile, Protein Secretion and Immunogenicity of Different *Sporothrix schenckii* Sensu Stricto Isolates Compared with S. Globosa and *S. brasiliensis* Species. Virulence 2013, 4, 241–249.

[30] Marimon, R.; Gené, J.; Cano, J.; Trilles, L.; Lazéra, M.D.S.; Guarro, J. Molecular Phylogeny of Sporothrix Schenckii. J. Clin. Microbiol. 2006, 44, 3251–3256.

[31] Zhang, Y.; Hagen, F.; Stielow, B.; Rodrigues, A.M.; Samerpitak, K.; Zhou, X.;

Feng, P.; Yang, L.; Chen, M.; Deng, S.; et al. Phylogeography and Evolutionary

Patterns in *Sporothrix* Spanning More than 14 000 Human and Animal Case Reports. Persoonia Mol. Phylogeny Evol. Fungi 2015, 35, 1–20.

[32] Marimon, R.; Cano, J.; Gené, J.; Sutton, D.A.; Kawasaki, M.; Guarro, J.;
Sporothrix Brasiliensis, S.G.; Mexicana, S. Three New *Sporothrix* Species of Clinical Interest. J. Clin. Microbiol. 2007, 45, 3198–3206.

[33] de Beer, Z.W.; Duong, T.A.; Wingfield, M.J. The Divorce of *Sporothrix* and Ophiostoma: Solution to a Problematic Relationship. Stud. Mycol. 2016, 83, 165–191.

[34] Rodrigues, A.M.; De Hoog, S.; De Camargo, Z.P. Emergence of Pathogenicity in the *Sporothrix schenckii* Complex. Med. Mycol. 2013, 51, 405–412.

[35] Marimon, R.; Genè, J.; Cano, J.; Guarro, J. *Sporothrix luriei*: A Rare Fungus from Clinical Origin. Med. Mycol. 2008, 46, 621–625.

[36] Rodrigues, A.M.; Cruz Choappa, R.; Fernandes, G.F.; de Hoog, G.S.; de Camargo, Z.P. *Sporothrix chilensis* Sp. Nov. (Ascomycota: Ophiostomatales), a Soil-Borne Agent of Human Sporotrichosis with Mild-Pathogenic Potential to Mammals. Fungal Biol. 2016, 120, 246–264.

[37] Rodrigues, A.M.; de Melo Teixeira, M.; de Hoog, G.S.; Schubach, T.M.P.; Pereira, S.A.; Fernandes, G.F.; Bezerra, L.M.L.; Felipe, M.S.; de Camargo, Z.P.

Phylogenetic Analysis Reveals a High Prevalence of *Sporothrix brasiliensis* in Feline Sporotrichosis Outbreaks. PLoS Negl. Trop. Dis. 2013, 7, e2281.

[38] Marques De Macedo, P.; Sztajnbok, D.C.N.; Camargo, Z.P.; Rodrigues, A.M.; Lopes-Bezerra, L.M.; Bernardes-Engemann, A.R.; Orofino-Costa, R. Dacryocystitis due to *Sporothrix brasiliensis*: A case report of a successful clinical and serological outcome with low-dose potassium iodide treatment and oculoplastic surgery. Br. J. Dermatol. 2015, 172, 1116–1119.

[39] Orofino-Costa, R.; Unterstell, N.; Carlos Gripp, A.; De Macedo, P.M.; Brota, A.;
Dias, E.; De Melo Teixeira, M.; Felipe, M.S.; Bernardes-Engemann, A.R.; Lopes-Bezerra, L.M. Pulmonary Cavitation and Skin Lesions Mimicking Tuberculosis in a
HIV Negative Patient Caused by *Sporothrix brasiliensis*. Med. Mycol. Case Rep. 2013, 2, 65–71.

[40] Castro, R.A.; Kubitschek-Barreira, P.H.; Teixeira, P.A.C.; Sanches, G.F.;
Teixeira, M.M.; Quintella, L.P.; Almeida, S.R.; Costa, R.O.; Camargo, Z.P.; Felipe,
M.S.S.; et al. Differences in Cell Morphometry, Cell Wall Topography and Gp70
Expression Correlate with the Virulence of *Sporothrix brasiliensis* Clinical Isolates.
PLoS ONE 2013, 8, e75656.

2 CHAPTER 2 - The first autochthonous human case of sporotrichosis by Sporothrix brasiliensis in paraguay

Authors: Mirtha Gabriela Santacruz Silvero^{1*}, Carolina Melchior do Prado^{2,3*}, Bram Spruijtenburg^{3,4,5}, Federico Augusto Lacarrubba Codas⁶, Maria Leticia Ojeda¹, Bruna Jacomel Favoreto de Souza Lima^{2,3}, Nancy Segovia Coronel¹, José Pereira Brunelli⁷, Vânia Aparecida Vicente², Theun de Groot^{3,4,5}, Flávio Queiroz-Telles^{2,8*}, Eelco F.J. Meijer^{3,4,5*}

1 Regional Epidemiological Laboratory, Faculty of Health Sciences, National University of the East, Minga Guazú, Paraguay.

2 Postgraduate Program in Microbiology, Parasitology and Pathology, Biological Sciences, Department of Basic Pathology, Federal University of Parana, Curitiba, Brazil.

3 Radboudumc-CWZ Center of Expertise for Mycology, Nijmegen, The Netherlands.

4 Department of Medical Microbiology, Radboudumc, Nijmegen, The Netherlands.

5 Canisius-Wilhelmina Hospital (CWZ)/Dicoon, Nijmegen, The Netherlands.

6 Departament of Infectious Deseases, Tesãi Fundation, Ciudad del Este, Paraguay.

7 Ministry of Public Health and Social Welfare, Asuncion, Paraguay.

8 Department of Public Health, Hospital de Clínicas, Federal University of Paraná, Curitiba, Brazil.

* Co-first authorship and co-senior authors was assigned based on equal contributions of both groups

Correspondence:

Eelco F.J. Meijer

Department of Medical Microbiology, Radboudumc, Weg door Jonkerbos 100, 6532 SZ, Nijmegen, The Netherlands.

Email: Eelco.meijer@radboudumc.nl

Journal of Medical Mycology, Volume 35, Issue 1, March 2025, 101536; https://doi.org/10.1016/j.mycmed.2025.101536

Received 29 November 2024, Revised 13 January 2025, Accepted 3 February 2025, Available online 4 February 2025, Version of Record 8 February 2025.

2.1 Abstract

Sporotrichosis *by Sporothrix brasiliensis* is increasingly reported in South America. Here, we present the first autochthonous human case in Paraguay, transmitted by a local infected cat. After 63 days of clinical signs onset, the patient was correctly diagnosed and antifungal treatment started, highlighting the need to increase awareness for this emerging disease.

Keywords: Sporotrichosis; *Sporothrix brasiliensis* infection; Diagnosis; Zoonotic Infectious diseases.

2.2 Introduction

Sporotrichosis is the most prevalent and globally distributed saprozoonotic implantation mycoses, previously referred to as a subcutaneous mycosis. Infections are caused by fungi of the genus *Sporothrix*. Generally, transmission occurs by traumatic inoculation in sapronotic, enzootic, or zoonotic routes. Since the 1990s sporotrichosis is hyperendemic in Brazil, with *S. brasiliensis* being the most common etiological agent in human and animal disease [1]. This major public health threat spreads by infected cats through exudates direct contact or respiratory droplets [2-4]. While the disease is common in Brazil, the prevalence in neighboring countries is steadily increasing, including Argentina and Chile [5,6].

From Paraguay, there has been one report from 2017 describing human sporotrichosis among relatives who moved from Brazil with an infected cat, although the species was not identified [7]. In 2022 the first feline *S. brasiliensis* infections in Paraguay were reported [8]. Here we report the first autochthonous human case of cat-transmitted sporotrichosis by *S. brasiliensis* in Paraguay.

2.3 Case report

This study was approved by the Committee for Ethics in Research of the Federal University of Parana (number—CAAE 52726021.8.0000.0102). Also, the

patient assigned a consent form to publication of information about her and the study adheres to the Declaration of Helsinki.

On March 9, 2024 (T = 0d), a 39-year-old female resident of Presidente Franco, Paraguay, without known underlying diseases or immunosuppression, noticed a skin lesion on her right arm that initially looked like a mosquito bite, which progressed to an inflamed nodule. On April 2 (T = 24d), after consulting a medical center, she began treatment with cephalexin (oral 1 gram every 8 h for seven days) (Figure 1A, 1B). On April 9 (T = 31d), the skin lesion progressed, and reddish areas appeared that followed the lymphatic pathway, which was initially diagnosed as cellulitis, for which clindamycin was administered (oral 500 mg every 8 h for 10 days). On April 30 (T = 52d), trimethoprim-sulfamethoxazole (oral 160 mg/800 mg every 12 h for 10 days) was prescribed for insufficient clinical response to clindamycin. Finally, on May 3 (T = 55d) a cycle of vancomycin was started (1 gram every 24 h for 10 days). At this point, the skin lesions evolved to 13 swollen and painful nodules (Fig. 1C). All these treatments were accompanied by analgesics and anti-inflammatory drugs, except for no corticosteroids. Despite the use of antibiotics, no bacterial culture was performed.



Figure 1. Clinical image across different time points. First consult on April 2 (Figure 1A, 1B), evolution to 13 swollen and painful nodules on May 3 (Figure 1C), consult with infectious disease specialist on May 11, (Figure 1D), itraconazole treatment for 6 days on May 16 (Figure 1E), for 18 days on May 28 (Figure 1F), for 24 days on June

41

3 (Figure 1G), for 50 days on June 29 (Figure 1H), for 59 days on July 8 (Figure 1I), for 89 days on August 7 (Figure 1J), and for 112 days on August 30 (Figure 1K).

On May 11 (T = 63), the patient consulted an infectious disease specialist that examined the skin lesions (Fig. 1D). During the consult, the patient reported to be scratched by her cat and after taking note of the clinical history of cat (lesion on head and cytopathological examination with observation of yeast-like structures, suggestive for *Sporothrix*, in the exudate of the cat's skin lesion), the specialist immediately requested a culture. Secretion from the skin lesions was collected using a swab and smeared onto Sabouraud Dextrose Agar medium. On the same day, treatment with oral itraconazole (200 mg daily) was started. In time the patient subsequently demonstrated clear clinical improvement (Fig. 1E-K) and continued with the treatment to date (January 8 [T = 305d], 2025).

In March 8, 2024, almost simultaneously with the patient clinical signs onset, her cat was also appeared with a lesion on its head. Antibiotic treatment was started. When no improvement was observed, the cat was admitted to a veterinary clinic on March 20 and continued with other antibiotics. At this point the cat was already weakened by the disease. Samples of the cat were collected for cytology and on April 17, the result revealed the presence of cigar-shaped yeast structures, suggestive for *Sporothrix*. No culture was performed, and despite starting itraconazole treatment (oral, 50 mg daily) at the same day (almost two months of the beginning of the lesion), few days later the cat was euthanized due to the declining medical condition.

The culture obtained on May 11 originated from a needle aspirate from the largest nodule containing a purulent secretion, which was sent to the microbiological laboratory. On direct examination using microscopy, an abundant inflammatory reaction was observed. Gram staining showed no bacterial or fungal structures. Culture was performed on SDA at 27 °C. After five days, the development of flat, white colonies with short aerial mycelia was observed that melanized over time. Under the microscope, hyphae with spherical conidia located in the apical part of the conidiophore were identified, in a characteristic teardrop or daisy flower arrangement, typical of the *Sporothrix* genus.

Calmodulin (CaM) sequencing was performed as described earlier [9] and the isolate was identified as *S. brasiliensis*. The sequence was deposited to the NCBI

Genbank database under accession number PQ550639. Using short tandem repeat (STR) genotyping as described previously [10,11], the genotype of the isolate appeared identical to previously genotyped isolates from the South of Brazil (Fig. 2).

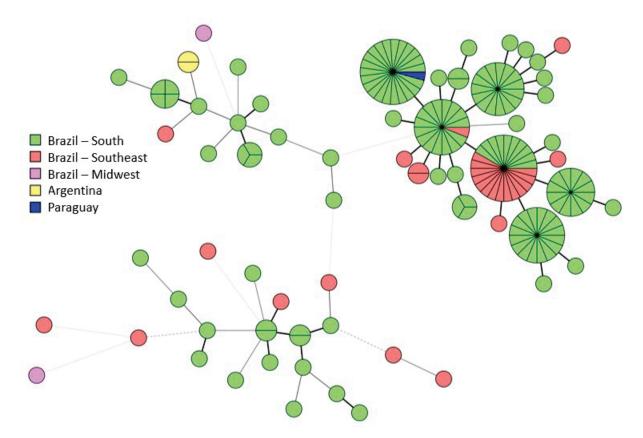


Figure 2. Minimum-spanning tree of 176 *Sporothrix brasiliensis* isolates, including one Paraguayan isolate from the current study. Branch lengths correlate with the similarity between isolates, which is further indicated by thick solid lines (variation in one allele), thin solid lines (variation in two alleles), thin dashed lines (variation in three alleles) and thin dotted lines (variation in four or more alleles). Isolates are colored according to different locations of origin.

Antifungal susceptibility testing was also performed by broth microdilution as outlined in the Clinical & Laboratory Standards Institute (CLSI) reference standard M38 for the mycelial form [12] and CLSI reference standard M27 for the yeasts form [13]. For the mycelial phase, isolate was cultured onto Potato Dextrose Agar (Difco Laboratories, Detroit, USA) plates at 30 °C for 7 days, which was confirmed microscopically. For the yeast phase, isolate was cultured onto Brain Heart Infusion (BHI) plates (Xebios Diagnostics GmbH, Düsseldorf, Germany), at 35 °C for 7 days,

followed by a second passage at BHI at 35 °C for 7 days, yeast presence was confirmed microscopically. *Aspergillus flavus* ATCC 204304 was used as quality control for the mycelial phase and *Pichia kudriavzevii* ATCC 6258 and *Candida parapsilosis* ATCC 22019 for the yeast phase. For both morphological phases, tentative epidemiological cut-off values (ECVs) for amphotericin B, itraconazole, voriconazole, posaconazole and terbinafine, were implemented [14]. All minimum inhibitory concentrations (MICs) were below the ECVs and for the other antifungals no elevated MIC values were found (Table 1).

Table 1. Minimum inhibitory concentrations (MICs) of the Paraguayan *Sporothrix brasiliensis* isolate in both yeast and mycelial phase according to clinical laboratory and standards institute (CLSI) guidelines. Concentrations in µg/mL.

AFG MFG Morphology AMB FLU ITC VOR POS ISA TRB 0.25 Yeast 32 0.063 0.25 0.063 0.125 0.125 0.125 0.125 ≤0.008* Mycelial 2 64 0.25 16 0.25 4 0.25 ≤0.008* AMB, amphotericin B; FLU, fluconazole; ITC, itraconazole; VOR, voriconazole; POS, posaconazole; ISA, isavuconazole; TER, terbinafine; AFG, anidulafungin; MFG, micafungin.

2.4 Discussion

While the first human sporotrichosis cases in Paraguay occurred in 2017, involving an imported cat, this case describes the first autochthonous case. Recently, feline cases in the same region were reported, known for their propensity to cause outbreaks [8]. Most sporotrichosis human cases present cutaneous or lymphocutaneous clinical forms, affecting lower and upper members and face [15]. Usually, a papular lesion starts on an inoculating site after some days or weeks, and may progress to nodular or ulcerative form. Lesions are characterized by swelling and itching erythematous halos [11]. In the lymphocutaneous form, lesions progress in a sporotrichoid pattern, following lymphatic vessels and reaching lymph nodes. The clinical signs of the current case are in line with this.

Despite noticing the clinical signs early and having an affected cat, it took a prolonged time to accurately diagnose human sporotrichosis and start antifungal treatment, while even antibiotics were administered initially. This highlights the need to raise awareness among clinicians and veterinarians for this disease, which is characterized by its typical sporotrichoid spread, and underscores the importance of a One Health approach and rational antifungal usage. Especially in the early stage of the infection, when itraconazole is administered accordingly, the treatment time is considerably lower than with more severe cases, as was shown here. Nonetheless, the isolate was found to have in vitro wildtype MICs to itraconazole, which was in line with the favorable clinical response to systemic itraconazole treatment.

STR genotyping demonstrated that this isolate is genetically closely related to isolates from Southern Brazil, suggesting an introduction by sick cats from there, although whole genome sequencing would be needed for definitive confirmation.

Our findings once more support the necessity of implementing an One Health approach to improve surveillance, diagnosis, and treatment of sporotrichosis, considering the importance of early diagnosis and treatment for an optimal prognosis.

Declaration of competing interest

EFJM received research grants from Mundipharma and Scynexis, is in the scientific advisory board for Pfizer and has received speaker fees from Gilead Sciences. All other authors declare no conflict of interest.

Funding

This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Finance Code 001. Was also supported by the Canisius-Wilhelmina Hospital (Grant CWZ_001421). Received fellowships from CNPq (grant number 312811/2018–7), Brasilia, Brazil. The work was supported by the Araucaria Foundation (http://www.fappr.pr.gov.br/, accessed on 31 June 2021) (NAPI grant number 113/2020).

2.5 References

[1] Rossow JA, Queiroz-Telles F, Caceres DH, Beer KD, Jackson BR, Pereira JG, Ferreira Gremião ID, Pereira SA. A One Health Approach to Combatting *Sporothrix* *brasiliensis*: Narrative Review of an Emerging Zoonotic Fungal Pathogen in South America. J Fungi (Basel) 2020:6(4):247.

[2] Schechtman RC, Falcão EMM, Carard M, García MSC, Mercado DS, Hay RJ.
Sporotrichosis: hyperendemic by zoonotic transmission, with atypical presentations, hypersensitivity reactions and greater severity. An Bras Dermatol 2022:97(1):1–13.
[3] de Andrade Galliano Daros Bastos F, Raimundo Cognialli RC, Rodrigues de Farias M, Dos Santos Monti F, Wu K, Queiroz-Telles F. Spread of *Sporothrix* spp. through respiratory droplets from infected cats: A potential route of transmission. Med Mycol 2022:60(11):myac079.

[4] Nahal J, Coelho RA, Almeida-Silva F, Bernardes-Engemann AR, Procópio-Azevedo AC, Rabello VBS, Loureiro RG, Freitas DFS, do Valle ACF, de Macedo PM, Oliveira MME, Silva MBTD, Zancopé-Oliveira RM, Almeida-Paes R, Gutierrez-Galhardo MC, Figueiredo-Carvalho MHG. Non-Zoonotic Transmission of Sporotrichosis: A Translational Study of Forty-Three Cases in a Zoonotic Hyperendemic Area. J Fungi (Basel) 2024:10(9):610.

[5] Etchecopaz A, Toscanini MA, Gisbert A, Mas J, Scarpa M, Iovannitti CA, Bendezú K, Nusblat AD, Iachini R, Cuestas ML. *Sporothrix brasiliensis*: A Review of an Emerging South American Fungal Pathogen, Its Related Disease, Presentation and Spread in Argentina. J Fungi (Basel) 2021:7(3):170.

[6] Rodrigues AM, Cruz Choappa R, Fernandes GF, de Hoog GS, de Camargo ZP. *Sporothrix chilensis* sp. nov. (Ascomycota: Ophiostomatales), a soil-borne agent of human sporotrichosis with mild-pathogenic potential to mammals. Fungal Biol 2016:120(2):246–64.

[7] García Duarte JM, Wattiez Acosta VR, Fornerón Viera PML, Aldama Caballero A, Gorostiaga Matiauda, GA, Rivelli de Oddone VB, Pereira Brunelli JG. Esporotricosis trasmitida por gato doméstico. Reporte de un caso familiar. Rev. Nac. (Itauguá) 2017:9(2): 67-76.

[8] do Prado CM, Razzolini E, Santacruz G, Ojeda L, Geraldo MR, Segovia N, Pereira Brunelli J, Vicente VA, Svoboda WK, Queiroz-Telles F. First Cases of Feline Sporotrichosis Caused by *Sporothrix brasiliensis* in Paraguay. J Fungi (Basel) 2023:9(10):972.

[9] Bombassaro A, Spruijtenburg B, Medeiros F, Jacomel Favoreto de Souza Lima B, Ballardin LB, Farias MR, Vicente VA, de Queiroz-Telles F, Meis JF, de Groot T. Genotyping and antifungal susceptibility testing of *Sporothrix brasiliensis* isolates from Southern Brazil. Mycoses 2023:66(7):585-593.

[10] Spruijtenburg B, Bombassaro A, Meijer EFJ, Rodrigues AM, Grisolia ME, Vicente VA, de Queiroz-Telles F, Meis JF, de Groot T. *Sporothrix brasiliensis* genotyping reveals numerous independent zoonotic introductions in Brazil. J Infect 2023:86(6):610-613.

[11] Fernandez NB, Spruijtenburg B, Tiraboschi IN, Meis JF, Lugo A, López Joffre MC, Meijer EFJ. Genotyping and clonal origin of *Sporothrix brasiliensis* in human sporotrichosis cases in Argentina. Med Mycol Case Rep 2024:43:100633.

[12] Clinical and Laboratory Standards Institute (CLSI). Reference method for broth dilution antifungal susceptibility testing of filamentous fungi. 3rd ed. Pennsylvania: CLSI standard M38, 2017.

[13] Clinical and Laboratory Standards Institute (CLSI). Reference method for broth dilution antifungal susceptibility testing of yeasts. 4th. Pennsylvania: CLSI standard M38, 2017.

[14] Espinel-Ingroff A, Abreu DPB, Almeida-Paes R, Brilhante RSN, Chakrabarti A, Chowdhary A, Hagen F, Córdoba S, Gonzalez GM, Govender NP, Guarro J, Johnson EM, Kidd SE, Pereira SA, Rodrigues AM, Rozental S, Szeszs MW, Ballesté Alaniz R, Bonifaz A, Bonfietti LX, Borba-Santos LP, Capilla J, Colombo AL, Dolande M, Isla MG, Melhem MSC, Mesa-Arango AC, Oliveira MME, Panizo MM, Pires de Camargo Z, Zancope-Oliveira RM, Meis JF, Turnidge J. Multicenter, International Study of MIC/MEC Distributions for Definition of Epidemiological Cutoff Values for *Sporothrix* Species Identified by Molecular Methods. Antimicrob Agents Chemother 2017:61(10):e01057-17.

[15] Queiroz-Telles F, Bonifaz A, Rossow J, Chindamporn A. *Sporothrix* and Sporotrichosis. In: Rezaei N, editor. Encyclopedia of Infection and Immunity. Elsevier; 2022. p. 376–96.

3 CHAPTER 3 - Emergence of feline sporotrichosis in Puerto Iguazú (Argentina) near the Argentina-Brazil Border

Authors: Katherina Alicia Vizcaychipi; María Cecilia López-Joffre; Mónica Martínez; Jerson Andrés Cuéllar-Sáenz; Marina Ramos, Celeste Agüero, Natalia Olsina, Emanuel Grassi, Esteban Couto; Jorge Mendoza; Carolina Melchior do Prado; Jorge Pablo Castillo; Mabel D. Giménez; Karen E. DeMatteo; Álvaro A. Faccini-Martínez; Mariana Viale; Adriana Toranzo; Cristina Elena Canteros.

Author affiliations: Universidad del Salvador, Virasoro, Corrientes, Argentina (K.A.Vizcaychipi, J. Mendoza, J.P. Castillo); Instituto Misionero de Biodiversidad, Puerto Iguazú, Misiones, Argentina (K.A. Vizcaychipi, M. Martínez, E. Grassi); Instituto Nacional de Medicina Tropical ANLIS - "Carlos G. Malbrán", Puerto Iguazú, Misiones, Argentina (K.A. Vizcaychipi, E. Couto); Instituto Nacional de Enfermedades Infecciosas ANLIS - "Carlos G. Malbrán", Buenos Aires, Argentina (M.C. López19 Joffré, M. Viale, A. Toranzo, C.E. Canteros); Universidad Nacional de Colombia, Bogotá, Colombia (J.A. Cuéllar-Sáenz); Dirección Municipal de Zoonosis, Puerto Iguazú, Misiones, Argentina (M. Ramos, C. Agüero); Clínica veterinaria, Puerto Iguazú, Misiones, Argentina (N. Olsina); Universidad Federal de Paraná, Brasil (C. Melchior do Prado); Instituto de Genética Humana de Misiones, CONICET, Posadas, Misiones, Argentina; Universidad Nacional de Misiones, Posadas, Argentina (M.D. Giménez); Washington University in St Louis & WildCare Institute at the Saint Louis Zoo, Missouri, USA (K.E. DeMatteo); Hospital Militar Central, Bogotá, Colombia; Universidad Militar Nueva Granada, Bogotá, Colombia (Á.A. Faccini-Martínez).

Correspondence:

Katherina A. Vizcaychipi. Puerto Iguazú, Misiones, Argentina. Email: kvizcaychipi@gmail.com

Emerging Infectious Diseases

Received 08 December 2024, Revised 27 January 2025, Accepted 21 February 2025.

3.1 Abstract

We describe a large urban outbreak of feline sporotrichosis caused by *Sporothrix brasiliensis* in Argentina. Over a 7-month period in Puerto Iguazú, which borders Brazil, culture-proven sporotrichosis was identified in 9 cases across 7 households. Notably, 2 of these cases presented without cutaneous lesions, and 2 households experienced multiple cases, indicating potential intradomestic transmission.

3.2 Text

Sporotrichosis is an implantation mycosis caused by thermal-dimorphic fungi belonging to the *Sporothrix schenckii* complex (1). Among the pathogenic species, *Sporothrix brasiliensis* stands out for its high virulence, epidemic potential, and a zoonotic/enzootic transmission which occurring through bites, scratches, or contact with exudates from infected animals, particularly domestic cats (2-4).

In South America, *S. brasiliensis* was first identified in Brazil, and has since been reported in other Latin American countries (1-8). Over recent decades, sporotrichosis in Brazil has seen a significant epidemiological shift, marked by intense, widespread urban zoonotic outbreaks, initially concentrated in Rio de Janeiro, affecting cats, dogs, and humans (2, 9). These outbreaks have spread to several cities in the southern and southeastern states, including Foz do Iguaçu, located on the triple border between Argentina and Paraguay (2, 3, 5, 7, 10), and recently, a cat infection by *S. brasiliensis* was reported in Ciudad del Este, the Paraguayan side of the triple border (8). In Argentina, the first human isolation of *S. brasiliensis* was documented in 1986 in the south of province of Misiones, with no identified source of infection (4). Since then, zoonotic sporotrichosis cases have increased, with most occurrences reported in the central and southern regions of the country (1, 4).

This study reports the emergence of urban transmission of feline sporotrichosis in Puerto Iguazú, Misiones, Argentina (25°36'39"S, 54°34'49"W). Located in the extreme northeast of Argentina, on the border of the Triple Frontier, the city has a population of approximately 54,675 and is 1,278 km from Buenos Aires City. Puerto Iguazú is a major tourist destination due to the Iguazu Falls, and is

characterized by significant cross-border dynamics, including high population and commercial movement.

Between August 2023 and February 2024, we conducted an intensified passive surveillance on 21 domestic cats (*Felis catus*) from 12 households in Puerto Iguazú's urban area (Figure 1).

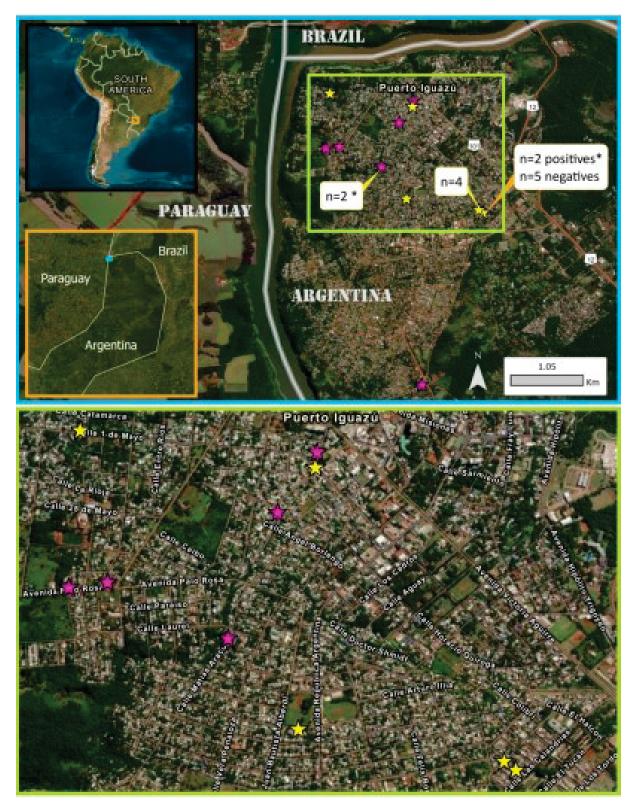


Figure 1. Map of Puerto Iguazú, Misiones, Argentina, located at the tri-border with Brazil and Paraguay, showing the spatial distribution of positive cases of feline sporotrichosis confirmed (n=9; pink) and ruled out cases (n=12; yellow) in the different neighborhoods studied. For those areas where multiple cases (positive and negative) overlap in location, the number of individual cases is indicated. The cases

of where intra-household transmission occurred between two cats (FSCMi24 - FSCMi25 and FSCMi41 – FSCMi43) are indicated by an *. In the spatial analysis, the distance between households with feline sporotrichosis cases was measured using nearest-neighbor calculation in ArcGIS Pro, which calculates the shortest distance between two points.

Cats with lesions consistent with feline sporotrichosis (ulcers, scabs, soft nodules, and ulcerated subcutaneous nodules with exudate) were included, as well as those without lesions but in contact with affected cats or living in areas with documented cases. None had received treatment before sample collection. Pet owners provided written consent, and clinical, epidemiological, and demographic data were recorded. All suspected cases and their contact were evaluated by veterinarians, with ongoing prevention and awareness campaigns directed at owners and the community.

Nasal and lesion swabs were collected from cats with skin lesions, and nasal swabs from cats without lesions. Diagnosis and species identification of *Sporothrix* were conducted using phenotypic (Giemsa stain and culture) and genotypic methods (sequencing ITS region and partial sequencing of the calmodulin gene) (1, 4, 10).

Among the 21 cases studied, 9 (42.9%) were suspected cases, while 12 (57.1%) were contacts. Feline sporotrichosis was confirmed by culture in 77.8% (7/9) of the suspected cases and 16.7% (2/12) of the contacts (Appendix Figure 1); of them, molecular confirmation of *S. brasiliensis* was achieved in 77.8% (7/9) and two samples were inconclusive due to mold and bacterial contamination, which hindered identification of *Sporothrix* species (Table 1).

Table 1. Features of domestic cats (*Felis catus*) suspected of feline sporotrichosis and their contacts, from 12 households in various neighborhoods of Puerto Iguazú's urban area, Misiones, Argentina; August 2023 to February 2024.

Household no. /Cat ID	Date	Sex/Age	Neutered	Inhabit*	Type/Location of lesion	Collected material	DE/Culture	Identification / DMic no.
1/FSCMi-0111	1-Aug-23	ි/Adult	No	Feral	Multiple dog bites	NS/LS/C	N/N	
2/FSCMI-017	13-Nov-23	∂/Adult	yes	Outdoor	NI	NS/C	N/N	
2/FSCMI-024	24-Nov-23	ੈ/Adult	Yes	Outdoor	NI	NS	ND/Sporothrix sp.	S. brasiliensis/247479
2/FSCMI-025*	13-Nov-23	∂/Adult	Yes	Outdoor	NI	NS	N/Sporothrix sp.	S. brasiliensis/247478
2/FSCMI-019	13-Nov-23	₽/Adult	Yes	Outdoor	NI	NS	N/N	
3/FSCMI-021	16-Nov-23	₽/Adult	No	Indoor	Localized/Head	NS/LS	yeast/Sporothrix sp.	NC
4/FSCMI-023	24-Nov-23	්/Adult	Yes	Outdoor	Multiple/Head, dorsum, and extremities	NS/LS	ND/Sporothrix sp.	S. brasiliensis/247481
5/FSCMI-026	24-Nov-23	ੈ/Adult	Yes	Feral	Localized/Head	NS/LS	ND/Sporothrix sp.	S. brasiliensis/247480
6/FSCMI-031	20-Dec-23	♂/Kitten	No	Outdoor	NI	NS	N/N	
6/FSCMI-032	20-Dec-23	⊊/Juvenile	No	Outdoor	NI	NS	N/N	
6/FSCMI-033	20-Dec-23	♂/Kitten	No	Outdoor	NI	NS	N/N	
7/FSCMI-034	20-Dec-23	ੈ/Adult	Yes	Outdoor	NI	NS	N/N	
7/FSCMI-035	20-Dec-23	∂/Adult	Yes	Outdoor	NI	NS	N/N	
7/FSCMI-036	20-Dec-23	∂/Adult	Yes	Outdoor	NI	NS	N/N	
7/FSCMI-037	20-Dec-23	♀/Adult	Yes	Outdoor	NI	NS	N/N	
8/FSCMI-038	20-Dec-23	⊊/Juvenile	Yes	Outdoor	N	NS	N/N	
9/FSCMI-040	10-Jan-24	♀/Juvenile	No	Feral	Multiple/Head and extremities	NS/LS	yeast/Sporothrix sp.	NC
10/FSCMi-041	15-Jan-24	ੈ/Adult	Yes	Outdoor	Multiple/Head, dorsum, and extremities	NS/LS	yeast/Sporothrix sp.	S. brasiliensis/247599
10/FSCMi-043	22-Jan-24	ੈ/Adult	Yes	Outdoor	Multiple/Head and extremities	NS/LS	yeast/Sporothrix sp.	S. brasiliensis/247600
11/FSCMi-0421	15-Jan-24	⊊/Juvenile	No	Outdoor	Alopecia	NS/LS	ND/Microsporum canis	
12/FSCMi-049	28-Feb-24	<i>∛</i> /Senior	Yes	Outdoor	Multiple/Head, dorsum, and extremities	NS/LS	yeast/Sporothrix sp.	S. brasiliensis/247735

* The cat had frequent sneezing. ♂ Male, ♀ Female. ¥ Inhabit; Indoor: Cats kept exclusively indoors or in enclosed spaces, without outdoor access; Outdoor: Cats with free access to roam outdoors, including streets, neighboring properties, or vacant lots; Feral: Cats living in the wild with minimal or no human interaction. NI: No injuries. NS: Nasal Swab. LS: Lesion Swab. C: Claw. N: Negative. ND: Not Determined. DE: Direct examination of the sample, with Giemsa staining. NC: Inconclusive due to mold and bacterial contamination. ¶Dog bites and diagnosis of dermatophytosis.

Among the two confirmed cases in contacts, sneezing was identified as the sole symptom in one case, and two suspected cases were ruled out due to differential diagnosis (dog bite and dermatophytosis) (Table 1). All cases have been reported to health authorities.

Feline sporotrichosis cases were identified in 58,3% (7/12) of the households. In 2 households, multiple cases were detected, suggesting intradomestic transmission. However, in 5 households, only one cat with sporotrichosis was found. Most of the cats had free access to streets, neighboring properties, and vacant lots. The average distance between the nearest households with feline sporotrichosis cases was 1.84 km ± 1.22 km (range: 0–4.54 km) (Table 1, Figure 1).

Our results describe the largest outbreak the S. brasiliensis in Argentina to date. We

identified 9 proven feline sporotrichosis in 7 months in Puerto Iguazú, province of Misiones, being higher than that reported for the whole country (1). The outbreak reflects transmission dynamics similar to the Brazilian epidemic, with evidence of multiple foci of transmission and asymptomatic carriers that facilitate the spread of the fungus (2, 5, 7, 10). In addition, the nearest epidemic focus is on the Brazilian side of the triple border (7), and to our knowledge, the latest cases of feline sporotrichosis in Argentina have been reported in Buenos Aires and Santa Cruz (1).

Asymptomatic carriers hinder control efforts by delaying diagnosis and treatment. Screening all contacts of confirmed cases is essential to minimize the risk of transmission (1, 10). Addressing these challenges requires mandatory case reporting and public health measures. The expansion of this emerging disease outside Brazil in contiguous countries calls for coordinated cross-border One Health actions and context-specific interventions, which will be crucial to safeguard local communities and tourists (1,5). Further studies are needed to understand its transmission dynamics.

Ethical Considerations

This study adhered to established ethical standards for veterinary research, ensuring the welfare and humane treatment of all animals involved. Sample collection was performed by qualified veterinarians using minimally invasive methods to reduce discomfort and stress. Procedures followed veterinary best practices and complied with international guidelines. Informed consent was obtained from all pet owners, who were briefed on the study objectives, methods, and potential benefits. The study protocol was reviewed and approved by the Institutional Ethics Committee of the Faculty of Agricultural and Veterinary Sciences, Universidad del Salvador (CICUAE - FCAyV, USAL-06-2021), with permits for the collection of natural resources and/or genetic material granted by the Institutio Misionero de Biodiversidad (IMiBio) and the Ministerio de Ecología y Recursos Naturales (Expte. 9950- 70-2023-1). The strains were deposited in the culture collection of the Mycology Department at INEI-ANLIS (DMic), Buenos Aires, and in the Biobank of the Instituto Misionero de Biodiversidad, Puerto Iguazú, Misiones.

Acknowledgment

The data presented are part of a broader ongoing project, Proyecto SIGEVA-USAL 2022-2025, entitled Estudio eco-epidemiológico y sanitario de *Sporothrix brasiliensis* en localidades correntinas y misioneras del Corredor Jesuítico Guaraní Argentino. We extend our gratitude for the support and collaboration of the Ministerio de Salud Pública (Misiones) and IMiBio, as well as trust and cooperation of the Consejo de Veterinarios de Misiones, Zoonosis Iguazú, private veterinary clinics in Puerto Iguazú, the Sociedad Rural de Ganado Menor (Misiones), and all pet owners involved in the study.

3.3 References

1. Etchecopaz A, Toscanini MA, Gisbert A, Mas J, Scarpa M, Iovannitti CA, et al. *Sporothrix brasiliensis*: a review of an emerging South American fungal pathogen, its related disease, presentation and spread in Argentina. J Fungi (Basel). 2021; 7:170. https://doi.org/10.3390/jof7030170

 Rodrigues AM, Hoog GS de, Camargo ZP de. *Sporothrix* species causing outbreaks in animals and humans driven by animal–animal transmission. PLoS Pathog. 2016;12:e1005638. https://doi.org/10.1371/journal.ppat.1005638
 Rabello VBS, Almeida MA, Bernardes-Engemann AR, Almeida-Paes R, Macedo PM de, Zancopé-Oliveira RM. The historical burden of Sporotrichosis in Brazil: a systematic review of cases reported from 1907 to 2020. Braz J Microbiol. 2022; 53:231. https://doi.org/10.1007/s42770-021-00658-1

4. Córdoba S, Isla G, Szusz W, Vivot W, Hevia A, Davel G, et al. Molecular identification and susceptibility profile of *Sporothrix schenckii* sensu lato isolated in Argentina. Mycoses. 2018; 61:441–8. https://doi.org/10.1111/myc.12760

5. Rossow JA, Queiroz-Telles F, Caceres DH, Beer KD, Jackson BR, Pereira JG, et al. A One Health approach to combatting *Sporothrix brasiliensis*: narrative review of an emerging zoonotic fungal pathogen in South America. J Fungi (Basel). 2020; 6:247. https://doi.org/10.3390/jof6040247

6. Thomson P, González C, Blank O, Ramírez V, Río C del, Santibáñez S, et al. Sporotrichosis outbreak due to *Sporothrix brasiliensis* in domestic cats in Magallanes, Chile: a One-Health-approach study. J Fungi (Basel). 2023; 9:226. https://doi.org/10.3390/jof9020226

7. Prado C, Chiyo L, Santi C, Cognialli R, Reis G, Geraldo M, et al. P464 Feline sporotrichosis: an emerging disease in the Brazilian side of the Southern Triple Border. Med Mycol. 2022; 60(Suppl 1): myac072P464.

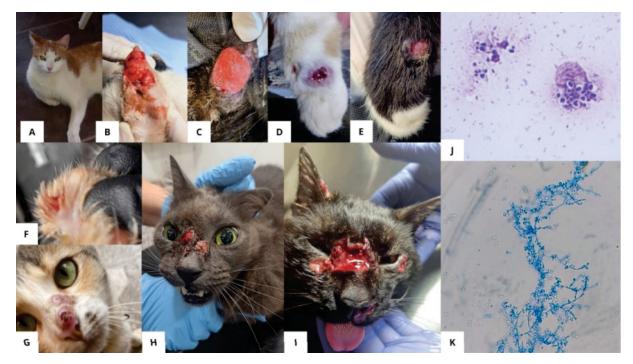
https://doi:10.1093/mmy/myac072.P464

8. do Prado CM, Razzolini E, Santacruz G, Ojeda L, Geraldo MR, Segovia N, et al. First cases of feline Sporotrichosis caused by *Sporothrix brasiliensis* in Paraguay. J Fungi (Basel). 2023; 9:972. https://doi.org/10.3390/jof9100972

9. Santos AR dos, Misas E, Min B, Le N, Bagal UR, Parnell LA, et al. Emergence of zoonotic Sporotrichosis in Brazil: a genomic epidemiology study. Lancet Microbe. 2024;5:e282–90. https://doi.org/10.1016/s2666-5247(23)00364-6

10. Cognialli RCR, Cáceres DH, Bastos FAGD, Cavassin FB, Lustosa BPR, Vicente VA, et al. Rising incidence of *Sporothrix brasiliensis* infections, Curitiba, Brazil, 2011-2022. Emerg Infect Dis. 2023; 29:1330-9. https://doi.org/10.3201/eid2907.230155

Appendix



Appendix Figure 1. Feline sporotrichosis and phenotypic studies. A. Cat without lesions, with frequent sneezing (FSCMi25). B - C. Multiple ulcerative lesions in the ear (FSCMi021 - FSCMi026). D - E. Ulcerative lesion on forelimbs (FSCMi043 -

FSCMi049). F. Ulcerative lesion around the claw (FSCMi040). G. Crusty ulcerative lesions on the nose (FSCMi043). H - I. Multiple crusty ulcerative lesions on the head (FSCMi041 - FSCMi026). J. Cigar-shaped oval capsulated yeasts (Giemsa stain. 1000x). K. Micromorphology of *Sporothrix* sp. Colonies (lactophenol cotton blue. 400x).

4 CHAPTER 4 - Absence of initial elevated itraconazole minimum inhibitory concentrations in *Sporothrix brasiliensis* from felines with treatment failure

Authors: Carolina Melchior do Prado*, Bram Spruijtenburg*, Emanuel Razzolini, Luciana Chiyo, Carlos Santi, Caroline Amaral Martins, Gabriela Santacruz, Nancy Segovia, José Pereira Brunelli, Regielly Caroline Raimundo Cognialli, Jacques F. Meis, Vânia Aparecida Vicente, Theun de Groot, Eelco F.J. Meijer*, Flávio Queiroz-Telles*

Author affiliations: Federal University of Paraná, Curitiba, Brazil (C.M. do Prado, E. Razzolini, R.C.R. Cognialli, V.A. Vicente, F. Queiroz-Telles); Radboudumc-CWZ Center of Expertise for Mycology, Nijmegen, The Netherlands (C.M do Prado, B. Spruijtenburg, J.F. Meis, T. de Groot, E.F.J. Meijer); Radboudumc, Nijmegen, The Netherlands (B. Spruijtenburg, J.F. Meis, E.F.J. Meijer), Canisius-Wilhelmina Hospital (CWZ)/Dicoon, Nijmegen, The Netherlands (B. Spruijtenburg, T. de Groot, E.F.J. Meijer); Zoonosis Control Center, Foz do Iguaçu, Brazil (L. Chiyo, C. Santi, C.A. Martins); National University of the East, Minga Guazú, Paraguay (G. Santacruz, N. Segovia); Ministry of Public Health and Social Welfare, Asuncion, Paraguay (J.P. Brunelli); University of Cologne, Cologne, Germany (J.F. Meis).

Address for Correspondence: Flávio Queiroz-Telles, Department of Public Health, Hospital de Clínicas, Federal University of Paraná, Curitiba, Brazil. Fone: +55 (41) 99972-1828. Email: queiroz.telles@uol.com.br. https://orcid.org/0000-0001-7034-2418.

4.1 Abstract

Cat-transmitted sporotrichosis caused by *Sporothrix brasiliensis* is an emerging zoonosis in Latin America. As treatment of feline sporotrichosis is often not effective, we determined whether this is caused by infection of *S. brasiliensis* strains with preexisting elevated itraconazole MICs, which is the drug of choice. At the triple border region of Brazil, Paraguay and Argentina from 2021 to 2023, 108 *S. brasiliensis* strains were isolated from felines before antifungal treatment. Antifungal susceptibility testing (AFST) to nine antifungal compounds was evaluated for the mycelial and yeast phase. The main clinical presentation was cutaneous disseminated sporotrichosis (61%) and was the only form showing sporotrichosis-induced mortality (61%). AFST demonstrated only non-wild type MICs for terbinafine (n = 9). While MIC levels were low for most antifungals, they were higher in the mycelial phase. We conclude that the varying clinical presentations with large differences in mortality were not caused by pre-existing elevated itraconazole MICs.

4.2 Text

Sporotrichosis is a globally neglected epizoonotic and sapronotic disease, mostly involving infection of the skin and subcutaneous tissues, caused by fungi of the Sporothrix genus and the most prevalent implantation mycosis (1). Sporothrix species are thermally dimorphic fungi from the order Ophiostomatales, showing filamentous forms at 25-30°C and yeast-like forms at temperatures of 35-37°C as in mammals (2). The main clinical pathogenic species include S. brasiliensis, S. schenckii, S. globosa and S. luriei. S. schenckii and S. globosa are usually transmitted via the sapronotic route, involving traumatic implantation with plant/soil debris (3). Remarkably, in the last 30 years transmission of S. brasiliensis to humans, other felines and canines through infected cats by the zoonotic route has led to various outbreaks in Brazil and other Latin American countries (4-7). Currently cattransmitted sporotrichosis (CTS) by S. brasiliensis is a major public health concern in Latin America, since infections are rapidly spreading from Brazil to other countries and cases were described in Brazil (8,9), Argentina (5,10), Paraguay (4), and Chile (6). In addition, imported cases in United Kingdom (11) and United States (12) were recently reported. Transmission by infected cats, via yeast form (13), occurs through bites, scratches, direct contact with exudate from skin lesions and probably through respiratory droplets, via cat sneezing (2,14). Cats are the primary animal hosts and main source of infection for other cats, dogs and humans (15).

Outbreaks often involve clonal zoonotic transmission (8,16). To curb these outbreaks, one of the necessary measures is antifungal treatment of cats (15). The drug of choice is itraconazole, although various refractory cases have been reported (17,18). Whether treatment failure is due to high antifungal MICs is unknown as this is poorly investigated (19). Recently high antifungal MICs against itraconazole were reported in isolates obtained from both cats and humans (19–21). It is not clear

whether strains with reduced susceptibility are also transmitted or whether this only develops during treatment. Antifungal susceptibility testing, applicable to both the yeast and mycelial form, has not been standardized leading to different protocols. As a consequence, published MICs are currently difficult to compare. In the present study, we investigate the spread of feline sporotrichosis in the triple border region between Brazil, Paraguay and Argentina by molecular genotyping. Additionally, MICs of common antifungals using microbroth dilution methods of both the yeast and mycelial phase, were obtained, to determine whether cats with sporotrichosis had S. *brasiliensis* with elevated itraconazole MICs at the onset of treatment.

4.3 Materials and methods

4.3.1 Isolate and data collection

Between July 2021 to October 2023, swab samples were collected from 108 symptomatic cats presenting with lesions compatible with sporotrichosis. Cats were selected through notification of the owners, health agents of the Zoonosis Control Center of Foz do Iguaçu (FIG) (CCZ – FIG, Brazil), veterinarians from private clinics and hospitals, and the receipt of suspected animals by CCZ. In FIG, the service was provided at the homes of the cats or at the place of indication by the citizen of stray cats. In Paraguay the service was provided at private veterinary clinics. All cats lived in the triple border region between Brazil, Paraguay and Argentina. Clinical and environmental data were obtained with questionnaires send to the owners of each cat. All cats were evaluated and classified according to the types of the lesions and divided in 3 groups: 1) cutaneous disseminated; 2) fixed cutaneous and 3) extracutaneous. Cartographic bases from the Brazilian Institute of Geography and Statistics (IBGE) were used for georeferencing the coordinates. Quantus Gis software (QGIS) was used to assemble the maps. To determine the clinical outcome, felines were followed for the complete duration of treatment, or until they passed away or were lost to follow-up.

4.3.2 Diagnosis

Sporotrichosis was diagnosed via fungal culture as previous described (4). Swabs collected from the wounds were sent for fungal culture. Specimens were cultivated on Sabouraud dextrose agar (SDA, KASVI, Pinhais, Brazil) containing chloramphenicol and incubated at 25-27°C for up to 10 days. Micromorphology of colonies was performed and *Sporothrix* growth was confirmed. Calmodulin (CaM) sequencing was performed for species identification as previous described (4). As control isolates, *S. brasiliensis* CBS 133017 (KP101458.1), *S. schenckii* CBS 117440 (KP101386.1), *S. globosa* CBS 129721 (KP101478.1), *S. luriei* ATCC 18616 (KT427639.1), *S. mexicana* Ss133 (JF811341.1), *S. chilensis* Ss470 (KP711816.1), *S. humicola* CBS 118129 (KX590808.1) and *S. phasma* CBS 119721 (KX590795.1) were included. Sequences generated in this study were deposited to NCBI Genbank (accession numbers OR501574, OR501573 and PQ741608-PQ741713) (Table S1).

4.3.3 Short tandem repeat genotyping

Genotyping of isolates was performed using short tandem repeats (STRs) as described before (16). Briefly, nine markers were amplified with three multiplex PCRs and subsequently analyzed with a 3500 XNL genetic analyzer (Applied Biosystems, Foster City, CA, USA). Copy numbers were determined using GeneMapper 5 (Applied Biosystems) and the genetic relatedness between the current and previously genotyped isolates was assessed with BioNumerics v7.6.1 (Applied Maths NV, Sint-Martems-Latem, Belgium) (16).

4.3.4 Antifungal susceptibility testing (AFST)

AFST was performed for the mycelial and yeast phase of all isolates. For the mycelial phase, isolates were cultured on Potato Dextrose Agar (Difco Laboratories, Detroit, USA) plates at 30°C for 7 days, and the absence of yeast cells was confirmed microscopically. AFST for the mycelial phase was performed by broth microdilution as outlined in the Clinical & Laboratory Standards Institute (CLSI) reference standard M38 for filamentous fungi (22). *Aspergillus flavus* ATCC 204304 was used as quality control. Amphotericin B (Bristol Myers Squib, Woerden, The Netherlands), fluconazole (Merck, Darmstadt, Germany), itraconazole (Janssen Cilag, Breda, The Netherlands), voriconazole (Pfizer Central Research, New York, USA), posaconazole

(Merck), isavuconazole (Basilea Pharmaceutica, Basel, Switzerland), terbinafine (VWR, Leicestershire, UK), anidulafungin (Merck) and micafungin (Astellas Pharma, Tokyo, Japan) were tested. The minimum inhibitory concentrations (MICs) were read visually after 72 hours of incubation at 30°C. For amphotericin B, fluconazole, itraconazole, voriconazole, posaconazole, and isavuconazole, the MIC endpoint was the lowest concentration that produced complete inhibition of growth. For echinocandins, it was the lowest concentration producing a visual change in the appearance of the growth, defined as the minimum effective concentration (MEC). For terbinafine, the MIC was defined as the lowest concentration at which there was ≤80% growth reduction when compared to the growth control. For mycelial phase, tentative epidemiological cut-off values (ECVs) according to Espinel-Ingroff were implemented for amphotericin B, itraconazole, voriconazole, posaconazole and terbinafine (23).

For the yeast phase, isolates were cultured onto Brain Heart Infusion (BHI) plates (Xebios Diagnostics GmbH, Düsseldorf, Germany), at 35°C for 7 days, followed by a second passage at BHI at 35°C for 7 days again, and the absence of filamentous fungi was confirmed microscopically. AFST was performed by broth microdilution according to the CLSI reference standard M27 for yeasts (24). *Candida krusei* ATCC 6258 and *Candida parapsilosis* ATCC 22019 were used as quality controls. The same antifungals as for the mycelial phase were tested. The MICs were read visually after 120 hours of incubation at 35°C. For amphotericin B and terbinafine, the MIC endpoint was identical as used for mycelial phase and for fluconazole, itraconazole, voriconazole, posaconazole, isavuconazole, anidulafungin and micafungin, the MIC was defined as the lowest concentration at which there was a \leq 50% growth reduction.

4.3.5 Ethics

This study was approved by the Committee for Ethics in Research of the Federal University of Paraná (number—CAAE 52726021.8.0000.0102) and by the Animal Use Ethics Committee of the Federal University of Paraná, Curitiba, Brazil.

4.4.1 Clinical epidemiology

Samples of symptomatic cats from the triple border region were collected between 2021 and 2023 and put on culture, obtaining 108 isolates of Sporothrix spp. originating from 88 households (Table S1). Of all animals, 100 were from FIG (Brazil), four from Ciudad del Este (CDE) (Paraguay) and four from Hernandarias (Paraguay). None of the animals had a previous travel history to other regions. Sporotrichosis cases were initially only found in neighborhoods in the eastern region of FIG (Figure 1A), while in following years cases were spreading to other regions, close to the country border, especially around the international bridges (Figure 1B and C). The prevalence of sporotrichosis cases was highest in the northern, southern and eastern districts (Table 1), regions with the highest density of humans (Figure S1). Furthermore, most cases were found at or close to favelas and poor urban communities (Figure S2), which are low-income dense housing settlements, characterized by poor socio-economic status, precarious conditions and lack of essential services, mostly found at the eastern region of FIG (Figure S2). Notably, all cats had easy access to the street, other homes, backyards and vacant lots. Feline sporotrichosis was more frequently in males (ration 2.8:1), adults, uncastrated cats, those not vaccinated for any disease, and those with little or no access to veterinary services, while most of them had an owner (Table 2). Characteristics of the cats with negative cultures can be seen in Table S2.

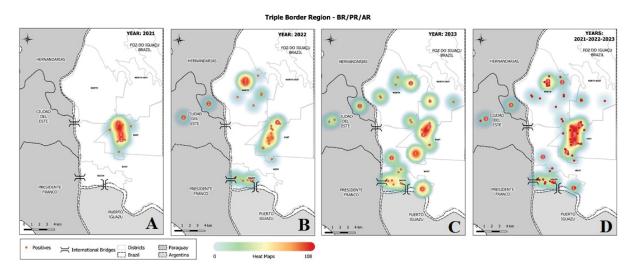


Figure 1. Heat map showing the spatial distribution of the animals with proven sporotrichosis in the triple border region (Brazil – white map; Paraguay – dark grey

map; Argentina – light grey map) between 2021 and 2023. Feline sporotrichosis cases and localization (red dots) in 2021 (A), 2022 (B), 2023 (C); and accumulated (D). Red circles with "2" and "3" refer to the number of positive cats in the same house.

District	Nºof residences	Nº of positive animals	Prevalence	in	1000
			residences		
North	32610	24	0.73		
West	32705	3	0.09		
South	16175	17	1.05		
North East	18804	4	0.21		
East	36315	52	1.43		

Table 1. Prevalence of sporotrichosis per district

Table 2. Characteristics of 108 cats with sporotrichosis

Characteristic	Category	Number of animals (%)
Sex	Male	80 (74%)
	Female	28 (26%)
Age	≤ 12 months	13 (12%)
	≥ 12 months	95 (88%)
Castration status	Uncastrated	81 (75%)
	Castrated	23 (21%)
	Unknown	4 (4%)
Vaccination status	Never vaccinated	64 (59%)
	Full vaccination scheme	11 (10%)
	Only primary vaccination	12 (11%)
	Occasional vaccination	4 (4%)
	Unknown	17 (16%)
Access to veterinary	No access	76 (70%)
	With access	32 (30%)
Tutored	With an owner	95 (88%)
	Stray cat	13 (12%)

The main clinical presentation consisted of cutaneous disseminated sporotrichosis (61%), followed by fixed cutaneous (34%) and extracutaneous (5%) sporotrichosis (Table 3). The mortality rate was 61% in those with the disseminated form, while cats with the fixed cutaneous or extracutaneous form did not die due to sporotrichosis. Ten animals were euthanized, as they lived on the street without owner, hindering any possibility of treatment. Other outcomes were lost to follow up (7%).

Clinical form	Outcome	Number of animals
Cutaneous disseminated	Death	40 (61%)
	Clinical cure	16 (24%)
	Lost to follow up	3 (4%)
	Euthanized	7 (11%)
Fixed cutaneous	Clinical cure	29 (78%)
	Lost to follow up	5 (14%)
	Euthanized	3 (8%)
Extracutaneous	Clinical cure	5 (100%)

Table 3. Outcome of 108 cats with sporotrichosis based on clinical form

All cats received itraconazole 25-100 mg daily, while those with the disseminated and extracutaneous form also received potassium iodide (2.5-20 mg/kg/24h, depending on severity of symptoms). For the clinically cured cats treatment duration ranged between two and 15 months with no correlation to the clinical form (Table 4). For the dead cats, 12 never received treatment and for the rest treatment duration ranged between 1 week and 7 months (Table 5).

Table 4. Treatment regime and duration of 50 clinical cured cats with sporotrichosis

 based on clinical form

Clinical form	Treatment regime and duration	Number of animals
Cutaneous	ITRA + PI 3 – 6 months	3
disseminated	ITRA + PI 8 – 12 months	11
	ITRA + PI 13 – 15 months	2
Fixed cutaneous	ITRA 2 – 6 months	12

	ITRA 7 – 12 months	17				
Extracutaneous	ITRA + PI 2 – 6 months	5				
ITRA, itraconazole; PI, potassium iodide.						

Table 5. Treatment regime and duration of 40 dead cats with sporotrichosis

Clinical form	Treatment regime and duration	Number of animals
Cutaneous disseminated	No treatment applied	12
	ITRA + PI 1 – 4 weeks	12
	ITRA + PI 2 – 4 months	6
	ITRA + PI 5 – 7 months	10

4.4.2 Phylogenetic analysis

With CaM sequencing, all 104 isolates were identified as *S. brasiliensis*, displaying low genetic variation within this gene (Figure S3). By performing STR genotyping, 20 genotypes were found, of which six were found earlier (16) (Figure 2). All isolates from the triple border region were highly related and all grouped within the Rio de Janeiro (RdJ) clade, a previously described dominant group of genotypes originating from RdJ (16). In this study they often clustered with isolates from other regions of Paraná, RdJ and several other states.

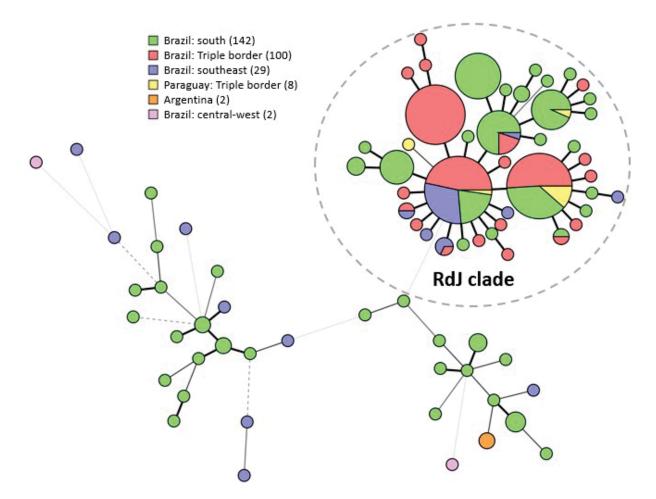


Figure 2. Minimum-spanning tree of 283 *Sporothrix brasiliensis* isolates, including 108 isolates from the current study. Isolates are colored according to different locations of origin and numbers in the color key represent the number of isolates. South: Paraná, Rio Grande do Sul; Southeast: Rio de Janeiro, Minas Gerais, São Paulo, Espírito Santo; Central-West: Federal District.

4.4.3 Minimum inhibitory concentration investigation

AFST of all isolates was performed for the mycelial and yeast phase. Based on the study of Espinel-Ingroff which proposed ECVs based on the M38 CLSI protocol, nine isolates were classified as non-wild-type for terbinafine (Table 6). For the other tested antifungals with available ECVs all isolates were classified as wildtype. For the mycelial phase, itraconazole and posaconazole had the highest in vitro activity, followed by amphotericin B. In contrast, fluconazole, voriconazole and isavuconazole, had low activity with high MICs. For the yeast phase, itraconazole, posaconazole and isavuconazole showed the highest activity followed by voriconazole and amphotericin B. Comparing susceptibility levels between both phases, isolates at the mycelial phase had higher MICs for all azoles and amphotericin B, while for terbinafine and echinocandins higher MICS were found at the yeast phase. Finally, the MIC50 of strains isolated from cats with disseminated disease that were cured were identical to those from cats that died (Table S3).

Table 6. Distribution of minimum inhibitory concentration (MIC) values against 108 clinical isolates of *Sporothrix brasiliensis* according to CLSI M38 and M27 guidelines. M lines refer to mycelial phase and Y lines refer to yeast phase. MICs in µg/mL

Antifur	ngal				≤0.063	0.125	0.25	0.5	1	2	4	8	16	32	≥64	MIC ₅₀ *
FLU	M														108	≥64
FLU Y								1	1	1	13	49	30	13	16	
			≤0.016	0.031	0.063	0.125	0.25	0.5	1	2	4	8	≥16			
AMB	Μ							1	38	57	12]		2
/	Υ					2	9	27	66	4		-				1
ITC	Μ					3	42	62	1					1		0.5
ne	Y		14	35	37	18	4									0.063
VOR	Μ											7	101	1		≥16
VOR	Y			2	8	18	26	19	21	13	1					0.25
POS	Μ					2	31	65	10					1		0.5
100	Y		11	29	36	28	4									0.063
ISA	Μ										31	51	26			8
	Υ		20	18	29	25	10	6								0.063
		≤0.008	0.016	0.031	0.063	0.125	0.25	0.5	1	2	4	≥8		-		
TRB	Μ				53	44	9									0.063
IND	Y			2	14	65	29									0.125
AFG	Μ	101	6	1												0.008
	Υ	7	9	20	34	32	6									0.063
MFG	Μ	101	2	5												0.008
	Υ	4	21	14	40	24	5									0.063

AMB, amphotericin B; FLU, fluconazole; ITC, itraconazole; VOR, voriconazole; POS, posaconazole; ISA, isavuconazole; TRB, terbinafine; AFG, anidulafungin; MFG, micafungin; M, mycelial phase; Y, yeast phase. Red dotted line indicates division of WT vs non-WT isolates based on ECV values when available. The ECV value for

voriconazole is 32 µg/mL. MIC50, the MIC value that inhibited 50% of isolates. *, for AFG and MFG the MEC50 (filamentous phase) was determined.

4.5 Discussion

This study is the first to report cases of CTS in triple border region of Brazil, Paraguay and Argentina. The epidemiological data shows that within three years S. brasiliensis-induced sporotrichosis spread across this region. The rapidly increasing number of feline cases highlights the severity of sporotrichosis as a public health problem and the potential for outbreaks (25). The data also suggests that CTS mainly affects an urban population with a high concentration of humans and likely also cats as compared to areas with a low density of humans (8,26). Notably, all cats had easy access to the street, other homes, backyards and vacant lots. Besides, cats can freely across national borders in this region and we observed introduction of sporotrichosis in Paraguay near to international bridges. Furthermore, cases correlate with poor socioeconomic status as the eastern region of FIG has the lowest standard of living. As a consequence, most cats had owners but had no access to the veterinarian (27). In most cases owners were financially not able to provide basic resources for their cats health, and in cases of sporotrichosis not capable to provide diagnosis and treatment. Therefore, public policies that provide these tools free of charge are crucial.

Although all reported sporotrichosis cases in both the Brazilian and Paraguayan side of the border were included, most isolates originated from Brazil, while the population number in the Brazilian and Paraguayan cities at the triple border region is similar (2024 population FIG: 295.000 (28), CDE: 339.000 (29)). The number of feline population is not known for both cities. The differences in sporotrichosis cases are partly due to the spread originating from the east of Brazil, which is halted by the river between both countries. In addition, differences between the healthcare systems of these two countries might play a role. In Brazil, the Health Unic System (SUS) is a decentralized system, meaning that the city decides how the resources from the state and the federal government will be used (30). In Paraguay there is a centralized health system, with most of the actions concentrated in the capital Asunción, which is 324 km from the border (30). Although the Epidemiological Laboratory in CDE, Paraguay is capable of to track cases and diagnosing

sporotrichosis in cats and humans for free, there are no enough clinicians and veterinarians available.

In this study, uncastrated and unvaccinated males represented the majority of patients with feline sporotrichosis, as previously described (31). Unvaccinated cats may have comorbidities such as FeLV, Calicivirus, Herpes, and panleukopenia, leading to immunosuppression and rapid evolution to the disseminated form of sporotrichosis (32). Castration reduces the behavior of territory disputes and sexual intercourse between males and females, which in cats usually involves fights with injuries, decreasing the chance of transmission to females and newborn kittens (33). Besides, production of testosterone can affect male's behavior. As the majority of cats in this study were over 12 months old, public interventions (vaccination and neutering programs) before this age would likely reduce the risk of transmission of sporotrichosis. Therefore, these measures are important to control and prevent sporotrichosis based on a One Health Approach, and reinforces the importance of public health education, especially about responsible feline ownership. Furthermore, the lack of awareness about this disease in health professionals is the main difficulty in identifying sporotrichosis in humans and animals, making searching for cases in these locations even harder (12). Thus, public health education on responsible feline ownership and increasing disease awareness in health professionals are the first step towards the prevention of sporotrichosis outbreaks and effective treatment of patients (34).

To show the genetic relatedness between the *S. brasiliensis* isolates, STR analysis was performed (35). All isolates clustered in the RdJ clade and were closely related to Brazilian isolates from the south and southeastern parts of the country. Curitiba and other cities in the Brazilian state of Paraná steadily report *S. brasiliensis* cases since 2011, and these isolates display identical or highly related genotypes (16). The introduction of *S. brasiliensis* in regions may happen by the movement of sick or colonized cats (11). Also, our isolates were closely related to those from Curitiba, which like FIG, belongs to Paraná state, although these regions are more than 600 kilometers apart. All our isolates clustered with the RdJ clade (16), but since different genotypes were identified, multiple introductions cannot be excluded. WGS of these isolates is needed to elucidate the origin of *S. brasiliensis* in this region.

Next, we determined MIC values for nine different antifungals against *S. brasiliensis* isolates with both pure yeast and mycelial phase, which was

microscopically confirmed. According to the interpretation of an international multicenter study for definition of tentative ECVs for mycelial S. brasiliensis (23), we found, with exception of nine non-WT isolates for terbinafine, only wild-type isolates for amphotericin B, itraconazole, voriconazole and posaconazole (36). This multicenter study used standard incubation of two to three days at 35°C according to the CLSI reference standard M38 for filamentous fungi. We found in this study that incubation at 35°C induces transition to the yeast phase taking up to two weeks for full transition. As microscopy was not performed in the multicenter study it cannot be excluded that the ECVs were established on a mixture of filamentous and yeasts phases. Moreover, many centers in this multicenter study were excluded due to insufficient or unsuitable data, suggesting suboptimal methodology or implementation thereof. Thus, additional studies are required to establish the ECVs, although the normal distribution of our MIC values indeed suggests an absence of non-WT isolates. Notably, mycelial phase MICs were overall higher than in the yeast phase, especially for the azoles fluconazole, voriconazole and isavuconazole. One explanation for this difference is the higher concentration of melanin in the cell wall in the filamentous phase. Melanin is associated with a reduced susceptibility to antifungal drugs. It is important to note however that MICs in the mycelial phase were read at 100% inhibition compared to growth control, while at the yeast phase this was 50%. Direct comparisons of the MICs between both phases should therefore be made with caution.

AFST results may not reflect in vivo treatment in the absence of clinical breakpoints (34). Nonetheless for itraconazole, which is the first-choice drug for feline treatment (15,34), MICs of all isolates in both phases were below the tentative ECV and similar results were reported earlier (17,37). In contrast, others recently found MICs of itraconazole and other azoles above the tentative ECV (19,38). Notably, the reported bimodal distribution with low and elevated MICs for itraconazole and the identification of cyp51 mutants (19) suggests that these MICs would also be well above the tentative ECV in conditions of pure mycelial and yeast phase (23). This discrepancy with our study might be due to the inclusion of different strains. Our collection consisted of one genotype only and strains were isolated before start of treatment. Smaller MIC differences could also be explained by differences in AFST protocols, including the mixed presence of filamentous and yeast phases as discussed earlier. In addition, other factors including panel preparation,

media/reagents, and inoculum preparation might influence AFST results. Finally, since the mycelial form is the one with the least variation in our genotypically similar isolates, is easiest to employ, and mycelial AFST data seemingly corresponds to in vivo failure of voriconazole (39), this phase might be most suitable for AFST. A multicenter evaluation comparing robust AFST methodologies in a genotypically variable cohort would be needed to establish the best method to determine antifungal susceptibility for *S. brasiliensis*.

We observed a high mortality in cats with the disseminated form and no sporothrichosis-related deaths in cats with the fixed cutaneous form. Importantly, all isolates were genetically similar and displayed initially low MICs of itraconazole, used for treatment in all cats. MIC50 levels of strains isolated from cats with the disseminated form that were cured were also not different from those that were not cured. Thus, the mortality in cats with the disseminated form is not due to initial elevated MICs, although we cannot exclude the development of reduced susceptibility in time, as we did not collect isolates after itraconazole treatment. The observation that transmission only involved itraconazole susceptible isolates, as we did not find an isolate with high MIC in any cat, suggests that an increase in MICs was probably uncommon, if present at all. Moreover, some cats from the same household became infected months after each other. Also in these households no elevated MICs were found. It is more likely that other factors, like disease progression, treatment variations and host factors, were involved in treatment failure in this cohort. Besides clinical outcomes, treatment duration was different in both clinical forms. Cats with the fixed cutaneous form were treated for fewer months. While disease progression is also likely to play an important role here, erratic itraconazole pharmacokinetics might be involved too. For feline sporotrichosis, the proposed drug dose by the Guideline for the management of feline sporotrichosis caused by S. brasiliensis is 100 mg/24h for cats over 3 kg. A robust dose-response study evaluating the efficacy of this dose, is lacking to our knowledge. When administering the medication, it is recommended to open the capsules over a small amount of wet food, however, there are currently no studies that verify the absorption degree of itraconazole administered in this way. In disseminated cases it is also unknown whether itraconazole can reach the mucous membranes at an adequate level for cure. However, suboptimal itraconazole blood levels prolong treatment and the risk for resistance development in other diseases (40,41). Optimal dosing to

reach effective serum itraconazole concentrations in severe disease would allow best standard of care but is currently unavailable for cats. In conclusion, early diagnosis and effective treatment seem crucial to prevent disease progression, mortality and transmission to other humans and animals, while reduced antifungal susceptibility has not been detected in the triple border region as of yet.

Conflict of interest

EFJM received research grants from Mundipharma and Scynexis, is in the scientific advisory board for Pfizer and has received speaker fees from Gilead Sciences. All other authors declare no conflict of interest.

Funding

This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Finance Code 001, the Canisius-Wilhelmina Hospital, Nijmegen, Netherlands (Grant CWZ_001421) and received fellowships from CNPq (grant number 312811/2018–7), Brasilia, Brazil. The work was supported by the Araucaria Foundation (http://www.fappr.pr.gov.br) (NAPI grant number 113/2020).

Appendices

Table S1 Accession numbers (NCBI), date of sampling and geographic data of the animals with proven sporotrichosis in the triple border region.

CMRP	ACCESSION NUMBER	DATE	NEIGHBORHOOD	CITY	COUNTRY
CMRP6060	PQ741666	07/07/2021	Jardim Alice I	Foz do Iguaçu	Brazil
CMRP5585	PQ741608	09/07/2021	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6061	PQ741667	09/07/2021	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6062	PQ741668	09/07/2021	Jardim Copacabana	Foz do Iguaçu	Brazil
CMRP5586	PQ741609	30/07/2021	Parque Residencial Morumbi	Foz do Iguaçu	Brazil
CMRP6063	PQ741669	30/07/2021	Parque Residencial Morumbi	Foz do Iguaçu	Brazil
CMRP6064	PQ741670	09/08/2021	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6065	PQ741671	09/08/2021	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6066	PQ741672	16/08/2021	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6067	PQ741673	16/08/2021	Portal da Foz	Foz do Iguaçu	Brazil

CMRP6068	PQ741674	16/08/2021	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6069	PQ741675	16/09/2021	Parque Residencial Santa Rita	Foz do Iguaçu	Brazil
CMRP5587	PQ741610	16/09/2021	Jardim Alice I	Foz do Iguaçu	Brazil
CMRP6070	PQ741676	24/09/2021	Jardim Europa	Foz do Iguaçu	Brazil
CMRP6071	PQ741677	24/09/2021	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6072	PQ741678	24/09/2021	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6073	PQ741679	24/09/2021	Parque Residencial Morumbi	Foz do Iguaçu	Brazil
CMRP5588	PQ741611	29/12/2021	Beverli Falls Park	Foz do Iguaçu	Brazil
CMRP5589	PQ741612	03/01/2022	Jardim das Flores	Foz do Iguaçu	Brazil
CMRP5590	PQ741613	18/01/2022	Jardim das Flores	Foz do Iguaçu	Brazil
CMRP5591	PQ741614	18/01/2022	Jardim das Flores	Foz do Iguaçu	Brazil
CMRP5592	PQ741615	01/02/2022	Jardim Curitibano	Foz do Iguaçu	Brazil
CMRP5593	PQ741616	04/02/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP5594	PQ741617	04/02/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP6074	PQ741680	10/03/2022	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6075	PQ741681	16/03/2022	Jardim Nacional	Foz do Iguaçu	Brazil
CMRP6077	PQ741682	05/04/2022	Cidade Nova II	Foz do Iguaçu	Brazil
CMRP6078	PQ741683	19/04/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP6079	PQ741684	19/04/2022	Cidade Nova II	Foz do Iguaçu	Brazil
CMRP6080	PQ741685	27/04/2022	Parque Residencial Santa Rita	Foz do Iguaçu	Brazil
CMRP6082	PQ741686	02/06/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP6083	PQ741687	02/06/2022	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6084	PQ741688	07/06/2022	Residencial Morumbi III	Foz do Iguaçu	Brazil
CMRP5785	PQ741618	07/06/2022	Parque Residencial Italia	Foz do Iguaçu	Brazil
CMRP6085	PQ741689	08/06/2022	Vila São Sebastião	Foz do Iguaçu	Brazil
CMRP6086	PQ741690	14/06/2022	Vila Borges	Foz do Iguaçu	Brazil
CMRP6087	PQ741691	14/06/2022	Vila Borges	Foz do Iguaçu	Brazil
CMRP6088	PQ741692	20/06/2022	Jardim São Paulo II	Foz do Iguaçu	Brazil
CMRP6089	PQ741693	21/06/2022	Jardim São Paulo II	Foz do Iguaçu	Brazil
CMRP6090	PQ741694	21/06/2022	Porto Belo	Foz do Iguaçu	Brazil
CMRP5786	OR501573	22/06/2022	-	Ciudad del Este	Paraguay
CMRP6091	PQ741695	07/07/2022	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6092	PQ741696	07/07/2022	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6093	PQ741697	14/07/2022	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6094	PQ741698	14/07/2022	Portal da Foz	Foz do Iguaçu	Brazil
CMRP6095	PQ741699	28/07/2022	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6096	PQ741700	11/08/2022	Vila Borges	Foz do Iguaçu	Brazil
CMRP6097	PQ741701	11/08/2022	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6098	PQ741702	19/08/2022	Vila São Sebastião	Foz do Iguaçu	Brazil
CMRP5787	OR501574	19/08/2022	-	Ciudad del Este	Paraguay
CMRP6099	PQ741703	24/08/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP6100	PQ741704	26/08/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP6101	PQ741705	26/08/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP6102	PQ741706	26/08/2022	Conjunto C	Foz do Iguaçu	Brazil
CMRP6103	PQ741707	06/09/2022	Jardim Residencial São Roque	Foz do Iguaçu	Brazil

CMRP6104	PQ741708	09/09/2022	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP6105	PQ741709	20/09/2022	Profilurb I	Foz do Iguaçu	Brazil
CMRP6106	PQ741710	05/10/2022	Jardim Bela Vista de Itaipu III	Foz do Iguaçu	Brazil
CMRP6107	PQ741711	05/10/2022	Jardim Oriente	Foz do Iguaçu	Brazil
CMRP6108	PQ741712	05/10/2022	Profilurb I	Foz do Iguaçu	Brazil
CMRP5791	PQ741619	30/11/2022	-	Hernandarias	Paraguay
CMRP5792	PQ741620	10/12/2022	-	Hernandarias	Paraguay
CMRP5915	PQ741621	19/01/2023	-	Hernandarias	Paraguay
CMRP5916	PQ741622	19/01/2023	-	Hernandarias	Paraguay
CMRP5917	PQ741623	12/05/2023	Portal da Foz	Foz do Iguaçu	Brazil
CMRP5918	PQ741624	12/05/2023	Portal da Foz	Foz do Iguaçu	Brazil
CMRP5919	PQ741625	25/05/2023	Portal da Foz	Foz do Iguaçu	Brazil
CMRP5940	PQ741640	25/05/2023	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP5941	PQ741641	25/05/2023	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP5921	PQ741626	01/06/2023	Vila Guarani	Foz do Iguaçu	Brazil
CMRP5922	PQ741627	08/06/2023	-	Ciudad del Este	Paraguay
CMRP5923	PQ741628	12/06/2023	Jardim Novo Horizonte	Foz do Iguaçu	Brazil
CMRP5924	PQ741629	12/06/2023	Jardim Novo Horizonte	Foz do Iguaçu	Brazil
CMRP5925	PQ741630	22/06/2023	Centro	Foz do Iguaçu	Brazil
CMRP5942	PQ741642	22/06/2023	Profilurb I	Foz do Iguaçu	Brazil
CMRP5926	PQ741631	28/06/2023	Jardim São Luiz	Foz do Iguaçu	Brazil
CMRP5927	PQ741632	28/06/2023	Jardim São Luiz	Foz do Iguaçu	Brazil
CMRP5943	PQ741643	30/06/2023	Vila A	Foz do Iguaçu	Brazil
CMRP5944	PQ741644	30/06/2023	Jd. Califórnia II	Foz do Iguaçu	Brazil
CMRP5928	PQ741633	06/07/2023	Jardim Guaira	Foz do Iguaçu	Brazil
CMRP5945	PQ741645	06/07/2023	Bubas	Foz do Iguaçu	Brazil
CMRP5946	PQ741646	11/07/2023	Profilurb I	Foz do Iguaçu	Brazil
CMRP5929	PQ741634	11/07/2023	Cohapar III	Foz do Iguaçu	Brazil
CMRP5947	PQ741647	13/07/2023	Conjunto C	Foz do Iguaçu	Brazil
CMRP5948	PQ741648	13/07/2023	Loteamento Bela Vista de Itaipu II	Foz do Iguaçu	Brazil
CMRP5930	PQ741635	13/07/2023	Jardim Cedro	Foz do Iguaçu	Brazil
CMRP5931	PQ741636	20/07/2023	Conjunto C	Foz do Iguaçu	Brazil
CMRP5949	PQ741649	20/07/2023	Portal da Foz	Foz do Iguaçu	Brazil
CMRP5932	PQ741637	26/07/2023	Jardim Duarte	Foz do Iguaçu	Brazil
CMRP5950	PQ741650	28/07/2023	Jardim Novo Horizonte	Foz do Iguaçu	Brazil
CMRP5933	PQ741638	28/07/2023	Jardim Novo Horizonte	Foz do Iguaçu	Brazil
CMRP5935	PQ741639	28/07/2023	Jardim Eliza II	Foz do Iguaçu	Brazil
CMRP5952	PQ741651	16/08/2023	Jardim California	Foz do Iguaçu	Brazil
CMRP5953	PQ741652	16/08/2023	Centro	Foz do Iguaçu	Brazil
CMRP5955	PQ741653	19/08/2023	-	Ciudad del Este	Paraguay
CMRP5956	PQ741654	24/08/2023	Jardim das Flores	Foz do Iguaçu	Brazil
CMRP5957	PQ741655	24/08/2023	Parque Residencial Morumbi II	Foz do Iguaçu	Brazil
CMRP5958	PQ741656	04/09/2023	Jardim São Paulo II	Foz do Iguaçu	Brazil
CMRP5959	PQ741657	04/09/2023	Jardim São Paulo II	Foz do Iguaçu	Brazil
CMRP5960	PQ741658	04/09/2023	Conjunto Habitacional Fernanda	Foz do Iguaçu	Brazil

CMRP5961	PQ741659	06/09/2023	Jardim Tropical	Foz do Iguaçu	Brazil
CMRP5962	PQ741660	06/09/2023	Parque Ouro Verde	Foz do Iguaçu	Brazil
CMRP5963	PQ741661	12/09/2023	Vila Guarani	Foz do Iguaçu	Brazil
CMRP5964	PQ741662	12/09/2023	Parque Residencial Morumbi III	Foz do Iguaçu	Brazil
CMRP5965	PQ741663	13/09/2023	Cohapar III	Foz do Iguaçu	Brazil
CMRP5966	PQ741664	15/09/2023	Loteamento Bela Vista de Itaipu II	Foz do Iguaçu	Brazil
CMRP5967	PQ741665	15/09/2023	Loteamento Bela Vista de Itaipu II	Foz do Iguaçu	Brazil
CMRP6111	PQ741713	25/10/2023	Jardim São Luiz	Foz do Iguaçu	Brazil

 Table S2.
 Characteristics of 25 cats without sporotrichosis.

Characteristic	Category	Number of animals (%)
Sex	Male	17 (68%)
	Female	8 (32%)
Age	≤ 12 months	3 (12%)
	≥ 12 months	22 (88%)
Castration status	Uncastrated	12 (48%)
	Castrated	12 (48%)
	Unknown	1 (4%)
Vaccination status	Never vaccinated	14 (56%)
	Full vaccination scheme	4 (16%)
	Only primary vaccination	3 (12%)
	Occasional vaccination	3 (12%)
	Unknown	1 (4%)
Access to veterinary	No access	22 (88%)
	With access	3 (12%)
Tutored	With an owner	24 (%)
	Stray cat	1 (%)

Table S3. Comparison between clinical outcome with MIC50.

Clinical form	Outcome	MIC50																	
		AMB		FLU		ITC		VOR		POS		ISA		TRB		AFG		MCF	=
		М	Y	М	Y	M	Y	М	Y	М	Y	М	Y	М	Y	М	Y	М	Y
Cutaneous	Death	2	1	64	16	0.	0.	16	0.	0.5	0.06	8	0.	0.0	0.	0.0	0.	0.	0.
disseminated						5	0		3		3		06	63	12	08	06	00	06
							6		7				3		5		3	8	3
							3		5										

	Clinical cure	2	1	64	16	0.	0.	16	0.	0.5	0.06	8	0.	0.0	0.	0.0	0.	0.	0.
						5	0		5		3		06	63	12	08	12	00	06
							4						3		5		5	8	3
							7												
	Lost to	2	1	64	16	0.	0.	16	1	0.5	0.12	8	0.	0.0	0.	0.0	0.	0.	0.
	follow up					5	1				5		25	63	06	08	06	00	06
							2								3		3	8	3
							5												
	Euthanized	1	0.	64	32	0.	0.	16	1	0.25	0.06	8	0.	0.0	0.	0.0	0.	0.	0.
			5			25	0				3		06	63	12	08	03	00	06
							3						3		5		1	8	3
							1												
Fixed	Clinical cure	2	1	64	16	0.	0.	16	0.	0.5	0.06	8	0.	0.0	0.	0.0	0.	0.	0.
cutaneous						5	0		2		3		06	63	12	08	06	00	06
							6		5				3		5		3	8	3
							3												
	Lost to	1	0.	64	32	0.	0.	16	0.	0.25	0.06	8	0.	0.1	0.	0.0	0.	0.	0.
	follow up		5			25	0		2		3		06	25	12	08	12	00	06
							6		5				3		5		5	8	3
							3												
	Euthanized	2	1	64	32	0.	0.	16	0.	0.5	0.03	8	0.	0.1	0.	0.0	0.	0.	0.
						5	0		2		1		01	25	12	08	03	00	01
							3		5				6		5		1	8	6
							1												
Extracutaneous	Clinical cure	2	1	64	16	0.	0.	16	0.	0.5	0.06	16	0.	0.1	0.	0.0	0.	0.	0.
						5	0		5		3		06	25	25	08	01	00	06
							6						3				6	8	3
							3												
															I				

Residences Per District

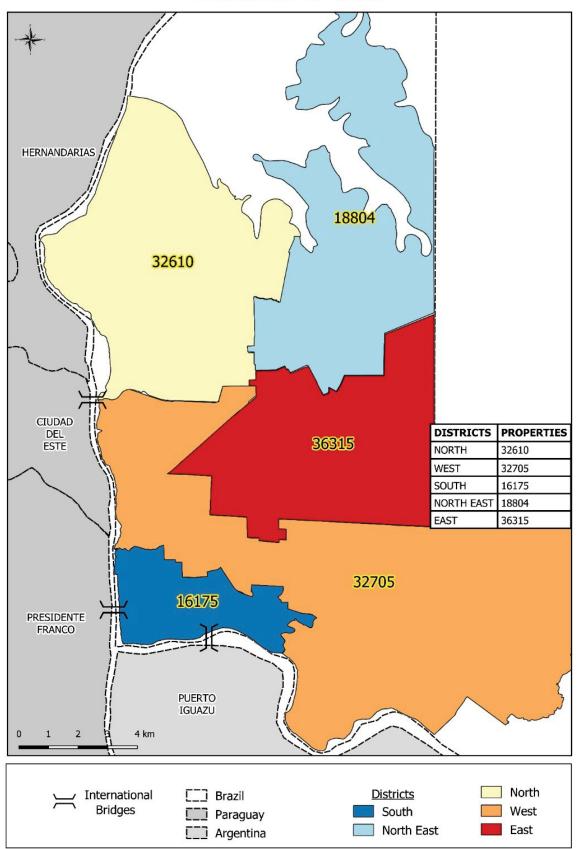


Figure S1. Number of residences per district of Foz do Iguaçu, Brazil.

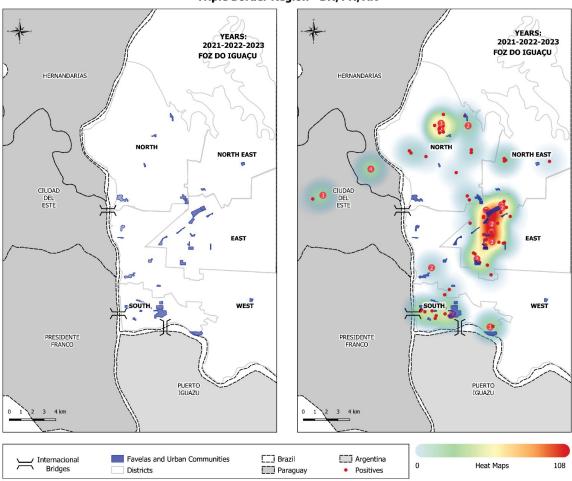


Figure S2. On the left side is the location of the favelas and poor urban communities (FPUC) in Foz do Iguaçu, Brazil. On the right side is the Map with FPUC + all sporotrichosis cases 2021-2023.

78

Triple Border Region - BR/PR/AR

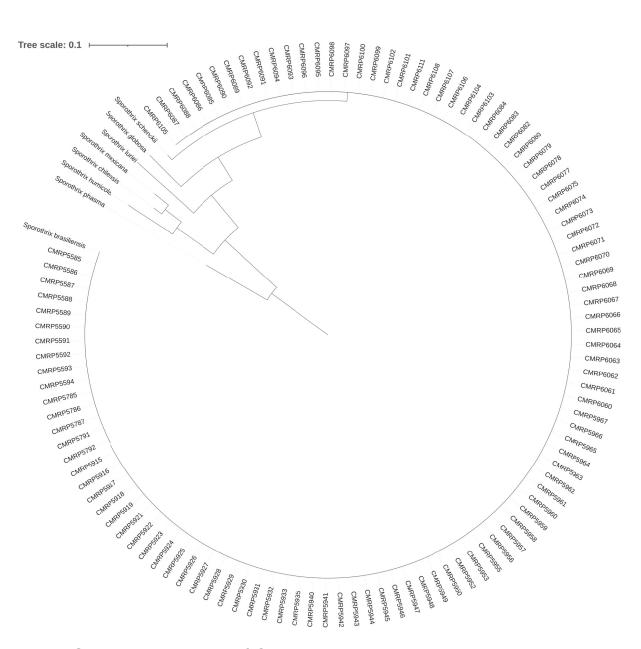


Figure S3. Phylogenetic tree of *Sporothrix brasiliensis* based on multiple sequence alignment of calmodulin sequences. The tree was rooted to *Sporothrix phasma*.

4.6 References

1. Rossow JA, Queiroz-Telles F, Caceres DH, Beer KD, Jackson BR, Pereira JG, et al. A One Health Approach to Combatting *Sporothrix brasiliensis*: Narrative Review of an Emerging Zoonotic Fungal Pathogen in South America. J Fungi (Basel).

2020;6(4):247. doi: 10.3390/jof6040247

2. Rodrigues AM, Della Terra PP, Gremião ID, Pereira SA, Orofino-Costa R, de Camargo ZP. The threat of emerging and re-emerging pathogenic *Sporothrix* species. Mycopathologia. 2020;185(5):813-842. doi:10.1007/s11046-020-00425-0

3. Rodrigues AM, Gonçalves SS, de Carvalho JA, Borba-Santos LP, Rozental S, Camargo ZP de. Current Progress on Epidemiology, Diagnosis, and Treatment of Sporotrichosis and Their Future Trends. J Fungi (Basel). 2022;8(8).

doi:10.3390/jof8080776

4. do Prado CM, Razzolini E, Santacruz G, Ojeda L, Geraldo MR, Segovia N, et al. First Cases of Feline Sporotrichosis Caused by *Sporothrix brasiliensis* in Paraguay. J Fungi (Basel). 2023;9(10):972. doi: 10.3390/jof9100972

 5. Etchecopaz A, Toscanini MA, Gisbert A, Mas J, Scarpa M, Iovannitti CA, et al. Sporothrix brasiliensis: A Review of an Emerging South American Fungal Pathogen, Its Related Disease, Presentation and Spread in Argentina. J Fungi (Basel).
 2021;7(3):170. doi: 10.3390/jof7030170

Thomson P, González C, Blank O, Ramírez V, Río CD, Santibáñez S, et al.
 Sporotrichosis Outbreak Due to *Sporothrix brasiliensis* in Domestic Cats in
 Magallanes, Chile: A One-Health-Approach Study. J Fungi (Basel). 2023;9(2):226.
 doi: 10.3390/jof9020226

7. Gallo S, Arias-Rodriguez C, Sánchez-Cifuentes EA, Santa-Vélez C, Larrañaga-Piñeres I, Gaviria-Barrera ME, et al. First three cases of cat-associated zoonotic cutaneous sporotrichosis in Colombia. Int J Dermatol. 2022;61(10):1276-1279. doi: 10.1111/ijd.16377

8. Cognialli RCR, Cáceres DH, Bastos FAGD, Cavassin FB, Lustosa BPR, Vicente VA, et al. Rising Incidence of *Sporothrix brasiliensis* Infections, Curitiba, Brazil, 2011-2022. Emerg Infect Dis. 2023 Jul;29(7):1330-1339. doi: 10.3201/eid2907.230155

 Rabello VBS, Almeida MA, Bernardes-Engemann AR, Almeida-Paes R, de Macedo PM, Zancopé-Oliveira RM. The Historical Burden of Sporotrichosis in Brazil: a Systematic Review of Cases Reported from 1907 to 2020. Braz J Microbiol.
 2022;53(1):231-244. doi: 10.1007/s42770-021-00658-1

10. Etchecopaz AN, Lanza N, Toscanini MA, Devoto TB, Pola SJ, Daneri GL, et al. Sporotrichosis caused by *Sporothrix brasiliensis* in Argentina: Case report, molecular identification and in vitro susceptibility pattern to antifungal drugs. J Mycol Med. 2020;30(1):100908. doi: 10.1016/j.mycmed.2019.100908

11. Barnacle JR, Chow YJ, Borman AM, Wyllie S, Dominguez V, Russell K, et al. The first three reported cases of *Sporothrix brasiliensis* cat-transmitted sporotrichosis outside South America. Med Mycol Case Rep. 2022;20;39:14-17. doi:

10.1016/j.mmcr.2022.12.004

12. Kaadan MI, Dennis M, Desai N, Yadavalli G, Lederer P. One Health Education for Future Physicians: A Case Report of Cat-Transmitted Sporotrichosis. Open Forum Infect Dis. 2020;7(3):ofaa049. doi: 10.1093/ofid/ofaa049

13. Queiroz-Telles F, Cognialli RC, Salvador GL, Moreira GA, Herkert PF, et al. Cutaneous disseminated sporotrichosis in immunocompetent patient: Case report and literature review. Med Mycol Case Rep. 2022;36:31-34. doi:

10.1016/j.mmcr.2022.05.003.

14. Bastos F, Farias M, Monti F, Cognialli R, Lopuch L, Gabriel A, et al. P462 Spread of *Sporothrix brasiliensis* from the sneeze of infected cats: a potential novel route of transmission. Med Mycol. 2022;60(1):myac072P462. doi:

10.1093/mmy/myac072.P462

15. Gremião IDF, Martins da Silva da Rocha E, Montenegro H, Carneiro AJB, Xavier MO, de Farias MR, et al. Guideline for the management of feline sporotrichosis

caused by Sporothrix brasiliensis and literature revision. Braz J Microbiol.

2021;52(1):107-124. doi: 10.1007/s42770-020-00365-3

16. Spruijtenburg B, Bombassaro A, Meijer EFJ, Rodrigues AM, Grisolia ME, Vicente VA, et al. *Sporothrix brasiliensis* genotyping reveals numerous independent zoonotic introductions in Brazil. J Infect. 2023;86(6):610-613. doi: 10.1016/j.jinf.2023.02.034

17. Roldán Villalobos W, Monti F, Ferreira T, Sato S, Telles F, Farias M. Therapeutic efficacy of isavuconazole and potassium iodide in a cat with refractory sporotrichosis. Vet Dermatol. 2023;34(6):624-628. doi: 10.1111/vde.13188

18. Nakasu CCT, Waller SB, Ripoll MK, Ferreira MRA, Conceição FR, Gomes ADR, et al. Feline sporotrichosis: a case series of itraconazole-resistant *Sporothrix brasiliensis* infection. Braz J Microbiol. 2021;52(1):163-171. doi: 10.1007/s42770-020-00290-5

19. Ribeiro Dos Santos A, Gade L, Misas E, Litvintseva AP, Nunnally NS, Parnell LA, et al. Bimodal distribution of azole susceptibility in *Sporothrix brasiliensis* isolates in Brazil. Antimicrob Agents Chemother. 2024;68(4):e0162023. doi:

10.1128/aac.01620-23

20. Ramos MLM, Almeida-Silva F, de Souza Rabello VB, Nahal J, Figueiredo-Carvalho MHG, Bernardes-Engemann AR, et al. In vitro activity of the anthelmintic drug niclosamide against *Sporothrix* spp. strains with distinct genetic and antifungal susceptibility backgrounds. Braz J Microbiol. 2024;55(2):1359-1368. doi:

10.1007/s42770-024-01301-5

21. Waller SB, Ripoll MK, Pierobom RM, Rodrigues PRC, Costa PPC, Pinto FDCL, et al. Screening of alkaloids and withanolides isolated from Solanaceae plants for antifungal properties against non-wild type *Sporothrix brasiliensis*. J Mycol Med. 2024;34(1):101451. doi: 10.1016/j.mycmed.2023.101451

22. Clinical and Laboratory Standards Institute (CLSI). Reference method for broth dilution antifungal susceptibility testing of filamentous fungi. 3rd ed. Pennsylvania: CLSI standard M38, 2017.

23. Espinel-Ingroff A, Abreu DPB, Almeida-Paes R, Brilhante RSN, Chakrabarti A, Chowdhary A, et al. Multicenter, International Study of MIC/MEC Distributions for Definition of Epidemiological Cutoff Values for *Sporothrix* Species Identified by Molecular Methods. Antimicrob Agents Chemother. 2017;61(10):e01057-17. doi: 10.1128/AAC.01057-17

24. Clinical and Laboratory Standards Institute (CLSI). Reference method for broth dilution antifungal susceptibility testing of yeasts. 4th. Pennsylvania: CLSI standard M27, 2017.

25. Rodrigues AM, de Melo Teixeira M, de Hoog GS, Schubach TM, Pereira SA, Fernandes GF, et al. Phylogenetic analysis reveals a high prevalence of *Sporothrix brasiliensis* in feline sporotrichosis outbreaks. PLoS Negl Trop Dis. 2013;7(6):e2281. doi: 10.1371/journal.pntd.0002281

26. Silva CE, Valeriano CA, Ferraz CE, Neves RP, Oliveira MM, Silva JC, et al. Epidemiological features and geographical expansion of sporotrichosis in the state of Pernambuco, northeastern Brazil. Future Microbiol. 2021;16:1371-1379. doi:

10.2217/fmb-2021-0142

27. Alzuguir CLC, Pereira SA, Magalhães MAFM, Almeida-Paes R, Freitas DFS, Oliveira LFA, et al. Geo-epidemiology and socioeconomic aspects of human sporotrichosis in the municipality of Duque de Caxias, Rio de Janeiro, Brazil, between 2007 and 2016. Trans R Soc Trop Med Hyg. 2020;114(2):99-106. doi: 10.1093/trstmh/trz081 28. IBGE – INSTITUTO BRASILEIRO DE GEOGRAFIA E ESTATÍSTICA . Cidades: Foz do Iguaçu. Rio de Janeiro: IBGE, 2022. [cited 2024 Nov]. Available from: https://cidades.ibge.gov.br/brasil/pr/foz-do-iguacu/panorama.

29. World Population Review. Ciudad del Este, Paraguay Population 2024. [cited 2024 Nov]. Available from: https://worldpopulationreview.com/cities/paraguay/ciudad-del-este.

30. do Prado CM, Svoboda WK, Chiyo L, Queiroz-Telles F. Fundamentos de Saúde Única (One Health) e Planejamento Estratégico Situacional para Implementação de Política Pública de Saúde para Prevenção e Controle da Esporotricose na Região da Tríplice Fronteira (Brasil, Paraguai, Argentina). In: Zilly A, da Silva RMM, eds. Saúde Pública Na Região Da Fronteira Brasil-Paraguai-Argentina. São Carlos: Pedro & João Editores; 2022. p101-118.

31. Pereira SA, Gremião IDF, Kitada AAB, Boechat JS, Viana PG, Schubach TMP. The epidemiological scenario of feline sporotrichosis in Rio de Janeiro, State of Rio de Janeiro, Brazil. Rev Soc Bras Med Trop. 2014;47(3):392-393. doi:10.1590/0037-8682-0092-2013

32. de Miranda LHM, Meli M, Conceição-Silva F, Novacco M, Menezes RC, Pereira SA, et al. Co-infection with feline retrovirus is related to changes in immunological parameters of cats with sporotrichosis. PLoS One. 2018;13(11):e0207644. doi:

10.1371/journal.pone.0207644

33. Araújo AA, Codeço C, Freitas DFS, de Macedo PM, Pereira SA, Gremião IDF, et al. Mathematical model of the dynamics of transmission and control of sporotrichosis in domestic cats. PLoS One. 2023;18(2):e0272672. doi:

10.1371/journal.pone.0272672

34. Lloret A, Hartmann K, Pennisi MG, Ferrer L, Addie D, Belák S, et al.

Sporotrichosis in cats: ABCD guidelines on prevention and management. J Feline Med Surg. 2013;15(7):619-23. doi: 10.1177/1098612X13489225

35. Fernandez NB, Spruijtenburg B, Tiraboschi IN, Meis JF, Lugo A, López Joffre MC, et al. Genotyping and clonal origin of *Sporothrix brasiliensis* in human sporotrichosis cases in Argentina. Med Mycol Case Rep. 2024;43:100633. doi: 10.1016/j.mmcr.2024.100633

36. Fichman V, Almeida-Silva F, Francis Saraiva Freitas D, Zancopé-Oliveira RM, Gutierrez-Galhardo MC, Almeida-Paes R. Severe Sporotrichosis Caused by *Sporothrix brasiliensis*: Antifungal Susceptibility and Clinical Outcomes. J Fungi (Basel). 2022;9(1):49. doi: 10.3390/jof9010049

37. Reis EGD, Pereira SA, Miranda LHM, Oliveira RVC, Quintana MSB, Viana PG, et al. A Randomized Clinical Trial Comparing Itraconazole and a Combination Therapy with Itraconazole and Potassium Iodide for the Treatment of Feline Sporotrichosis. J Fungi (Basel). 2024;10(2):101. doi: 10.3390/jof10020101

38. Waller SB, Dalla Lana DF, Quatrin PM, Ferreira MRA, Fuentefria AM, Mezzari A. Antifungal resistance on *Sporothrix* species: an overview. Braz J Microbiol.

2021;52(1):73-80. doi: 10.1007/s42770-020-00307-z

 39. Fernández-Silva F, Capilla J, Mayayo E, Guarro J. Modest efficacy of voriconazole against murine infections by *Sporothrix schenckii* and lack of efficacy against *Sporothrix brasiliensis*. Mycoses. 2014;57(2):121-4. doi: 10.1111/myc.12112
 40. Chen Y, Ma F, Lu T, Budha N, Jin JY, Kenny JR, et al. Development of a Physiologically Based Pharmacokinetic Model for Itraconazole Pharmacokinetics and Drug-Drug Interaction Prediction. Clin Pharmacokinet. 2016;55(6):735-49. doi: 10.1007/s40262-015-0352-5 41. Prentice AG, Glasmacher A. Making sense of itraconazole pharmacokinetics. J Antimicrob Chemother. 2005;56 Suppl 1:i17-i22. doi: 10.1093/jac/dki220

CONCLUSION

This research identified the first autochthonous cases of feline and human sporotrichosis caused by *S. brasiliensis* in the border region between Brazil and Paraguay, as well as an emerging outbreak in Argentina. It was also revealed that treatment failures in cats are not due to pre-existing itraconazole resistance but may be related to other factors, such as pharmacokinetic/pharmacodynamic issues, disease progression, and host conditions.

These findings highlight the urgent need to strengthen disease surveillance, increase public awareness, such as educating veterinarians, health professionals, and the public about early detection and preventive measures, and adopt a coordinated One Health approach to address this growing public health threat in Latin America.

The increase in cases in Latin America, especially in high transboundary mobility regions, like the one we studied, presents significant challenges for public health authorities. Climate change, urbanization, and high densities of cats with access to the street exacerbate the spread of sporotrichosis.

The study emphasizes the importance of early diagnosis and effective treatment to prevent disease progression and transmission. Public health measures, such as vaccination and sterilization programs for cats, are critical for controlling sporotrichosis. Additionally, raising awareness among health professionals and the public is essential for timely interventions.

Molecular diagnostics such as Short Tandem Repeat Genotyping are useful to monitor the spread of *S. brasiliensis* and its introduction to new areas. Future studies are warranted to assess whether this method for AFST is better reproducible between laboratories and whether the high mortality can be attributed to mere late clinical presentation and/or pharmacokinetics and pharmacodynamics (PK/PD) issues in cats.

REFERENCES

ALZUGUIR, C. L. C. et al. Geo-epidemiology and socioeconomic aspects of human sporotrichosis in the municipality of Duque de Caxias, Rio de Janeiro, Brazil, between 2007 and 2016. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, v. 114, n. 2, p. 99-106, 2020. DOI: 10.1093/trstmh/trz081.

ARAÚJO, A. A. et al. Mathematical model of the dynamics of transmission and control of sporotrichosis in domestic cats. *PLoS One*, v. 18, n. 2, e0272672, 2023. DOI: 10.1371/journal.pone.0272672.

BARNACLE, J. R. et al. The first three reported cases of *Sporothrix brasiliensis* cattransmitted sporotrichosis outside South America. *Medical Mycology Case Reports*, v. 39, p. 14–17, 2023.

BOMBASSARO, A. et al. Genotyping and antifungal susceptibility testing of *Sporothrix brasiliensis* isolates from Southern Brazil. *Mycoses*, v. 66, n. 7, p. 585-593, 2023.

CASTRO, R. A. et al. Differences in cell morphometry, cell wall topography and Gp70 expression correlate with the virulence of *Sporothrix brasiliensis* clinical isolates. *PLoS ONE*, v. 8, e75656, 2013.

CLINICAL AND LABORATORY STANDARDS INSTITUTE (CLSI). *Reference method for broth dilution antifungal susceptibility testing of filamentous fungi*. 3rd ed. Pennsylvania: CLSI standard M38, 2017.

CHEN, Y. et al. Development of a physiologically based pharmacokinetic model for itraconazole pharmacokinetics and drug-drug interaction prediction. *Clinical Pharmacokinetics*, v. 55, n. 6, p. 735-749, 2016. DOI: 10.1007/s40262-015-0352-5.

CLINICAL AND LABORATORY STANDARDS INSTITUTE (CLSI). *Reference method for broth dilution antifungal susceptibility testing of yeasts*. 4th ed. Pennsylvania: CLSI standard M38, 2017.

COGNIALLI, R. C. R. et al. Rising incidence of *Sporothrix brasiliensis* infections, Curitiba, Brazil, 2011–2022. *Emerging Infectious Diseases*, v. 29, p. 1330–1339, 2023.

CORDOBA, S. et al. Molecular identification and susceptibility profile of *Sporothrix schenckii* sensu lato isolated in Argentina. *Mycoses*, v. 61, p. 441–448, 2018. DOI: 10.1111/myc.12760.

CRISOSTOMO, R. M. et al. Diagnostic accuracy and cost-benefit analysis of a new serologic test to detect sporotrichosis due to *Sporothrix brasiliensis* in serum samples from cats. *Journal of Fungi*, v. 9, n. 5, p. 509, 2023. DOI: 10.3390/jof9050509.

FAGUNDES-PEREYRA, W. J. et al. Cell-free antigens from *Sporothrix brasiliensis* lead to protection against sporotrichosis in mice. *Medical Mycology*, v. 58, n. 1, p. 37–45, 2020. DOI: 10.1093/mmy/myz019.

FERREIRA, L. S. et al. Histopathological patterns of feline sporotrichosis due to *Sporothrix brasiliensis. Mycopathologia*, v. 183, n. 3, p. 505–517, 2018. DOI: 10.1007/s11046-017-0175-1.

GARCÍA-LOMBARDÍA, J. et al. Human sporotrichosis: recommendations from the Working Group of the Spanish Mycology Association (AEM). *Revista Iberoamericana de Micología*, v. 40, n. 1, p. 1–9, 2023. DOI: 10.1016/j.riam.2022.09.004.

GREMION, G. et al. First case of feline sporotrichosis in Switzerland: diagnostic and therapeutic challenges. *Medical Mycology Case Reports*, v. 41, p. 14–17, 2023. DOI: 10.1016/j.mmcr.2023.01.002.

GREMION, G. et al. The international emergence of *Sporothrix brasiliensis*: a public health concern. *The Lancet Microbe*, 2024. DOI: 10.1016/S2666-5247(23)00318-7.

KASAI, M. et al. Comparison of the efficacy and safety of oral potassium iodide, itraconazole, and terbinafine for sporotrichosis: a retrospective cohort study. *Journal of Dermatology*, v. 51, n. 1, p. 46–53, 2024.

LANDAZURI, A. et al. Itraconazole capsule dose proportionality and pharmacokinetics in healthy male volunteers. *Pharmacotherapy*, v. 26, n. 4, p. 469–476, 2006.

LIMA-NETO, R. G. de et al. Development of *Sporothrix brasiliensis* keratitis in a patient with a history of recurrent herpetic keratitis: a case report. *Case Reports in Ophthalmological Medicine*, 2023. DOI: 10.1155/2023/5580200.

LOPES-BORGES, J. et al. High frequency of itraconazole resistance in *Sporothrix brasiliensis* from feline sporotrichosis cases in Southern Brazil. *Journal of Fungi*, v. 9, n. 2, p. 166, 2023.

MIRANDA, L. H. M. et al. Large outbreak of sporotrichosis in humans and cats, Rio de Janeiro, Brazil. *Emerging Infectious Diseases*, v. 17, n. 2, p. 197–204, 2011. DOI: 10.3201/eid1702.100812.

OLIVEIRA, M. M. E. de et al. The sporotrichosis outbreak in Brazil: towards a diagnostic strategy for suspected cases. *PLoS Neglected Tropical Diseases*, v. 12, n. 12, p. e0006842, 2018. DOI: 10.1371/journal.pntd.0006842.

OLIVEIRA, M. M. E. de et al. Development and evaluation of a serodiagnostic enzyme-linked immunosorbent assay using a purified and specific antigen to detect *Sporothrix brasiliensis* infection. *Journal of Clinical Microbiology*, v. 53, n. 12, p. 4074–4080, 2015. DOI: 10.1128/JCM.02072-15.

OLIVEIRA, M. M. E. de et al. In vitro activity of antifungal drugs against *Sporothrix brasiliensis*, *S. schenckii*, and *S. globosa* clinical isolates. *Antimicrobial Agents and Chemotherapy*, v. 55, n. 9, p. 4480–4483, 2011. DOI: 10.1128/AAC.00176-11.

RIBEIRO, L. S. et al. First isolation of *Sporothrix brasiliensis* from the nail of a domestic cat in Rio de Janeiro, Brazil. *Mycopathologia*, v. 185, n. 5, p. 803–807, 2020. DOI: 10.1007/s11046-020-00478-z.

RODRIGUES, A. M. et al. The threat of emerging and re-emerging fungal infections in animals and humans. *The Journal of Fungi*, v. 7, n. 9, p. 696, 2021. DOI: 10.3390/jof7090696.

RODRIGUES, A. M. et al. Sporothrix brasiliensis: a review of an emerging South American fungal pathogen, its related disease and management. *Medical Mycology*, v. 58, n. 5, p. 579–593, 2020. DOI: 10.1093/mmy/myz127.

SANCHES, F. C. et al. Combined treatment with potassium iodide and itraconazole in a case of cutaneous sporotrichosis: a successful alternative for refractory cases. *Revista da Sociedade Brasileira de Medicina Tropical*, v. 56, e0134, 2023. DOI: 10.1590/0037-8682-0482-2022.

SANTOS, C. et al. Antifungal susceptibility of *Sporothrix* species collected in Brazil. *Medical Mycology*, v. 59, n. 5, p. 472–478, 2021. DOI: 10.1093/mmy/myab004.

SANTOS, I. B. dos et al. *Sporothrix brasiliensis* isolates from feline sporotrichosis show differential virulence in *Galleria mellonella*. *Virulence*, v. 14, n. 1, p. 2223174, 2023. DOI: 10.1080/21505594.2023.2223174.

SANTOS, K. T. et al. Virulence and antifungal susceptibility of *Sporothrix brasiliensis* strains isolated from human cases of sporotrichosis in São Paulo, Brazil. *Journal of Fungi*, v. 8, n. 11, p. 1141, 2022. DOI: 10.3390/jof8111141.

SARAIVA, F. M. S. et al. Association of diabetes mellitus and sporotrichosis: A retrospective study. *PLOS ONE*, v. 18, n. 2, p. e0281026, 2023. DOI: 10.1371/journal.pone.0281026.

SILVA, G. R. da et al. Histopathological features and detection of *Sporothrix brasiliensis* in skin samples of cats with sporotrichosis. *Journal of Comparative Pathology*, v. 178, p. 10–19, 2020. DOI: 10.1016/j.jcpa.2020.06.004.

SILVA, M. B. T. da et al. Rapid and efficient diagnosis of feline sporotrichosis by direct examination of cutaneous lesions. *The Veterinary Journal*, v. 187, n. 2, p. 258–260, 2011. DOI: 10.1016/j.tvjl.2009.11.014.

TOLEDO, C. E. C. et al. Investigation of circulating microRNAs as potential biomarkers in sporotrichosis. *Journal of Fungi*, v. 9, n. 1, p. 104, 2023. DOI: 10.3390/jof9010104.

XAVIER, M. O. et al. Sporotrichosis in felines and canines in Rio Grande do Sul, Brazil: a retrospective study (2005–2015). *Mycoses*, v. 61, n. 1, p. 34–39, 2018. DOI: 10.1111/myc.12708.

ZHANG, Y. et al. Phylogeography and evolutionary patterns in *Sporothrix* spanning more than 100 years. *Mycoses*, v. 68, n. 1, p. 3–15, 2025. DOI: 10.1111/myc.13976.