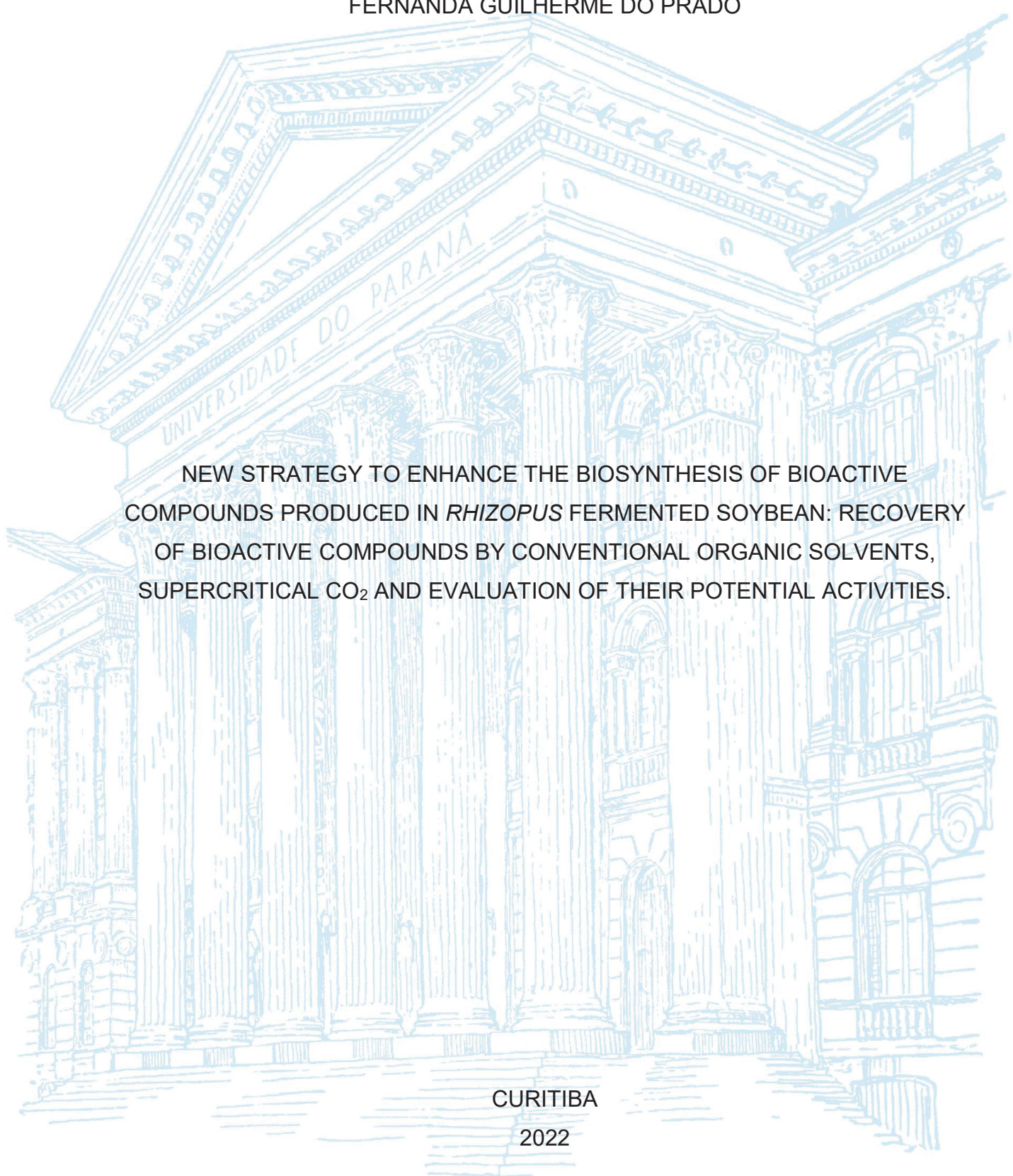


UNIVERSIDADE FEDERAL DO PARANÁ

FERNANDA GUILHERME DO PRADO



NEW STRATEGY TO ENHANCE THE BIOSYNTHESIS OF BIOACTIVE
COMPOUNDS PRODUCED IN *RHIZOPUS* FERMENTED SOYBEAN: RECOVERY
OF BIOACTIVE COMPOUNDS BY CONVENTIONAL ORGANIC SOLVENTS,
SUPERCRITICAL CO₂ AND EVALUATION OF THEIR POTENTIAL ACTIVITIES.

CURITIBA

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Tese apresentada ao curso de Pós-Graduação em Engenharia de Bioprocessos e Biotecnologia, Setor de Tecnologia, Universidade Federal do Paraná, como requisito parcial à obtenção do título de Doutor em Engenharia de Bioprocessos e Biotecnologia.

Orientador: Prof^o. Dr^o: Carlos Ricardo Soccol

CURITIBA

2022

DADOS INTERNACIONAIS DE CATALOGAÇÃO NA PUBLICAÇÃO (CIP)
UNIVERSIDADE FEDERAL DO PARANÁ
SISTEMA DE BIBLIOTECAS – BIBLIOTECA CIÊNCIA E TECNOLOGIA

Prado, Fernanda Guilherme do.

New strategy to enhance the biosynthesis of bioactive compounds produced in *Rhizopus* fermented soybean : recovery of bioactive compounds by conventional organic solvents, supercritical CO₂ and evaluation of their potential activities. / Fernanda Guilherme do Prado. – Curitiba, 2022.

1 recurso on-line : PDF.

Tese (Doutorado) – Universidade Federal do Paraná, Setor de Tecnologia, Programa de Pós-Graduação em Engenharia de Bioprocessos e Biotecnologia.

Orientador Prof. Dr. Carlos Ricardo Soccol.

1. Biotecnologia. 2. Engenharia - Alimentos. 3. Soja como alimento. 4. Indústria alimentícia. I. Soccol, Carlos Ricardo. II. Universidade Federal do Paraná. Programa de Pós-Graduação em Engenharia de Bioprocessos e Biotecnologia. III. Título.

Bibliotecário: Nilson Carlos Vieira Junior CRB-9/1797



MINISTÉRIO DA EDUCAÇÃO
SETOR DE TECNOLOGIA
UNIVERSIDADE FEDERAL DO PARANÁ
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BIOPROCESSOS E BIOTECNOLOGIA - 40001016036P8

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Os membros da Banca Examinadora designada pelo Colegiado do Programa de Pós-Graduação ENGENHARIA DE BIOPROCESSOS E BIOTECNOLOGIA da Universidade Federal do Paraná foram convocados para realizar a arguição da tese de Doutorado de **FERNANDA GUILHERME DO PRADO** intitulada: **New strategy to enhance the biosynthesis of bioactive compounds produced in Rhizopus fermented soybean: Recovery of bioactive compounds by conventional organic solvents, supercritical CO₂ and evaluation of their potential activities**, sob orientação do Prof. Dr. CARLOS RICARDO SOCCOL, que após terem inquirido a aluna e realizada a avaliação do trabalho, são de parecer pela sua APROVAÇÃO no rito de defesa.

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ATA Nº166

**ATA DE SESSÃO PÚBLICA DE DEFESA DE DOUTORADO PARA A OBTENÇÃO DO
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No dia seis de julho de dois mil e vinte e dois às 14:00 horas, na sala 03 (DEFESA FECHADA), Prédio de Engenharia de Bioprocessos e Biotecnologia, foram instaladas as atividades pertinentes ao rito de defesa de tese da doutoranda **FERNANDA GUILHERME DO PRADO**, intitulada: **New strategy to enhance the byosynthesis of bioactive compounds produced in Rhizopus fermented soybean: Recovery of bioactive compounds by conventional organic solvents, supercritical CO2 and evaluation of their potential activities**, sob orientação do Prof. Dr. CARLOS RICARDO SOCCOL. A Banca Examinadora, designada pelo Colegiado do Programa de Pós-Graduação ENGENHARIA DE BIOPROCESSOS E BIOTECNOLOGIA da Universidade Federal do Paraná, foi constituída pelos seguintes Membros: CARLOS RICARDO SOCCOL (UNIVERSIDADE FEDERAL DO PARANÁ), GUSTAVO HENRIQUE COUTO (UNIVERSIDADE TECNOLÓGICA FEDERAL DO PARANÁ), MARIA GIOVANA BINDER PAGNONCELLI (UNIVERSIDADE TECNOLÓGICA FEDERAL DO PARANÁ), LUIZ ALBERTO JUNIOR LETTI (UNIVERSIDADE FEDERAL DO PARANÁ). A presidência iniciou os ritos definidos pelo Colegiado do Programa e, após exarados os pareceres dos membros do comitê examinador e da respectiva contra argumentação, ocorreu a leitura do parecer final da banca examinadora, que decidiu pela APROVAÇÃO. Este resultado deverá ser homologado pelo Colegiado do programa, mediante o atendimento de todas as indicações e correções solicitadas pela banca dentro dos prazos regimentais definidos pelo programa. A outorga de título de doutora está condicionada ao atendimento de todos os requisitos e prazos determinados no regimento do Programa de Pós-Graduação. Nada mais havendo a tratar a presidência deu por encerrada a sessão, da qual eu, CARLOS RICARDO SOCCOL, lavrei a presente ata, que vai assinada por mim e pelos demais membros da Comissão Examinadora.

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Dedico essa tese à minha família,
em especial aos meus pais Irineu e Cleusa.

AGRADECIMENTOS

Sobretudo, a Deus. Aprendi que a vida é um sopro! Sou grata à Ele todos os dias por esse privilégio, **a vida**. Especialmente por Ele ter me concedido uma nova vida, um novo livro branco nas mãos para que eu possa escrever novas histórias. Agradeço por me abençoar todos os dias durante toda essa caminhada e por todos os obstáculos dispostos no caminho, foram eles que me moldaram e me fizeram mais forte.

Aos meus pais, Irineu e Cleusa, dedico muito mais que os meus agradecimento, dedico tudo o que sou e faço. Eu poderia escrever tantas páginas de agradecimento quanto os artigos que esculpem essa tese. Obrigada por tanto amor, por estarem sempre comigo, ao meu lado e por lutarem por mim. Obrigada por segurarem a minha mão, tão forte.. mas tão forte, que hoje eu estou aqui. Obrigada por todo o apoio que recebo, que me dá forças e coragem para alçar o vôo que for necessário. Todas as minhas conquistas e sucesso são e sempre serão dedicados e divididos com vocês e por vocês. Obrigada por serem os principais responsáveis por quem eu sou hoje.

Aos meus irmão, Giovani e Victória, ter vocês é a certeza de que eu nunca estarei sozinha. A distância é o que nos une. Obrigada pelas conversas diárias, pelas trocas de memes e piadas, estes que deixam o dia mais leve. Saibam que são a minha base, e eu estarei sempre aqui para vocês.

À minha amiga-irmã, Fabricia Lorrane, por partilhar comigo esses seis anos. Você faz parte da minha história, e se eu estou aqui hoje é porque um dia, quando menos esperávamos, você estava comigo. Obrigada por todos os momentos que vivemos, momentos estes que não serão jamais esquecidos. Foram seis anos que ocupam lindas páginas no meu livro da vida. Encerramos juntas esse ciclo e tenho certeza que os caminhos que tomamos serão repletos de luz. Estarei sempre aqui torcendo e vibrando o seu sucesso, você merece!

Ao Prof^o. Dr. Carlos Ricardo Soccol pela orientação. Por sua assistência e envolvimento dedicado em todas as etapas desse trabalho. Por todas as conversas, ideias, sugestões e troca de experiências que foram essenciais para o resultado final desse projeto. Por toda a compreensão.

À Prof^a. Dr. Maria Giovana Binder Pagnoncelli por sua co-orientação e amizade. Por se tornar a minha mãe de Curitiba. Obrigada por todos os ensinamentos,

por todos esses anos que estivemos juntas pesquisando, buscando, compartilhando momentos e oportunidades. Obrigada pelo incentivo, conversas e apoio nos momentos difíceis em que alguns resultados não vieram.

Ao Prof. Dr. Gilberto Vinícius de Melo Pereira, pelos conselhos, atenção e disposição em ajudar.

À Prof^a. Maria Rosa Machado Prado, pela colaboração.

Aos meus amigos da Usina Piloto e Cenbapar. Obrigada por todo o companheirismo e carinho, vocês estiveram comigo em todos os momentos. Obrigada pela energia que à mim emanavam no momento que eu mais precisei. Obrigada por me acolherem, por todas as trocas que tivemos. Até aqui viajamos juntos! Eu torço pelo sucesso de cada um de vocês e espero que as experiências compartilhadas neste percurso tornem-se a alavanca para nunca desistirmos e alcançarmos nossos objetivos e sonhos. E lembrem-se: vocês podem e vão muito além! Contem sempre comigo.

Ao Programa de Pós-Graduação de Engenharia de Bioprocessos e Biotecnologia da UFPR e aos professores e técnicos do Programa de Pós-graduação de Engenharia de Bioprocessos e Biotecnologia pelo auxílio e colaboração em diversos aspectos.

A CAPES, pelo suporte financeiro.

Ao conhecimento e crescimento que o Doutorado me proporcionou.

E por fim, desejo exprimir os meus agradecimentos a todos aqueles que de alguma forma, permitiram que esta tese se concretizasse.

Os meus mais sinceros agradecimentos!

“Não fui eu que lhe ordenei? Seja forte e corajoso! Não se apavore, nem se desanime, pois o Senhor, o seu Deus, estará com você por onde você andar.”

(Josué 1:9)

RESUMO

A busca por alternativas terapêuticas vem crescendo e ganhando destaque cada vez mais nos dias atuais. Uma das estratégias consiste no consumo de alimentos que são ricos em antioxidantes naturais e outros compostos bioativos que apresentam capacidade de prevenir ou reduzir processos patogênicos associados a doenças metabólicas. Devido à presença de muitos compostos com propriedades bioativas, incluindo antioxidantes, os alimentos fermentados são relacionados a uma série de efeitos benéficos para a saúde e surgem como uma alternativa promissora. A fermentação tem sido utilizada como uma forma potente de melhorar as propriedades da soja e dos seus componentes. Neste contexto, este estudo teve como objetivo em um primeira etapa avaliar o uso de diferentes substratos a fim de aumentar a produção de compostos bioativos através do bioprocessamento da soja por *Rhizopus oligosporus* NRRL 2710, e por seguinte comparar os rendimentos obtidos de diferentes métodos, nomeadamente Soxhlet e CO₂ + cosolvente supercrítico. A suplementação soja + arroz integral mostrou um aumento significativo na concentração de compostos fenólicos, expressa como equivalente á ácido gálico (GAE), (0,697 a 6,447 mg GAE g⁻¹) em comparação com o uso apenas da soja (1,792 a 3,10 mg GAE g⁻¹). Tal processo resultou em um produto rico em isoflavona e aglycone, contendo diferentes compostos antioxidantes, tais como ácido transcinâmico, ácido gálico, miricetina, quercetina e kaempferol. O substrato fermentado também mostrou grande potencial para inibir a enzima hialuronidase (atividade anti-inflamatória) e contra o crescimento de células tumorais CaCo-2 (atividade antitumoral). O Soxhlet foi utilizado como método de extração de referência para efeitos de comparação; foi realizado com 7 solventes orgânicos incluindo n-hexano, éter de petróleo, acetato de etilo, acetona, etanol, metanol e água. Para a extração de Soxhlet, os rendimentos mais altos e mais baixos obtidos foram 45,24% e 15,56% utilizando metanol e hexano, respectivamente. Além disso, foi alcançado um aumento de 18,62% no rendimento da extração quando a soja fermentada foi seca antes das extrações. A extração utilizando CO₂ supercrítico mais etanol como modificador estático (scCO₂ + EtOH) apresentou, a alta pressão (25 MPa) e temperatura (80°C), um teor de compostos fenólicos de 1391,9 µg GAE g⁻¹ e eliminação radical de 0,17 g g⁻¹, atingindo um rendimento de 42,87%. Todos os extractos mostraram actividade anti-inflamatória contra a enzima hialuronidase e os compostos bioactivos identificados por HPLC-DAD, incluindo ácido gálico, ácido transcinâmico, daidzeína e genisteína. Estes resultados mostraram que o produto obtido pode servir como um ingrediente de valor agregado para a industria alimenticia e farmaceutica e que as técnicas de extração utilizadas foram eficientes no rendimento do processo e os solventes de maior polaridade têm uma influência significativa. Foi também demonstrada a viabilidade técnica da scCO₂ + EtOH para obter um produto de alto valor agregado.

Palavras-chave: Soja fermentada. Atividade antioxidante. Compostos fenólicos. Extração por solventes orgânicos. Extração por fluidos supercríticos. Anti-inflamatório. Antitumoral.

ABSTRACT

The search for therapeutic alternatives has been growing and gaining prominence more and more these days. One of the strategies is the consumption of foods that are rich in natural antioxidants and other bioactive compounds that have the capacity to prevent or reduce pathogenic processes. Due to the presence of many compounds with bioactive properties, including antioxidants, fermented foods are related to a number of beneficial health effects and emerge as a promising alternative. Fermentation has been used as a potent way of improving the properties of soybean and their components. In this context, this study aimed in a first step to evaluate the use of different substrates in order to increase the production of bioactive compounds through the bioprocessing of soybean by *Rhizopus oligosporus* NRRL 2710, and next to compare the yields obtained from different methods, namely Soxhlet and CO₂ + supercritical solvent. The soy + brown rice supplementation showed a significant increase in the concentration of phenolic compounds, expressed as gallic acid equivalent (GAE), (0.697 to 6.447 mg GAE g⁻¹) compared to using soy alone (1.792 to 3.10 mg GAE g⁻¹). Such a process resulted in a product rich in isoflavone and aglycone, containing different antioxidant compounds such as trans-cinnamic acid, gallic acid, myricetin, quercetin and kaempferol. Soxhlet was used as a reference extraction method for comparison purposes; it was performed with 7 organic solvents including n-hexane, petroleum ether, ethyl acetate, acetone, ethanol, methanol, and water. For Soxhlet extraction, the highest and lowest yields obtained were 45.24% and 15.56% using methanol and hexane, respectively. In addition, an increase of 18.62% in extraction yield was achieved when fermented soybeans were dried before extractions. Extraction using supercritical CO₂ plus ethanol as a static modifier (scCO₂ + EtOH) showed, at high pressure (25 MPa) and temperature (80°C), a phenolic compound content of 1391.9 µg GAE g⁻¹ and radical elimination of 0.17 g g⁻¹, achieving a yield of 42.87%. All extracts showed anti-inflammatory activity against hyaluronidase enzyme and bioactive compounds identified by HPLC-DAD, including gallic acid, trans-cinnamic acid, daidzein and genistein. These results showed that the product obtained can serve as a value-added ingredient for the food and pharmaceutical industries and that the extraction techniques used were efficient in yielding the process and the higher polarity solvents have a significant influence. The technical feasibility of scCO₂ + EtOH to obtain a high value-added product was also demonstrated.

Keywords: Fermented soybean. Antioxidant activity. Phenolic compounds. Extraction by organic solvents. Supercritical fluid extraction. Anti-inflammatory. Antitumoral.

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1 INTRODUCTION

Soybean (*Glycine max L.*) and its derivatives are present in the human diet all over the world, being one of the cheapest and most abundant sources of vegetable protein consumed. It is one of the world's main *commodities* and accounts for 60% of global oilseed production. World production in the 2021/2022 harvest was estimated by the United States Department of Agriculture at 385.5 million tons. It is projected an increase by 21.5 million tons over the past crop (CONAB, 2022; USDA, 2022a; ZHOU *et al.*, 2022).

Soy is thought to be the most abundant source of phenolic compounds in the human diet, being an important class of phytochemical antioxidants and have been widely used by the food and pharmaceutical industries (LI *et al.*, 2020; WANG *et al.*, 2016). In the human diet soy is the richest source of isoflavones (phytoestrogenic compounds with estrogenic activity), and these are divided into two basic phytochemical categories: glycosides and aglycones (JASKULSKI *et al.*, 2017; LI *et al.*, 2017; MOORE; FRANKS; FOX, 2017).

Fermented soy products have phenolic compounds available in the aglycone form and consequently have higher bioactive activities than "*in natura*" soybeans. This because aglycone forms are characterized by being more quickly absorbed by the organism and have higher estrogenic and antioxidant potential (KAYANO *et al.*, 2012; PRADO *et al.*, 2021).

Several studies in the literature mention fermented soy products and derived bioactive compounds, however, information that present the comparative effects of extraction methods and operating conditions is vague (PRADO *et al.*, 2021; SANJA *et al.*, 2018). Traditional extraction methods have been used to recover many bioactive compounds from plant materials. However, these methods require large amounts of

organic solvents, which is not interesting because they end up becoming costly and environmentally harmful processes, resulting from the use of aggressive and toxic chemicals (ALESSANDRA *et al.*, 2019).

Bioactive components of interest, such as phenolic compounds, carotenoids, alpha-acids, methylxanthines, and vitamins are present in plant sources. Phenolics consist of flavonoids, isoflavones, tannins, and other related ones (ALVAREZ *et al.*, 2019; GRAJEDA-IGLESIAS *et al.*, 2016). In addition to the high protein content, other bioactive and remarkably rich components such as isoflavones, anthocyanins, and saponins are present in soy. These compounds are known to have antioxidant capacity that is related to different health benefits (GIORDANO *et al.*, 2015; MANI; MING, 2017).

Antioxidants are among the main health components that consumers look for in food products because of their benefits in preventing many diseases caused by cellular oxidative processes and reactive oxygen species (ROS), such as aging and chronic diseases like diabetes and cancer (Prior *et al.*, 1998; Melo *et al.*, 2006; Mukhopadhyay, 2006).

In the context, methods that encompass the principles of "green chemistry" are an alternative that reduces or eliminate extractive waste (ALVAREZ *et al.*, 2019). Supercritical carbon dioxide (scCO₂) appears as a modern method for the extraction of natural compounds with high added value (Attard *et al.*, 2018). A considerable advantage of this method is the manipulation of temperature and pressure that adjust the properties of supercritical fluids, such as density, with the aim of increasing the selectivity of the solvent (AHMED *et al.*, 2017).

The use of the co-solvent is a useful strategy to enhance the solvent power, increase the extraction of polyphenols, and improve the selectivity regarding the extraction yield of compounds from plant matrices (Santos *et al.*, 2017). This is

because the solubility of long-chain molecules and high polar compounds in supercritical CO₂ is limited and this disadvantage can be solved using a polar co-solvent (SOLTANI; MAZLOUMI, 2017).

There are few reports on extractions of fermented soybeans by filamentous fungi and evaluation of the biological activities of the extracts obtained from the resulting material. Therefore, this study evaluated the extracts obtained by different extraction techniques - solid-liquid, Soxhlet, with different organic solvents, and extraction with CO₂ + supercritical ethanol (scCO₂ + EtOH). The antioxidant activity (AA), total phenolic content (TPC), phenolic compounds (by HPLC), fatty acid profile (FA), β -glucosidase activity, anti-inflammatory activity, antitumor activity against CaCo-2 and MCF-10 cells and oxidative stability were evaluated in order to verify the technical feasibility of the process and the recovery of molecules with high bioactive potential for the pharmaceutical and food industries.

2 OBJECTIVES

2.1 GENERAL OBJECTIVES

This work aimed at the production and quantification of bioactive compounds by fermentative process from soybean fermented by filamentous fungi and evaluation of biological activities, as well as evaluating the fermented product and potential for use in pharmaceutical and food industries.

2.2 SPECIFIC OBJECTIVES

- To verify the increase in antioxidant activity of the fermented after soybean supplementation with brown rice
- To evaluate the concentration of total phenolic compounds during the fermentation process kinetics

- To evaluate the changes in isoflavone profiles (genistein and daidzein) during fermentation
- Evaluate the increased anti-inflammatory and antitumor potential after the fermentation process
- Evaluate the different yields of Soxhlet extraction using organic solvents with different polarities
- Verify the influence of the drying process on the extraction yields in both extraction methods used, Soxhlet and scCO₂ + EtOH
- Check the extraction efficiency using scCO₂ + EtOH at different pressure and temperature

3 LITERATURE REVIEW

3.1 SOYBEAN

Soybean (*Glycine max*) is one of the most widely grown oilseed crops in the world and one of the cheapest and most abundant sources of vegetable protein consumed as a food and dietary supplement (JIA *et al.*, 2020). The soybean has in its composition about 40% protein, 20% lipids, 35% carbohydrates, 5% minerals, and 10% moisture, in addition to other compounds such as fatty acids, vitamins, flavonoids, isoflavones, phenolic acids, and saponins (MEDIC; ATKINSON; HURBURGH, 2014; REDDY; DUKE, 2015).

In the context between global supply and demand for food, soy stands out for being a highly nutritious grain capable of providing a range of by-products for human consumption, in addition to attributing an important role in the production of animal protein (chicken, pork, and beef) (DE LIMA *et al.*, 2018). Of the world oilseed production, 60% corresponds to soy and, of this total, only 6% is consumed in the form of grains, whole grains, and fermented products. The remaining 94% is processed industrially, being transformed into oil for human consumption, production of biodiesel,

and development of chemicals, food, and cosmetics (18-20% of this total). The remainder crushed is usually transformed into protein-rich bran used for animal feed (PESSOA *et al.*, 2019).

The world soybean production in the 2019/2020 harvest was estimated by the U.S. Department of Agriculture (USDA) at 337.298 million tons, in a total world area occupied by the cultivation of 122.647 million hectares, corresponding to the average productivity of 2.750 kg/ha (United States Department of Agriculture USDA, 2022b). The largest suppliers are Brazil, the United States and Argentina (Table 1) (CONAB, 2022; United States Department of Agriculture USDA, 2022b).

TABLE 1. PRODUCTION, PRODUCTIVENESS AND TOTAL PLANTED AREA IN HECTARES OF SOYBEAN IN 2019/2020.

	Area (million ha)	Production (million ton)	Productiveness (kg/ha)
Brazil	36.95	124.844	3.379
United States	30.33	96.676	3.187
Argentina	17.1	50.0	2.92
World	122.647	337.298	2.750

SOURCE: (CONAB, 2022; United States Department of Agriculture USDA, 2022b)

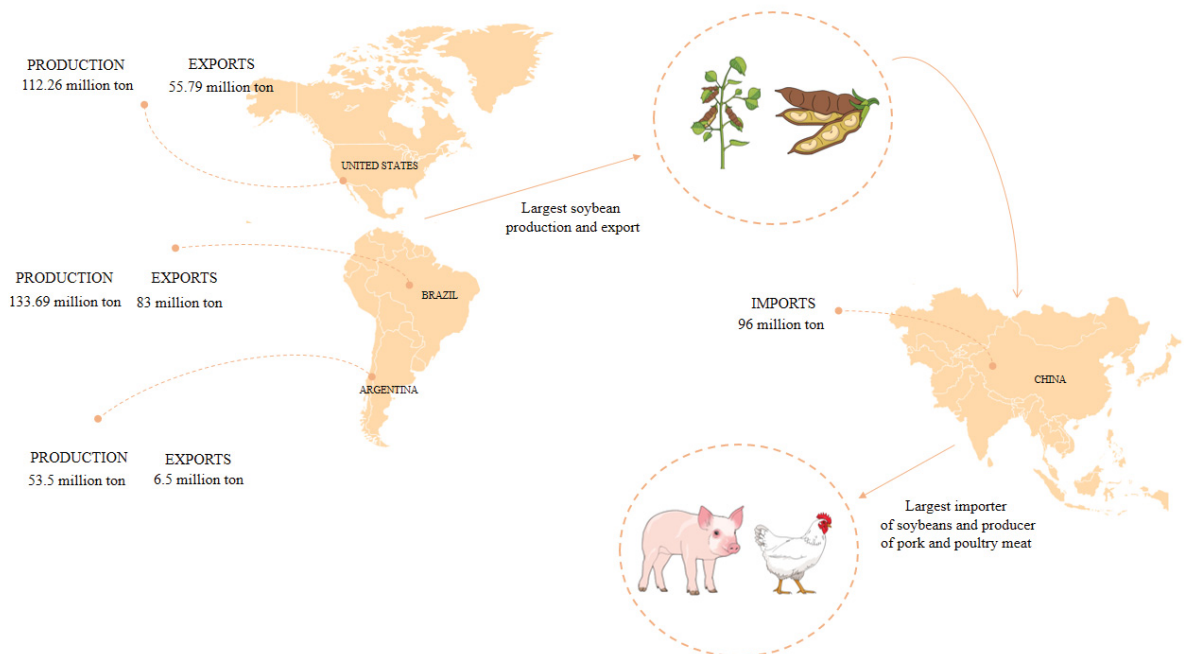
USDA projects the 2021/22 world soybean crop at 372.56 million tons. Brazil remains the world's largest producer of soy, however the forecast was cut from 144 million, in the previous bulletin, to 139 million tons. Brazilian ending stocks of soy were also reduced from 28.25 to 23.55 million tons but estimates for Brazilian soy exports were maintained at 94 million tons. Argentina's production was also projected to fall by 46.5 million tons, a cut of 3 million from the previous report. For the US, the USDA brought a slight increase from 120.45 to 120.7 million tons (USDA, 2022a).

Asian countries are characterized by high grain consumption. In recent years, due to increased demand and domestic supply, China has become the largest importer of soybeans, accounting for more than 60% of world soy trading. This is because Chinese production accounts for only 20% of domestic demand and a large amount of

soy is needed to supply pork feed. Compared to other crops, the application of soy resources is potentially greater, taking the efficiency of land use as follows: a soybean yield of 3000 kg/ha becomes a product to produce 343 kg of beef, 600 kg of swine, or 1200 kg of birds (GARRETT; RAUSCH, 2016; WU *et al.*, 2020).

In the 2020/2021 harvest, there is an estimate for Chinese imports of 96 million tons. Brazil also assumes the position of largest grain exporter, with more than 60% of the harvest destined for this purpose. Motivated by the high Chinese demand and the percentage, Brazilian exports are expected to reach 83 million tons. The global export forecast is 312.80 million tons, of which 55.8 million tons are from the United States and only 6.5 million tons from Argentina (Figure 2) (United States Department of AgricultureUSDA, 2022b).

FIGURE 1 - SOYBEAN PRODUCTION AND EXPORT OF LARGEST PRODUCERS AND ESTIMATE FOR IMPORTS IN THE 2020/2021 HARVEST.



SOURCE: The author (2022).

Plant genotype, location, climate, water, and maturity group are responsible for the quality characteristics of soy, e.g., protein, oil, fatty acids, soluble sugars, and isoflavones. The grain consists of 8% husks, 90% cotyledons, and 2% hypocotyls. The protein composition of the soybean grain on a dry basis is on average 40% (Table 2). The storage of proteins occurs in the intermediate layers of the grain, cotyledons, and hypocotyls. In the shell, the outer layer, there are greater amounts of carbohydrates and cellulosic material (ASSEFA *et al.*, 2019; PERKINS, 1995).

TABLE 1 - NUTRITIONAL COMPOSITION OF SOYBEAN

Components	% grain	Chemical Composition (% dry weight)			
		Proteins	Lipids	Carbohydrates	Others
Husks	8	9	1	86	4.3
Hypocotyls	2	41	11	43	4.4
Cotyledons	90	43	23	29	5.0
Total	100	40	20	35	5.0

* Minerals, vitamins, phytates and isoflavones.

SOURCE: The author (2022).

In addition to the high protein content, the soybean contains other bioactive and remarkably rich components, such as isoflavones, anthocyanins, and saponins. They are compounds known to have antioxidant capacity that is related to different health benefits (GIORDANO *et al.*, 2015; MANI; MING, 2017).

3.2 NUTRITIONAL CHANGES IN FERMENTED SOYBEAN PRODUCTS

Fermentation enriches the nutritional value of foods by increasing the content of vitamins, essential amino acids, or fatty acids, allowing detoxification and removal of anti-nutritional factors. In addition to proteins and isoflavones, soybean is made up of numerous other functional and nutritional substances, increased in the fermentation

process through microbial biotransformations Microorganisms with abilities to produce specific hydrolytic enzymes, such as protease, amylase, and β -glucosidase, play a fundamental role in increasing functional properties (KUMAR *et al.*, 2016; XU; DU; XU, 2015; ZHENG *et al.*, 2017).

The modification of isoflavones occurs through β -glucose enzymes, which degrade cellulose, hydrolyze the β -D-cellulose terminal non-reducing glucoside bond, and, consequently, release β -D-glucose. Thus, the amount of glycosides decreases by hydration by β -glucosidase, increasing the amount of isoflavones aglycones (FLORINDO *et al.*, 2018; GUADAMURO *et al.*, 2017). After ingesting the aglycone isoflavones, such as genistein and daidzein, they are absorbed by the blood vessels, hydrolyzed in the small and large intestine by intestinal hydrolytic enzymes and microbial glycosidases through deglycosylation, increasing their bioactive potential (RAIMONDI *et al.*, 2009).

Some final characteristics of the fermented product, as well as the changes that occur during soy fermentation, are related to the type of microorganism used. In tempeh, for example, several amylases, lipases, and proteases are produced by fungi of the genus *Rhizopus spp.* These hydrolyze macronutrients into simpler, water-soluble compounds, resulting in the production of vitamins, phytochemicals, and antioxidant constituents (AHMAD *et al.*, 2015). The increased antioxidant effect of tempeh can still be attributed to the increased levels of polyphenols released by cell wall degradation by enzymes secreted by *Rhizopus* fungi during preparation in the boiling stage and through the course of fermentation (KULIGOWSKI; PAWŁOWSKA; JASIŃSKA-KULIGOWSKA, 2017).

Due to the metabolic activity of starter cultures, the levels of vitamin B complexes are also increased during fermentation. In tempeh *Rhizopus* and the

bacteria, *K. pneumoniae* and *C. freundii* are the main producers of vitamin B₁₂. In natto, *Bacillus* is the agent responsible for the increase of vitamin K₂ (KAMAO *et al.*, 2007; MO *et al.*, 2013). The functionality of these vitamins is well known, being essential bioactive substances that act in the coordination of the nervous system and the development of the brain, some studies, still confirm the significant increase in gamma-aminobutyric acid (GABA) in fermented soy products, responsible for the regulation of the central nervous system (XU; CAI; XU, 2017).

Microbial proteolytic enzymes involved in fermentation processes hydrolyze the protein content into peptides. The length of the chain and the composition and sequence of amino acids interfere with the biological activity of the peptide and, during enzymatic hydrolysis in fermentation and digestion, inactive bioactive peptides are released. Furthermore, the bioconversion of high molecular weight proteins into minors increases the solubility (CHAN; LIU, 2016; YOUNG *et al.*, 2011). One of the main biochemical changes that occur during fermentation is the hydrolysis of proteins by microbial proteases and the enrichment of nutritional effects depends on this reaction (MEINLSCHMIDT *et al.*, 2015).

Angiotensin-converting enzyme (ACE) inhibitory peptides are generated by proteolytic degradation of glycinin and β -conglycinin, which consists of protein fractions from soybean. This enzyme acts in the conversion of angiotensin I into angiotensin II and inactivation of the bradykinin vasodilator, raising blood pressure and the risk of cardiovascular disease. Hydrophobic amino acids (Try, Phe, Trp, Ala, Ile, Val, and Met) or positively charged amino acids (Arg and Lys) show greater affinity with ACE. There are three classifications for ACE-inhibiting peptides, being (1) true inhibitor, unaffected by gastrointestinal digestion, (2) substrate, converted into other peptides with less activity in gastrointestinal digestion, and (3) prodrug, converted to true inhibitors by

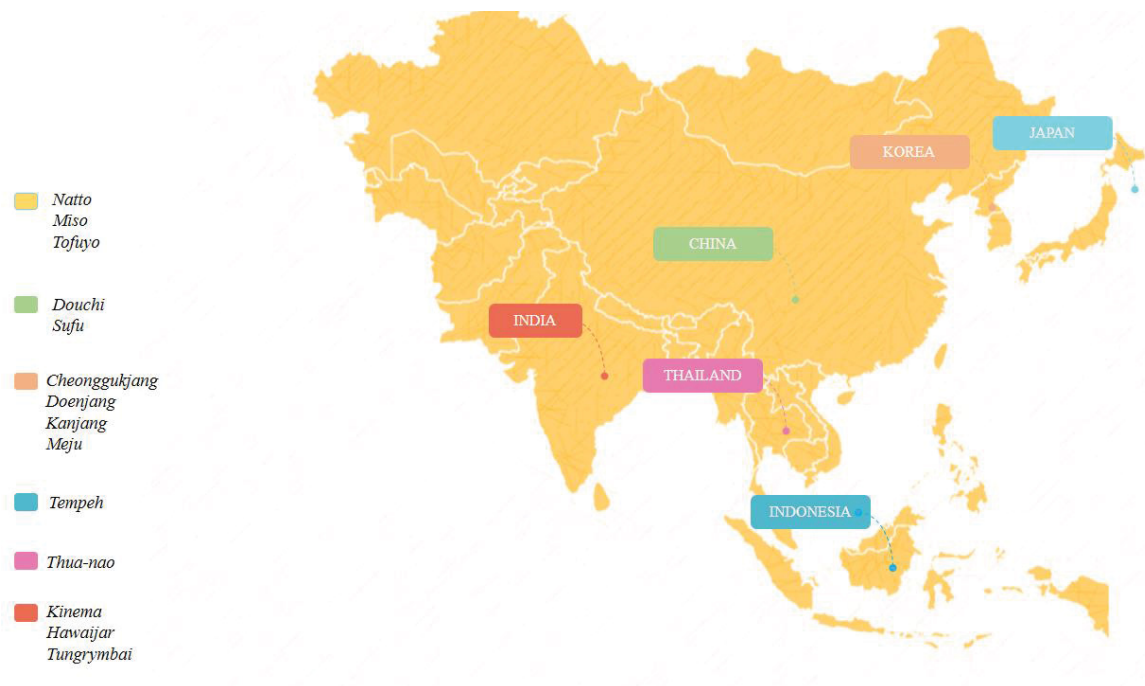
gastrointestinal digestion (HE *et al.*, 2012; RAI; SANJUKTA; JEYARAM, 2017). Still, several other nutritional changes are reported as a consequence of the soybean fermentation process, such as the increase in total soluble iron, the level of folic acid, the composition of tocopherol, with the levels of beta-, gamma- and delta-tocopherol being increased (MANI; MING, 2017).

In summary, many metabolic activities and biotransformations occurred during the soybean fermentation process. There are several beneficial health effects of the final fermented product, and its consumption is related to a series of bioactivities that will be mentioned below.

3.3 FERMENTED SOYBEAN PRODUCTS

The production and consumption of fermented soy are widespread in Asian countries. The main soybean products include *natto*, *miso*, *tofuyo* (Japan), *douchi*, *sufu* (China), *cheonggukjang*, *doenjang*, *kanjang*, and *meju* (Korea), *tempeh* (Indonesia), *thua-nao* (Thailand), *kinema*, *hawaijar*, *tungrymbai* (India) (Figure 3). In addition to other widely consumed products, such as sauce, pasta, and soybean milk (Production of bioactive peptides during soybean fermentation and their potential health benefits SANJUKTA; RAI, 2016). Records of some production methods were found in *Cheminyoshul*, a Chinese manuscript dating back to 530-550 dC, and others in the Korean manuscript *Samkuksaki*, dating from the 1392s, pointing to the consumption of fermented soybeans since the 12th century (MAH, 2015).

FIGURE 2 - SOY FERMENTED PRODUCTS IN ASIAN COUNTRIES



SOURCE: The author (2022).

The large consumption of soybeans in Asian countries is related to the widespread adoption of grain cultivation. The variety of appropriate climates and geographic regions resulted in highly sizable crops, making soybean a staple in the region. With insufficient meat consumption, these fermented foods played a vital role as a source of protein in the Asian diet (PARK; LEE; MAH, 2019; WU *et al.*, 2004).

In ancient times, the basic idea behind soybean fermentation was the preservation of food. In the current perspective, the research is interested and directed to the application of fermentation to improve the bioactive components of soybean, responsible for health benefits, and reduce anti-nutritional factors (DIFO *et al.*, 2014; RAI; KUMARASWAMY, 2015).

The difference between fermented soybeans is based on several parameters, but mainly due to the microorganism used in the process. Thus, fermented soybean products are different in terms of aroma, texture, and therapeutic and nutraceutical

values. Some fermentations occur only with bacteria, others using only filamentous fungi, and, in many cases, both these microbial groups are used in the fermentation. Some products are fermented only with *Bacillus* (*natto*, *kinema*, *chungkookjang*); some are fermented with fungi *Aspergillus oryzae*, *Mucor* spp. *Rhizopus* spp. and *Fusarium* spp. (*douchi*, *tempeh*, *miso*, *tofu*) and in some cases both microorganisms are used, as in the case of *doenjang*, where the bacteria involved in this process would be *B. subtilis* and fungi include *Rhizopus* spp., *Mucor* spp., *Geotrichum* spp., and *Aspergillus* spp. (CAO *et al.*, 2009; JUNG; LEE; JEON, 2014; SANJUKTA; RAI, 2016; STEINKRAUS *et al.*, 1959; YASUDA, 2011).

Soybean protein and isoflavones are the main functional constituents of fermented soybean foods. Soybean consists of one of the plant sources with the highest abundance of isoflavones. Because their chemical structure is similar and has an affinity for estrogen receptors, these compounds are usually called phytoestrogens (DAI *et al.*, 2019; ZHU *et al.*, 2020).

The native forms of isoflavones have their bioavailability compromised because they are usually combined with sugars that minimize their absorption through the human intestinal tract. Isoflavones are categorized into two groups: glycosides and aglycone. The beneficial and functional effects of isoflavones on health are conferred to their aglycone forms, which are absorbed more quickly by the body. In unfermented soybean, the presence of aglycone isoflavones is 2–3% of the total composition, this content being mainly corresponded to β -glucoside isoflavones (CAO *et al.*, 2019; HANDA *et al.*, 2019; ZHANG; YANG; HUANG, 2019).

Thus, the biotransformation of glycosidic forms into aglycones through fermentation is a desirable process to increase and produce more biologically active forms. In fermented soy products, the aglycone values vary from 40 to 100%. The

conversion of the glycoside into isoflavone aglycones occurs through the action of β -glycosidase produced by microorganisms during the fermentation process. In addition to a higher absorption rate, aglycone forms have greater antioxidant activity than glycosidic forms. This explains the fact that the consumption of fermented soybean products in Asian countries is associated with the reduction of chronic diseases since the consumption of natural antioxidants is efficient in reducing the harmful impacts of reactive oxygen species (ROS) and in adjusting the body's antioxidant load (CHEN, M. *et al.*, 2014; PRADO *et al.*, 2021; XIAO *et al.*, 2015).

Soybean proteins, they have many enzyme inhibitors, such as proteinase and trypsin inhibitors, which make them less digestible. During fermentation, proteolytic enzymes generated by microbial populations hydrolyze proteins into peptides and free amino acids responsible for antioxidant activity and increasing the digestibility of soybean protein (MAI; TECHNOLOGY; RAI, 2011; SANJUKTA *et al.*, 2015).

In addition to proteins and isoflavones, soybean is made up of numerous other functional and nutritional substances, such as fatty acids, vitamins, peptides, minerals, flavonoids, phenolic acids, and saponins (MAN *et al.*, 2011).

Natto consists of popular and traditional food in Japan that has been consumed since the 17th century and produced by fermenting soybeans cooked with strains of *Bacillus subtilis* var natto. It is known to have large amounts of peptides because, in the fermentation process, proteins are cleaved by extracellular proteases produced by the *Bacillus* strain, which increases the free amino acid content by 10% to 30%. Studies report that the proteins derived from this food consist of at least seventeen different amino acids, including glutamic acid, glutamine, aspartic acid, leucine, proline, serine, lysine, methionine, threonine, glycine, isoleucine, tyrosine, phenylalanine, histidine, arginine, alanine and valine (MANABE, 2011; SATO *et al.*, 2018). The consumption of

natto has been shown to have an anti-aging effect, prolonging life expectancy, due to the metabolites found in natto extracts, for example, the enzyme nattokinase. Often, reduced life expectancy is caused by oxidative stress, and the relatively high antioxidant activities of natto are mentioned (INOMATA; MIYAKAWA; AIHARA, 2018).

Miso is a fermented soybean paste, prepared from steamed soy, salt, and koji - cooked cereal or soy malted with *Aspergillus oryzae*. It is a traditional Japanese spice used to add flavor to soups and dishes consumed at breakfast by most Japanese families for over 1000 years. The process of maturation of the Miso takes from three to twenty-four months and involves several microorganisms, such as molds, yeasts and, lactic acid bacteria, which act by hydrolyzing the components of soybean (INOUE *et al.*, 2016; WANG, S. *et al.*, 2019). There are different types of miso, which vary according to local traditions and available ingredients; and this food can be classified according to the koji used: 1) rice miso, made by adding rice koji to soy; 2) barley miso with the addition of barley koji to soy and soy miso, made only with soybean. It is reported that during the miso ripening process, the peptides formed are made up of 3-20 amino acids and still include amino acids such as glutamic acid, aspartic acid, and proline (OGASAWARA; YAMADA; EGI, 2006; RATNANINGRUM *et al.*, 2018).

Tofu is a Japanese fermented soy curd similar to cream cheese a characteristic that results from the ripening or maturation process by proteases, carbohydrases and other catabolic enzymes found in red koji (*Monascus fungus*) or koji yellow (*Aspergillus oryzae*), used in the preparation of this food together with tofu (vegetable cheese based on soybean) (NURYANI *et al.*, 2018; YASUDA, 2011). Its functional properties were investigated and associated with the presence of bioactive peptides. Thus, tofu came to be seen not only as a nutritional accompaniment but

becoming a valuable source of protein (KUBA *et al.*, 2003; YASUDA; TACHIBANA; KUBA-MIYARA, 2012).

Douchi is a popular product consumed for at least 2.000 years by the Chinese, as a source of protein and flavoring ingredient. The preparation of the douchi is carried out in two stages: pre-fermentation, which consists of an aerobic process using several microorganisms as the starter culture (for example, *Aspergillus oryzae*, *Zygosaccharomyces rouxii*, *Lactobacillus plantarum* e *Bacillus subtilis*) and takes 12-15 days; and post-fermentation, where the addition and mixing of salt and other spices is carried out and left for 9 months in anaerobic fermentation, a process where the development of the special nutrients and flavor of the douchi occurs (CHEN, T. *et al.*, 2014; HE *et al.*, 2016). In recent years, douchi has attracted attention as a functional food source. Some studies have revealed the benefits of this food to health, including antioxidants, antihypertensive activity and even lowering blood pressure (TAN *et al.*, 2019; WANG *et al.*, 2008).

Mentioned as “Chinese cheese” due to its texture, sufu is a traditional fermented soy product that has been used as a flavor enhancer and appetizer. There are different types of sufu, which are produced by various processes in different locations in China through microbial fermentation, and based on the types of starter culture; the sufu can be classified into fungi fermented sufu (inoculated with *Actinomucor*, *Mucor*, or *Rhizopus*), sufu fermented by bacteria (inoculated with *Bacillus* or *Micrococcus*) and others (naturally inoculated) (HUANG *et al.*, 2018; YANG; TAN; KAN, 2021).

3.4 POTENTIAL BENEFICIAL HEALTH EFFECTS OF SOYBEAN FERMENTED PRODUCT

3.4.1 Antioxidant effect

Many normal reactions in the body form by-products such as free radicals, which are species with unpaired electrons. If the antioxidant defense systems are not efficient, there is an increase in tissue damage and oxidative stress, associated with cell apoptosis and the appearance of several chronic diseases (SANJUKTA *et al.*, 2015).

Due to the beneficial effects of the prevention of these diseases caused by cellular oxidative processes and reactive oxygen species, antioxidants, essential to prevent the formation and suppress the activities of reactive nitrogen and oxygen species, become the main compounds with benefits for health to be included in the diet (DASTMALCHI *et al.*, 2020).

Several fermented soybean foods have bioactive components, such as polyphenols, phenolic acid, saponins, sterol, and flavonoids, that protect against oxidative damage, with flavonoids and phenolics being fundamental compounds responsible for antioxidant activity (SHUKLA *et al.*, 2016).

Phytoestrogenic compounds and phenolic compounds are an important class of phytochemical antioxidants. In soybean and soybean products, aglycone forms are characterized by having greater estrogenic and antioxidant potential (WANG, D. *et al.*, 2019).

3.4.2 Anticancer effect

Environmental factors, especially diet, are considered to play a key role in carcinogenesis. The incidence of cancer in the Asian population is relatively low, and Asians traditionally consume large amounts of soy-based foods, which are rich in isoflavones (KIMURA, 2012). One of the first studies that linked cancer risk reduction and the consumption of fermented soy-based foods took place in Singapore in 1991, reporting that a soy-rich diet resulted in less breast cancer development in women in pre-menopause (LEE *et al.*, 1991).

Soybean isoflavones are believed to have the potential to reduce cancer risk through their antioxidant activity and estrogen-like structure. Genistein presents Estrogen binding affinity compared to estradiol (estrogen steroid hormone) receptors ER- α and ER- β of 4% and 87%, respectively. As a result, it binds to these receptors and plays an important role in preventing hormone-related cancers (KHOSRAVI; RAZAVI, 2021). In addition, genistein is a known tyrosine kinase inhibitor and acts by preventing topoisomerase and angiogenesis. Through these functions, their effect is evident in the cascades of proliferation signals. Some bioactive peptides from soybean can also prevent the growth of tumor cells, such as lunasin and saponins, repelling the formation of the cell membrane and promoting cell apoptosis (CALVELLO *et al.*, 2016; SINGH; YADAV; VIJ, 2019).

Several soybean products are mentioned for their anti-cancer potential. In Korea, fungi and *Bacillus* sp. are used in the fermentation process in Meju, a dry soybean block, which is used to produce other products such as Kanjang, Doenjang, and Gochuchang. The anti-cancer potential of Doenjang is associated with compounds, such as trypsin inhibitor, isoflavones, vitamin E and an unsaturated fatty acid that contributes to the biological effect. Still, Doenjang extracts invigorate

glutathione S-transferase and increase the vitalization of natural killer cells (SHIN; JEONG, 2015).

Mostly, the anticancer effects of fermented soy products are associated with isoflavones. Some studies have shown that methylation-mediated epigenetic gene silencing can be reversed. Genistein is said to have a broad-spectrum anticancer effect on cancers of the breast, prostate, esophagus, pancreas, stomach, and colon, and metacarcinoma, lymphoma, and neuroblastoma. It also acts as a positive regulator of the mRNA expression of several tumor suppressor genes, counteracting the function of growth-stimulating factors and inhibiting cell malignancy. Therefore, the consumption and inclusion of fermented soybean foods have stood out as a new therapy for the treatment of tumors (BILIR *et al.*, 2017; SUNDARAM *et al.*, 2018).

3.4.3 Anti-obesity and Antidiabetic effect

Physiologically, obesity consists of an imbalance between energy intake and consumption that indicates an excessive accumulation of fat in the tissue and is considered a major health problem that is advancing significantly worldwide. Obesity is generally correlated with diabetes and metabolic syndromes leading to hyperinsulinemia and dyslipidemia (HSIAO; HO; PAN, 2020).

As a result, interest in combating obesity and overweight is growing. Several studies show that a change in the diet prevents and alleviates a series of metabolic imbalances characterized by central obesity, dyslipidemia, and high fasting glucose. Some benefits are attributed to the physiologically active components of certain foods, therefore, they are used to prevent obesity and its complications (ROSAS-VILLEGAS *et al.*, 2017; WANG *et al.*, 2017).

The isoflavones daidzein and genistein found in high levels in fermented soybean foods are mentioned as having bioactivity, regulating the generation of lipids and thermogenesis *in vivo*. Through lipogenesis (synthesis of fatty acids and triglycerides), hyperlipidemia (high levels of fat particles in the blood), hyperglycemia (elevated blood glucose), and improved insulin resistance, aglycones and metabolites demonstrate their anti-obesity effects (GAMAN; STOIAN; ATANASIU, 2011). Some studies also associate these effects with the bioactive phytochemical content of fermented soybean foods as alpha-amylase and alpha-glucosidase inhibitors, protease inhibitors, hemagglutinin, and crude fibers, able to disturb normal metabolism and assist in the management of obesity and different metabolic disorders (SHARMA; BALUJA; PROFESSOR, 2015)

The consumption of isoflavones through soybean foods in the diet in HDL cholesterol and reduction of total cholesterol, LDL, and triglycerides. In addition to isoflavones, soy proteins, as well as peptides, are active ingredients that lower the levels of LDL cholesterol and triacylglycerols (SINGH; YADAV; VIJ, 2019).

There is an association between obesity and the transition from pre-menopause to post-menopause in women. This phase is correlated with the risk of several diseases due to the lack of hormonal regulation, including the accumulation of abdominal fat, hypertriglyceridemia, and high levels of low-density lipoprotein cholesterol (LDL-C), reduced high-density lipoprotein cholesterol (HDL-C), elevated blood pressure (BP) and impaired glucose tolerance/diabetes. Studies show that the consumption of fermented soybean foods rich in isoflavones has beneficial effects on the distribution of body fat and lipid profile in women during the menopause period, due to the structural similarity of these compounds with estrogen, their greater affinity

for estrogen receptors and circulating concentration in the human body (HUANG *et al.*, 2016; SQUADRITO *et al.*, 2013).

Clinical and experimental studies indicate that the population with obesity and overweight is more vulnerable to type 2 diabetes mellitus (DM2), with obesity dramatically increasing the likelihood of DM2. Individuals with type 2 diabetes mellitus are at increased risk of cardiovascular disease, even with aggressive control of glucose, cholesterol, and blood pressure (GUO; LING, 2015; VIEIRA *et al.*, 2017).

Oxidative stress (OS) is closely associated with obesity and diabetes. Free fatty acids, which at high levels influence the production of reactive oxygen species (ROS) through mitochondrial electron transport chain complexes and enzymes in endothelial cells, decrease the bioactivity of nitric oxide, activate pro-inflammatory signaling pathways causing damage to cellular proteins and organelles. Damaged, more oxidizing mitochondrial enzymes enhance oxidative stress and cellular dysfunction. Chronic exposure to ROS negatively affects insulin signaling when stress pathways are activated. As a result, insulin resistance, glucose intolerance, β -cell, and mitochondrial dysfunctions are developed, advancing to a state of diabetes (PICARD *et al.*, 2013; RAINS; JAIN, 2011).

The consumption of foods rich in isoflavones is seen as a promising strategy in the treatment of diabetes and obesity. Genistein reduces the inflammatory state in obese people, decreases production, and neutralizes the effects of ROS, resulting in the relief of insulin resistance and, consequently, decreasing the risk of diabetes (BEHLOUL; WU, 2013). After ingestion, genistein enriches insulin resistance by increasing the production of insulin receptor substrate (IRS) 1, glucose transporter (GLUT) type 1, and N-terminal c-jun kinase, increasing the activity of superoxide dismutase, decreasing mitochondrial damage and lipid peroxidation. Daidzein, which

is also found in soybeans, activates GLUT4 and IRS1 in adipocytes, adding insulin-stimulated glucose uptake (LEI *et al.*, 2011).

Spermidine is a naturally occurring polyamine present in all living cells noted to play an important role in cellular functions and is found in different concentrations in fermented soy foods such as *Chunjang* (1.4 –12.8 mg/kg), *Doubanjiang* (0.18 mg/kg), *Douchi* (74.92 mg/kg) and *Sufu* (1.3 – 32.87 mg/kg) (CHOKSOMNGAM *et al.*, 2021; PARK; LEE; MAH, 2019).

Increased spermidine flux is associated with increased glucose and lipid metabolism. Many *in vivo* studies reveal that spermidine overexpression protects against diet-induced obesity, and an epidemiological study shows that foods rich in polyamines, such as spermidine, are associated with a lower occurrence of cardiovascular disease (CVD), corroborating the theory that spermidine is beneficial in the treatment of obesity (EISENBERG *et al.*, 2009; KRAUS *et al.*, 2014; SODA; KANO; CHIBA, 2012).

3.4.4 Anti-inflammatory effect

Inflammation is a natural biological mechanism in the human body in which the immune system protects against tissue damage due to physical trauma, harmful chemicals, and microbial agents. A wide range of progressive diseases is related to inflammation, resulting in dysregulation of cell signaling, the exaggerated appearance of cytokines, abrogation of the barrier function to inflammatory cells, and oxidative damage of tissues and organs, resulting in systemic disorders. Thus, some studies suggest that the reduction or inhibition of chronic inflammatory mechanisms can

prevent numerous diseases. Thus, a diet with anti-inflammatory components has beneficial biological activities (LEI *et al.*, 2018; YUSOF *et al.*, 2019).

In inflammation, macrophages can be activated by toll-like receptors (TLR) (e.g., lipopolysaccharide (LPS) and interferon- γ), which results in the production of several pro-inflammatory cytokine substances, such as inducible nitric oxide synthase (iNOS), cyclooxygenase-2 (COX-2), interleukin 1 β (IL-1 β), interleukin-6 (IL-6) and tumor necrosis factor-alpha (TNF- α) (MÜLLER *et al.*, 2017). Cytokines consist of soluble proteins that influence the behavior of other cells involved in cellular immunity and the inflammatory response. Thus, such pro-inflammatory substances contribute to the development of numerous metabolic disorders (GRAY; BLOCH, 2012; MURRAY; WYNN, 2011).

However, inflammation control appears as a promising therapeutic alternative to maintaining health and well-being. Foods that assist in this process should be considered. The use of fermented soy foods to control inflammatory processes is reported, and active metabolites, such as proteins, phenolic compounds, and antioxidants, are found and associated with anti-inflammatory effects (DIA; BRINGE; MEJIA, 2014).

In the inflammatory reaction, the macrophage produces nitric oxide (NO), which is usually detected as iNOS (inducible nitric oxide synthase). (LIAO *et al.*, 2010; SZLISZKA *et al.*, 2011). Isoflavones act as inhibitors of NO production and consequently cancel the production of IL-1 β and TNF- α pro-inflammatory cytokines. After ingestion, these compounds inhibit the expression of COX-2, the production of pro-inflammatory cytokines and the activation of the nuclear transcription factor kappa-B (NF- κ B). Thus, the expression of several genes during inflammatory responses is controlled and regulation of innate and adaptive immunity occurs. Isoflavones still

affect the mechanisms of inflammation containing the inflammatory process through different intracellular signaling pathways triggered by AP-1, PPAR, Nrf2 and MAPKs (CHEN *et al.*, 2018; YUSOF *et al.*, 2019).

Epidemiological investigations show the associations between different soy foods and inflammatory markers, including highly sensitive C-reactive protein (hs-CRP), interleukin (IL) -6 and IL-18. The high levels of intake of these foods, including miso and soy sauce, are related to a reduction in the serum level of IL-6, a pro-inflammatory cytokine associated with several chronic diseases (ASSOCIATIONS BETWEEN INTAKE OF DIETARY FERMENTED SOY FOOD AND CONCENTRATIONS OF INFLAMMATORY MARKERS.PDF, [s. d.]

Above all, the anti-inflammatory effects of isoflavones are confirmed by the fact that they act in the elimination of reactive oxygen species (ROS), which are directly involved in inflammation

3.4.5 Preventive effect against cardiovascular disease

Some evidence reports the association between high soy consumption and the preventive effect against cardiovascular diseases (CVD), such as a lower risk of ischemic heart disease (IHD) or stroke. Soy protein and isoflavones are the constituents responsible for the lower risk of CVD, in addition to their beneficial effects such as lipid profile, arterial stiffness, blood pressure, and endothelial functions (NAGATA *et al.*, 2017).

Nattokinase (NK) is an enzyme contained in the sticky component of natto, cheese-like food made from soybeans fermented with *Bacillus subtilis*, which can dissolve thrombi and fibrin. Because it is considered stable in the gastrointestinal tract,

NK becomes an appropriate agent for oral thrombolytic therapy (DABBAGH *et al.*, 2014).

It is because NK acts directly degrading fibrin or activates other fibrinolytic enzymes, such as pro-urokinase and tissue plasminogen activator (t-PA). NK also inactivates plasminogen activator inhibitor-1 (PAI-1) *in vitro*, the primary inhibitor of t-PA, resulting in the enhancement of fibrinolysis (TAKABAYASHI *et al.*, 2017; YATAGAI *et al.*, 2008).

The development of intravascular thrombi causes a variety of CVDs. Studies suggest that natto has broad thrombolytic efficacy and its ingestion has protective effects against CVD (KUROSAWA *et al.*, 2015).

3.4.6 Neuroprotective effect

The human brain is singularly vulnerable to oxidative damage and has high oxygen consumption, in addition to having a relatively high content of polyunsaturated fatty acids (PUFA), which are sensitive to oxidation. Otherwise, neurons are particularly sensitive to disturbances in the balance between antioxidants and the production of reactive oxygen species (ROS), since the levels of antioxidant defense in the brain are negligible. High content of active redox metals is found in the brain, which promotes the formation of ROS and is associated with the development of pathologies (KANG *et al.*, 2016).

Glial cells, such as astrocytes, oligodendrocytes, and microglia, are cell types present in the central nervous system that play a beneficial role in the viability and survival of neural cells. The latter is like macrophages found in the brain and plays an important role in defending the host and repairing tissues in the central nervous system

(CNS). In response to extracellular stimuli, microglia release pro-inflammatory mediators, such as nitric oxide (NO), prostaglandin E₂ (PGE₂), reactive oxygen species (ROS), and pro-inflammatory cytokines, such as interleukin (IL)-1 β and tumor necrosis factor (TNF)- α that works to restore CNS homeostasis by eliminating pathogens and infected cells (DANDONA; AJAY; SANDEEP, 2010; LOANE; BYRNES, 2010).

A possible therapeutic approach for the treatment of neurodegenerative diseases is to control microglial activation and reduce the number of pro-inflammatory factors since the overproduction of inflammatory mediators and cytokines causes chronic neuroinflammation, develops several neurodegenerative diseases, and can occasionally lead to neural cell death (DENG *et al.*, 2011).

Studies show that isoflavones are protective against neuronal cell death, elevate existing neuronal function, and boost neuronal regeneration. Thus, interest in the consumption of fermented soybean foods rich in isoflavones is growing due to their supposed beneficial effects, such as the ability of genistein to inhibit the apoptotic signaling cascade in neurons (DIAS *et al.*, 2012).

3.4.7 Anti-aging effect

Aging is seen as an inevitable, universal, multifactorial and, complex progressive decline in the physiological functions of all living beings, affecting the condition of relative stability and making it susceptible to age-related injuries and diseases (LARA; MICHAEL J.; MARGARET, 2016; MAGALHÃES; STEVENS; THORNTON, 2017).

Healthy aging is the surest way to extend your health. Thus, therapies that help achieve healthy aging become an efficient path to longevity in humans. In this

search for the longevity of the body, antioxidant therapy has beneficial effects, pointing out the role of dietary antioxidants. The accumulation of oxidized molecules, such as lipid peroxides, proteins, and damaged DNA mediated by oxidative stress (OS), is the result of the aging process and the administration of antioxidants can prevent oxidation or exclude the production of free radicals, characterized by affecting the rate of aging (GAMAN; STOIAN; ATANASIU, 2011; LEONARDI, 2017; REVIEWS, 2016).

Traditionally, fermented soybean food products are mentioned as having anti-aging properties. These effects are associated with the isoflavones aglycone genistein and daidzein (PAN *et al.*, 2012). Studies suggest the anti-aging effect of Tempeh in the pre and postmenopausal with the maintenance of the quality of the uterus, the improvement of skin quality, and bone strength (KHOSRAVI; RAZAVI, 2021; SAPBAMRER; VISAVARUNGROJ; SUTTAJIT, 2013). It is because isoflavones act in the replacement of estrogen, improving the quality of life of postmenopausal women (DAS *et al.*, 2020).

Some research shows that neuroinflammation is related to low-grade systemic inflammation, common in aging. The cascade of neuroinflammation also correlated with systemic inflammation is one of the most widely accepted suspicions regarding Alzheimer's disease (AD), one of the main common forms of age-related dementia. Antioxidant and anti-inflammatory nutrients are mentioned as agents that help to reduce or delay the development of AD (SHIRLEY STEFFANY MUÑOZ; SANDRA MARIA LIMA, 2018).

Neuroinflammation in the brain can be reduced by the isoflavones present in fermented soy foods, known to have antioxidant activity. A promoter of proinflammatory activity, IL-1 β , is decreased while a potent anti-inflammatory cytokine, IL-10, is increased. Also, the intake of isoflavones increases cognitive capacity and

prevents oxidative damage in neurons. In AD neurodegeneration, the most damaging sequel is memory loss, with the first implicit mechanism being the deficit of cholinergic neurons, where the transmission of information is canceled due to the lack of neurotransmitters such as acetylcholine (ACh). Research suggests that isoflavones reverse amnesia by increasing acetylcholine and reducing levels of acetylcholinesterase (AHMAD *et al.*, 2014; CHAN *et al.*, 2018; OVERK *et al.*, 2010).

After dietary supplementation with Chungkookjang, a fermented soybean paste from Korea cultivated by *Bacillus* sp., components, such as the isoflavonoids daidzein and genistein, elevated the activity of superoxide dismutase, an important free radical scavenging enzyme (JEONG *et al.*, 2021). It has also been reported that, by improving memory functions and other neurological indications, gerbils induced by stroke are improved (KIM *et al.*, 2021). These, express cerebral ischemia after transitory artery occlusion, and afterward, have global neural cell death due to the addition of oxidative stress, and neuroinflammation (PARK *et al.*, 2011; YEON *et al.*, 2020).

Oxidative stress is the cause of reduced life expectancy. Some findings suggest that Natto significantly prolonged the life of nematodes, increased resilience to oxidative stress and postponed the accumulation of lipofuscin, a characteristic of aging cells. This is cited for its anti-aging effect, relying on actions such as preventing heart attacks, stroke, osteoporosis, bowel disease, and improved cognitive function, especially with age (IBE *et al.*, 2013; NAGATA *et al.*, 2017).

The anti-aging effect of fermented soy foods can also be attributed to the high concentration of spermidine, bioactive polyamines found in high levels in foods such as natto and tempeh. Such a compound has several important functional and regulatory properties related to the physiology of cell aging, such as reversing memory

loss, improving the blood lipid profile, and reducing cardiovascular risks, inducing autophagy in damaged cells (SAGARA *et al.*, 2017).

3.4 SARS-COV-2 AND THE POTENTIAL APPLICATION OF FERMENTED SOY PRODUCTS IN THE PREVENTION

A new strain of Coronavirus not previously identified in humans was reported in Wuhan, China in December 2019, being identified as a beta type of Coronavirus β -CoV Group 2B. A total of seven human coronaviruses (HCoVs) have been identified before: HCoV-229E, HCoV-OC43, HCoV-NL63, HCoV-HKU1, SARS-COV - responsible for severe acute respiratory syndrome, MERS-COV - responsible for the Middle East respiratory syndrome, and recently the new coronavirus, called SARS-CoV-2 - responsible for causing the new severe respiratory inflammatory disease, COVID-19 (OPAS, 2021; SANTACROCE *et al.*, 2021).

As of early May 2022, a total of 514.918.067 confirmed cases and 6.240.940 deaths caused by COVID-19 have been reported to the World Health Organization (WHO) (WHO, 2021). SARS-CoV-2 enters the body using angiotensin-converting enzyme 2 (ACE2) and transmembrane protease, serine 2 (TMPRSS2), as target receptors to infect the cells. After affecting the epithelial cells of the human respiratory tract, the rapid replication of the virus leads to a storm of pro-inflammatory cytokines and chemokines. This hyperinflammatory state causes oxidative stress leading to damage to the alveolar and endothelial cells of the lung and chronic lung inflammation (MAHMUDPOUR *et al.*, 2020; ZENG *et al.*, 2020).

Given the pandemic scenario, numerous researches investigate risk factors, clinical manifestations, and possible preventive and therapeutic actions. Health

conditions such as obesity, diabetes, previous morbidities with risk of immunodeficiency, and chronic cardiovascular, renal, and respiratory diseases are also investigated for association and relationship to the high severity of COVID-19 (GOUDA *et al.*, 2021). Among the complications caused by the disease are ARDS, septic shock, coagulation dysfunction, metabolic acidosis, cardiac arrhythmia, kidney damage, liver dysfunction, heart failure, or secondary infection (COSTELA-RUIZ *et al.*, 2020).

A therapeutic strategy for the control of SARS-CoV-2 consists of identifying anti-inflammatory agents to act on the reduction of uncontrolled inflammation in patients and the receptors for the ACE-2 enzyme since it is widely expressed by epithelial cells of the lung, kidney, heart, blood vessels, and intestine (HUANG *et al.*, 2020). Flavonoid-derived bioactive compounds, such as isoflavones, are mentioned for their significant health benefits such as antibacterial, antioxidant, anticancer, anti-inflammatory, and immunomodulatory bioactivity (MUCHTARIDI *et al.*, 2020).

There are few studies related to dietary habits as a risk factor for COVID-19 instability. However, some differences in diet have been hypothesized to play a potential role in disease and fatality rate variability (CDC, 2020). Based on mechanistic and clinical data, vitamins and folate, polysaccharides and dietary fiber, lipids, peptides, and natural polyphenols are known to be necessary for the body's immune system against viruses (CALDER *et al.*, 2020).

For example, certain countries, such as Bulgaria, Greece, Romania, and Turkey, where there is high consumption of some types of fermented foods (cabbage and milk), are associated with lower mortality rates. The possible protective effects of antioxidants and angiotensin-converting enzyme (ACE) inhibiting peptides present in

fermented foods may justify this hypothesis (BOUSQUET *et al.*, 2020; MOHSENI *et al.*, 2021).

Adem *et al.* (ADEM *et al.*, 2020) performed a molecular docking study to identify the ability of 80 flavonoid compounds to bind to the 3-chymotrypsin-like protease (3CL^{pro}), a known enzyme important for SARS-CoV replication. Other polyphenols and flavonoids, such as daidzein and genistein (found in fermented soy products), have been proposed as potential inhibitors of the main SARS-CoV-2 protease (PENDYALAA; PATRASA, 2020).

As already mentioned, bioactive peptides with therapeutic properties, including antihypertensive antioxidant, antitumor, and antidiabetic, are present in fermented soy products. Fermented soy peptides have previously demonstrated activity against several viruses, including the SARS-CoV responsible for the SARS outbreak in 2003 (CHOURASIA *et al.*, 2020). In the soybean fermentation process, the proteolytic degradation of the soy protein fractions (glycine and β -conglycinin) generates the ACE inhibitory peptides (KUBA *et al.*, 2005). Studies have already mentioned the identification of ACE inhibitory and antihypertensive peptides in *Natto* and, also, two ACE inhibitory peptides isolated from *tofu* (SACHIE *et al.*, 2009; UBA *et al.*, 2003). Other foods, such as *douchi* (fermented by *A. egyptiacus*) and *sufu* (a soybean fermented by the fungus), have peptides with ACE inhibitory activity (MA; CHENG; YIN, 2013; ZHANG *et al.*, 2006).

Oba *et al.* (OBA *et al.*, 2021) conducted a study with *Natto* to investigate the antiviral activities of this food against SARS-CoV-2. The results showed that *Natto* extract fully inhibited severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection in the cells. The protease activities of *Natto* extract were able to proteolytically

degrade the British variant of the spike protein (receptor binding domain; RBD) of SARS-CoV-2, resulting in the inhibition of viral infections in cells.

A study conducted by Chourasia et al. (CHOURASIA *et al.*, 2020) from soy cheese fermented with *Lactobacillus delbrueckii* WS4, identified and selected peptides for antiviral activity in silico. A total of twenty-three peptide sequences were examined for binding affinity to critical residues of the SARS-CoV-2 RBD protein and important catalytic residues of the SARS-CoV-2 pro-enzyme 3CL using molecular docking. The authors also showed in molecular docking studies of the selected peptides that they revealed a potential peptide "KFVPKQPNMIL". This peptide showed a strong affinity for significant amino acid residues, for host cells (RBD) of the SARS-CoV-2 peak S1 glycoprotein that are responsible for binding the virus to the human ACE2 protein receptor and also an affinity for the important viral proteolytic enzyme 3CLpro. for viral replication.

Therefore, it can be concluded that fermented soy cheese could be explored as a prophylactic food for SARS-CoV-2 and related viruses. Furthermore, the multi-target inhibitor peptide, which effectively inhibited both viral proteins, could be used for in vitro and in vivo functions against SARS-CoV-2.

3.6 BIOACTIVE COMPOUNDS IN SOYBEAN FERMENTED PRODUCTS

Soy has long been consumed as a health food and fermented soybean products are important components of traditional diets in Asian countries. The benefits of fermented soy are attributed to its phytochemical content and bioactive compounds, which confer numerous benefits to human health (CUI *et al.*, 2020).

Many compounds are responsible for the bioactive properties of fermented foods mentioned in several studies. Isoflavones are compounds found in tempeh, which act as antioxidants and are also related to many chronic diseases (AMIN *et al.*, 2020). Surya and Romulo showed that tempeh extracts protect HepG2 cells (human liver cancer cell line) against induced oxidative stress by reducing ROS generation, and eventually cell death (SURYA; ROMULO, 2020).

Chungkookjang has compounds such as isoflavone aglycones, peptides, and dietary fiber, and is rich in poly- γ -glutamic acid (γ -PGA). Consumption of this food can act on memory impairment induced by Alzheimer's disease and cerebral ischemia, so it prevents and alleviates neural cell survival, thus improving brain insulin sensitivity and neuroinflammation (JEONG *et al.*, 2021).

To investigate the hypothesis that chungkookjang consumption improves sensitivity and insulin secretion capacity an animal model study reminiscent of the characteristics of type 2 diabetes in Asians was conducted. A high concentration of daidzein was observed and related to the anti-diabetic properties of chungkookjang, capable of improving glucose regulation by potentiating insulin secretion and reducing insulin resistance (JEONG *et al.*, 2020).

Recently, research in Japan has shown that higher consumption of natto and miso is associated with a lower risk of mortality (KATAGIRI *et al.*, 2020). These foods are sources of bioactive compounds such as nattokinase, bacillopeptidase F, vitamin K2, dipicolinic acid, γ -polyglutamic acid, isoflavones, vanillic acid and syringic acid, which have health-promoting effects (CAO *et al.*, 2019).

Nattokinase is shown to be responsible for anti-thrombotic and anti-coagulative activities. The anti-thrombotic effect of NK can be used for the treatment of cardiovascular diseases, and such a compound also acts on amyloid degradation

related to Alzheimer's disease and on the suppression of atherosclerosis, heart attack, and stroke in sick patients (CHAN *et al.*, 2021; REN *et al.*, 2017). The health benefits of miso are associated with the presence of isoflavones, such as 8-OH-daidzein, 8-OH-genistein, 6-OH-daidzein, which have strong antioxidant activity, this is related to a series of beneficial effects on human health (HIROTA *et al.*, 2004).

Doenjang is an important food consumed in Korea, such food shows strong activities against several carcinogens/mutagenic agents, such as aflatoxin B1. Park et al showed through studies that genistein and linoleic acid present in doenjang extracts have strong antimutagenic activities, being more effective among the other bioactive compounds found in this food, such as β -sitosterol, soy saponin, α -tocopherol, genistein and phytic acid (PARK *et al.*, 2003).

Fermented soy foods are composed of molecules, vitamins, and peptides that are found in greater availability after the fermentation process, which point to being a potential source of numerous health benefits. The bioactive compounds found in fermented soy products are associated with the microorganism used in the process as well as the traditional practice used in each region.

Several bioactive compounds are found in different fermented soy products, these and their health benefits are illustrated in Table 3.

TABLE 2 - BIOACTIVE COMPOUNDS OF DIFFERENT FERMENTED SOY PRODUCTS AND THEIR HEALTH BENEFITS.

Soybean products	Bioactive compounds	Health benefits	References
Tempeh	Isoflavone aglycone	Antioxidant properties	(ROMULO; SURYA, 2021)
Tempeh	Isoflavone aglycone	Protection of HepG2 cells from oxidative stress	(SURYA; ROMULO, 2020)
Tempeh	Genistein	Immunomodulatory Function	(AOKI <i>et al.</i> , 2020)
Tempeh	Trans-cinnamic acid	Antioxidant properties	(PRADO <i>et al.</i> , 2021)

Chungkookjang	Poly- γ -glutamic acid (γ -PGA)	Prevention of memory loss from Alzheimer's and cerebral ischemia	(JEONG <i>et al.</i> , 2021)
Chungkookjang	Daidzein	Anti-diabetic property	(JEONG <i>et al.</i> , 2020)
Chungkookjang	Poly- γ -glutamic acid	Anti-obesity effect	(CHOI <i>et al.</i> , 2016)
Chungkookjang	Bacillomycin D and surfactin	Antimicrobial activity	(LEE <i>et al.</i> , 2016)
Natto	Nattokinase (NK)	Anti-thrombotic and anti-coagulant activities	(CHAN <i>et al.</i> , 2021)
Natto	Vitamin K2	Reducing osteoporotic fracture risk	(KOJIMA <i>et al.</i> , 2019)
Natto	Bacillopeptidase F	Anti-thrombotic and blood pressure-lowering	(HITOSUGI; HAMADA; MISAKA, 2015)
Natto	Nattokinase (NK)	Fibrinolytic activity	(PAGNONCELLI <i>et al.</i> , 2017)
Miso	Isoflavones aglycones	Anti-tumoral activity	(ABE <i>et al.</i> , 2021)
Miso	Isoflavones aglycones	Protective effects against stroke	(WATANABE <i>et al.</i> , 2018)
Miso	Isoflavones aglycones	Sympathetic nerve activity	(ITO, 2020)
Doenjang	Linoleic acid and Genistein	Antimutagenic activity	(PARK <i>et al.</i> , 2003)
Doenjang	Genistein	Antimutagenic and anticancer activities	(JUNG; PARK, 2006)
Doenjang	Genistein	Anti-obesity effects	(CHA <i>et al.</i> , 2012)
Kinema	Poly- γ -glutamic acid (γ -PGA)	Suppression of postprandial hyperglycemia	
Kinema	Isoflavones aglycones	Antioxidant properties	(SANJUKTA; 2016)
Kinema	Group B saponins	Prevention of dietary hypercholesterolemia	(OMIZU <i>et al.</i> , 2011)
Douchi	β -glucosidase and protease	Antioxidant activity	(YANG <i>et al.</i> , 2019)
Sufu	Isoflavones aglycones	Enhancement of the physiological function	(LI-JUN <i>et al.</i> , 2004)

SOURCE: The author (2022).

3.8 RECENT PATENTS AND INNOVATIONS ON BIOACTIVE COMPOUNDS IN SOYBEAN FERMENTED PRODUCT

As presented throughout this review, the bioactive compounds present in fermented soy products are related to several beneficial activities for human health and well-being. Scientific studies confirm this potential, and to complement this view, a patent search was conducted showing recent advances and innovations in the use of such compounds.

The patent search was conducted on the Derwent Innovations Index patent database, on March 25, 2022, performing one search for each bioactive compound in the field "Title", namely *Linoleic acid*, *Daidzein*, *Genistein*, *Isoflavone aglycone*, *Nattokinase*, *Cinnamic acid*, and *Vitamin K2*, using the wildcard * to retrieve all documents containing the defined word roots. The keywords were combined with the International Patent Classification (IPC) A61P (Specific therapeutic activity of chemical compounds or medicinal preparations) using the Boolean operator AND. The IPCs A61P 003/04 (Anorexiant; Antiobesity agents), A61P 003/10 (Hyperglycaemia, e.g. antidiabetics), A61P 009/12 (Antihypertensives) and A61P 037/04 (Immunostimulants) were used to refine the search when necessary (WIPO, 2022). The time interval was the last five years, 2018 to 2022.

After analyzing the documents by reading the titles and abstracts, 334 documents were classified and analyzed using Microsoft Excel software. Along these years it is possible to observe an increase in the number of registered patent documents related to the development and innovation of products made from substances obtained in the soybean fermentation process. The apparent decrease in the number of documents in 2022 is attributed to the date of search (March 2022) and to the secrecy period of usually 18 months before publication. China (CN) and Japan (JP) were the countries that filed the most patents (Figure 4). China accounted for 169 registered patents, representing 50.6% of the total analyzed, while Japan registered 42 patents (12.57%).

The substances generally most abundant in the fermented soybean product were Nattokinase, Genistein and Cinnamic acid because they are widely used in pharmaceutical formulations, for instance, in the prevention and treatment of chronic diseases. This was observed in terms of technology, because the highest number of

documents was found for Nattokinase (79), followed by Genistein (69) and Cinnamic Acid (66).

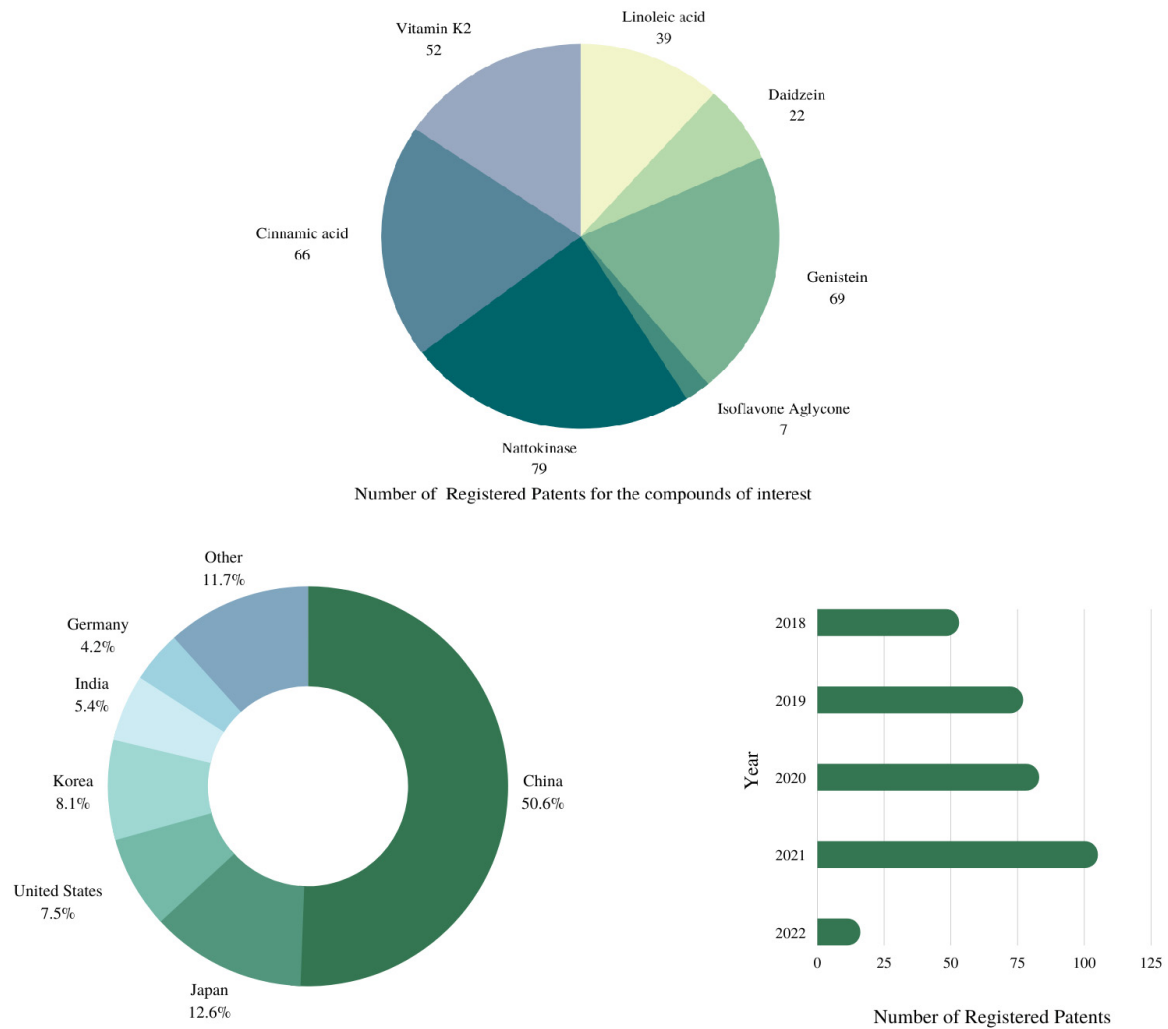


FIGURE 3 - NUMBER OF REGISTERED PATENTS FOR THE COMPOUNDS FOR DE INTEREST, COUNTRIES AND YEARS OF PUBLICATIONS.

SOURCE: The author (2022).

The main company (assignee) that contributed to these patent filings was HUGHES BIOTECHNOLOGY CO. LTD®, Taiwan, with 11.59% patent documents. This company is specialized in the development and manufacture of plant-based

nutraceuticals, focusing on the discovery and development of dietary ingredients that are based on the most current science, and reformulating existing ingredients to increase their potency. Other important assignees were DONGGUAN ANHAO PHARM CO. LTD[®], China, with 10.6% documents, a skin health management company that integrates product technology research and development, sales, and service, and KOBAYASHI PHARMACEUTICAL CO. LTD[®], Japan, with 7.5% documents, that develops ideas for pharmaceuticals and various other applications in daily life, such as dental hygiene skincare, and nutritional supplementation

4 MATERIAL AND METHODS

In this section, a description of the materials and methods used in this work is presented. In the following topics, the materials and methods used in the fermentation process, in performing the product extractions and in analyzing the activities and biological potential are mentioned.

4.1 MICROORGANISM

Fermentation process was performed using a fungus of the *Rhizopus oligosporus* NRRL 2710, belonging for ARS Culture Collection of the Northern Regional Research Laboratory (NRRL, USA). The strains were maintained in Potato Dextrose Agar (PDA) slants at 4 °C.

4.2 SUBSTRATE TREATMENT

The substrates used in this study (soybean and brown rice) were pre-soaked in deionized water containing 5% v/v of acetic acid. After 12 h, the substrate was thermally treated under flowing steam at 121 °C for 15 min. The excess water was removed, and the substrate cooled to the inoculation temperature at 30 °C.

4.3 FERMENTED SOYBEAN

The substrate thermally treated was inoculated with 10^7 spores g^{-1} of a dry substrate. The experiments were carried out in perforated trays at 30 °C for 72 h. After fermentation, the samples were crushed in a kitchen blender (Turbo Blender) for 5 min. A fraction was stored in a freezer, at $-18\text{ °C} \pm 2\text{ °C}$ for use in the extraction experiments (here named wet sample). Another fraction was dried in an oven at 50 °C until constant weight (here named dry sample). The samples were stored in a freezer at 4 °C until the use. The moisture of the raw material was measured using infrared moisture balance (Bel Engineering). All analyzes were performed in triplicate.

4.4 PREPARATION OF ETHANOL EXTRACTS

Alcoholic extraction of fermented substrates and control (non-fermented soybean) was performed according to the method described by (TYUG; PRASAD; ISMAIL, 2010). Dry powder was extracted with 70% (v/v) for one hour in a shaking at 120 rpm. The extracts were collected and centrifuged at 3000xg for 15 min. Supernatants were stored at -18 °C for further analysis.

4.5 SOXHLET EXTRACTION

Fermented soybeans were submitted to the Soxhlet extraction following the methodology described in AOCS procedures (AOCS, 1998), using a mass of solids to solvent volume of 1:40 (m/v) ratio. Different solvents, with different polarities, were tested to evaluate the extraction yield, as well as the extraction efficiency of phenolic compounds. Tests were carried out in triplicate for each solvent, for 6 hours of extraction. All solvents used in this study were of analytical grade. At the end of extraction, the solvent was removed with a rotary evaporator (RV 10 digital, Ika, Wilmington, USA) and then held in an oven in 50°C for 24 hours to remove residual solvent. The extract amounts were weighed in an analytical balance to calculate the extraction yield (wt%), as follows:

$$\text{Extraction yield (\%)} = \left(\frac{\text{mass of extracted oil (g)}}{\text{mass of sample (g)}} \right) \times 100 \quad (1)$$

Where the mass of the sample was expressed on a dry basis, taking into account the moisture content in all samples used before the extraction.

4.6 SUPERCRITICAL EXTRACTION

The supercritical extraction process was performed in a home-made laboratory scale extraction unit (batch extractor with 80 cm³ internal volume, L = 0.16 m, ϕ = 2.52 x 10⁻² m) as described in (CORREA *et al.*, 2016). Basically, the high-pressure extractor is connected to an ultrathermostated bath and a needle valve is used for the flow control at the outlet stream of the extractor. The system also consists of a high-

pressure syringe pump (ISCO, model 500D, Lincoln, NE 68504, USA), pressure and temperature sensors and transducers.

The extractions were performed using a liquid solvent modifier in a semi-batch fixed bed extraction process, using supercritical CO₂ as the solvent and ethanol as liquid solvent, named as scCO₂ + EtOH. The experimental procedure consisted of soaking the raw material in the solvent ethanol using a 1:1 (w/w) ratio kept the mixture static for 60 min. The mixture was loaded into the extraction vessel, and CO₂ laden until the pressure reached the extraction condition. Then, mixture containing the raw material (fermented solids) with ethanol and CO₂ was kept in a confinement period (static extraction step) for 60 min to assure the thermal and mechanical equilibration before one start the dynamic extraction period.

scCO₂+EtOH extractions were performed based on a 2² factorial experimental design with a center point, aiming to evaluate the effects of temperature from 40 to 80°C and pressure from 15 to 25 MPa on the extraction yield and the quality of the extract. The compressed solvent (supercritical CO₂) was pumped at a constant flow rate of $2.0 \pm 0.3 \text{ cm}^3 \text{ min}^{-1}$.

The extracts were collected in amber glass vessels and its weight determined at each extraction time intervals of 5 min.

4.7 ANALYTICAL PROCEDURES

4.7.1 Measurement of DPPH radical-scavenging activity

The antioxidant activity of the extracts was assessed based on the radical scavenging effect of the stable DPPH-free radical activity (1,1-diphenyl-2-

picrylhydrazyl) (Sigma-Aldrich, USA), as described by (BRAND-WILLIAMS; CUVELIER; BERSET, 1995). A methanol solution of DPPH (0.004% w/v) was prepared. One milliliter of methanolic DPPH solution was added to 250 μL sample extracts at different concentrations (50; 100; 150; 200; 250 μL μL^{-1}). After incubation in the dark at ambient temperature for 30 min, the absorbance was measured at 517 nm (Spectrum band, UV-VIS). Ethanol solution without the sample was considered as a control. IC_{50} value (index representing the concentration of the antioxidant which can reduce 50% of free radicals) was obtained using a standard curve. The inhibition rate (%) was calculated to the following equation:

$$\text{Inibição (\%)} = \left(\frac{A_{\text{controle}} - A_{\text{extrato}}}{A_{\text{controle}}} \right) \times 100 \quad (2)$$

Where:

A_{control} = DPPH absorbance (diluted with ethanol solution)

A_{extract} = DPPH + sample extract absorbance (diluted with ethanol solution)

Percent antioxidant activity was plotted against log absorbance values, generates a straight line to calculate the half-inhibitory concentration (IC_{50}) in g g^{-1} . The anti-free radical activity (expressed as IC_{50}) is defined as the amount of antioxidant substances needed to reduce 50% of the initial concentration of DPPH. The higher the power anti-free radical, the most effective antioxidant and consequently lower the IC_{50} value (BRAND-WILLIAMS; CUVELIER; BERSET, 1995). Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid) (Sigma-Aldrich, USA) was used as standard, prepared in concentrations from 1 to 100 μM of trolox mL^{-1} . The results comparing the equivalence of the antioxidant potential of the extracts with the standard potential were expressed.

4.7.2 Determination of total phenolic content

Total phenolic content of the extracts was determined by the Folin-Ciocalteu method (SWAIN; HILLS, 1959) with modifications. This test is based on the oxidation of phenolic groups with phosphomolybdic and phosphotungstic acids. After oxidation, the absorbance of the green-blue complex formed was measured at 760 nm. In a test tube, 0.1 mL of the sample was mixed with 0.5 mL of Folin-Ciocalteu reagent (10% v/v) and 0.4 mL of sodium carbonate (7.5% w/v). Samples were shaken for 1 h protected from light (reaction time). Ethanol was used as blank. The samples were analyzed in triplicate. A standard curve prepared using gallic acid monohydrate was used to calculate the total phenol content, which was expressed as gallic acid equivalent in $\mu\text{g g}^{-1}$ of the extract (Singleton, 1965).

4.8 DETERMINATION OF B-GLUCOSIDASE ACTIVITY.

β -glucosidase activity of ethanol extracts (0, 24, 48 and 72 h) was assayed by monitoring the release of p-nitrophenol (pNP) using p-Nitrophenyl- β -D-glucopyranoside (pNPG) as the substrate, according to the method described by Zhang et al., 2012. A total of 20 μl of appropriately diluted enzyme solution and 280 μl of 1.785 mM pNPG in 100 mM acetate buffer pH 4.8 were incubated for 15 min at 50 °C. The reaction was quenched by adding 400 μl of glycine buffer, and the amount of p-nitrophenol released during the reaction was determined with a spectrophotometer at 430 nm. A p-nitrophenol calibration curve was previously prepared to calculate the enzyme activity. One unit of β -glucosidase activity was defined as the amount of enzyme equivalent to the release of 1 μmol of p-nitrophenol per minute.

4.9 DETERMINATION OF FATTY ACIDS PROFILE

Fatty acid profile of fermented soybean oil was analyzed using a Shimadzu chromatograph (GC 2010 *Plus*), a capillary column (SH-Rtx-Wax (Shimadzu): 30 m x 0.32 mm x 0.25 μm), flame ionization detector (FID) and split injection mode (1:10). The injector and detector temperatures were 240 °C and 250 °C, respectively. The oven temperature was programmed to start at 100 °C during 5 min, followed by an increase up to 240 °C at a rate of 4 °C min^{-1} and maintained in this temperature for 5 min. The carrier gas was Helium at 32.5 $\text{cm}^3 \text{min}^{-1}$. The samples were prepared according official method (AOCS, 1998) to convert triacylglycerol and free fatty acid of samples into fatty acid methyl esters (FAMES). FAMES were identified by comparison with retention times of the standard mixture FAMES (Supelco, MIX FAME 37, St. Louis, MO 63103, USA). The quantification of fatty acid was conducted by normalization area procedure. Results were expressed as percentage of each individual fatty acid present in the sample.

4.10 IDENTIFICATION AND QUANTIFICATION OF ANTIOXIDANT COMPOUNDS BY RP-HPLC-DAD/UV

The ethanol extracts (0, 48 and 72 h) were filtered with Spritzen syringe filter of 0.22 μm (Millipore, Bedford, MA). The filtrate was used to quantify the polyphenolic compounds by High-performance liquid chromatography with UV–vis detection (HPLC-DAD/UV–vis). HPLC analysis was performed according to (Haminiuk et al., 2014), with a reversed phase Acclaim® 120 column, C18 5 μm 120 Å (4.6 mm×250 mm). The

column was maintained at 40 °C and the detection was recorded at 280, 300 and 320 nm. Polyphenolics and flavonols were detected in the range of 210 and 320 nm. The mobile phase was acidified water with phosphoric acid 1 % and methanol. The solvent gradient was as follows: 0–15 % B for 2 min, 15–25 % B for 5 min, 25–30 % B for 10 min, 30–35 % B for 15 min, 35–50 % B for 25 min, 50–60 % B for 30 min, 60–80 % B for 35 min, 80–100 % B for 45 min and 100–5 % B for 60 min. Flow rate: 1 mL min⁻¹. Standards used: for phenolic acids (gallic acid, chlorogenic acid, trans-cinnamic acid, caffeic acid, p-coumaric acid, ferulic acid) and for flavonols (rutin, myricetin, quercetin and kaempferol). Stock solutions of all standards were prepared in methanol and the calibration curves were obtained from triplicate injections of at least five concentrations and identification based on the retention times of the standards (Granato et al., 2011) in chromatogram at 280 nm. Retention times (min) for the standards at the conditions used were: gallic acid: 6.83 min; caffeic acid: 32 min; p-coumaric acid: 18.54 min; ferulic acid: 20.07 min; rutin: 25.03 min; myricetin: 28.22 min; trans-cinnamic acid: 32.40 min; kaempferol: 37.08 min.

4.11 HIGH – PERFORMANCE LIQUID CHROMATOGRAPHY OF PHENOL CONTENTS (HPLC)

The phenolic content of the extracts was analyzed in high performance liquid chromatography using an Agilent Technology 1200 Series system, coupled to a diode array detector (DAD) at wavelengths 235, 260, 275, 280, 290, 311, 357, 370 nm and a scanning from 190 nm to 600 nm. A Zorbax Eclipse XDB – C 18 (4.6 x 150 mm, 5 – micron) column was used at 0.7 mL min⁻¹ flow. The mobile phase utilized was methanol and acetic acid 2.5% (50:50 v/v). Chlorogenic acid, caffeic acid, ferulic acid, tocopherol, genistein, daidzein, trans-cinnamic acid, catechin, rutin, p-coumaric acid,

gallic acid, resveratrol and epicatechin (SIGMA Aldrich, EUA) was used as standards. To obtain the calibration curve, all standard reagents (SIGMA Aldrich, EUA) were solved in mobile phase and used at 1, 2, 5, 8 and 10 ppm. The injection volume was 10 μL and the run time was 36 min. The resulting chromatograms values were graphed, and linear equation was used to calculate the phenolic compounds contents of the samples. The samples were microfiltered through a hydrophilic membrane GV (Durapore) made of polyvinylidene difluoride (PVDF), with a pore size of 0.22 μm . The results were showed in $\mu\text{g g}^{-1}$ of the phenolics compounds identified.

4.12 EVALUATION OF ANTI-INFLAMMATORY ACTIVITY

In order to evaluate the anti-inflammatory activity with inhibition of the hyaluronidase enzyme, an *in vitro* method was used according to (STROMINGER; LELOIR; REISSIG, 1955). The anti-inflammatory potential was measured at concentrations of 0.5 to 2.5 $\mu\text{g mL}^{-1}$. For the sample analysis, was placed 50 μL of extract in different concentration and 0.5 mL of potassium salt of hyaluronic acid (Sigma-Aldrich, St. Louis, MO, USA) (1.2 mg hyaluronic acid per mL of 0.1 M acetate buffer, pH 3.6, containing 0.15 M NaCl) in a reaction tube. The control tube consisted of the same reagent of test tubes without ExPP. All tube were incubated for 5 min at 37 °C and after that 50 μL of the enzyme hyaluronidase (350 units of the enzyme hyaluronidase type IV-S from bovine *testes*, Sigma-Aldrich, St. Louis, MO, USA - dissolved in the same buffer substrate at concentration 6.5 mg/mL) was added, and incubated at 37 °C for 40 min. The reaction was stopped by adding 10 mL of 4 N sodium hydroxide solution and immediately placing 0.1 mL of 0.8 M potassium tetraborate into the reaction mixture and incubating it in a boiling bath for 3 min. After

the incubation time, we added 3 mL of 4-dimethylaminobenzaldehyde (DMAB) (10% solution in glacial acetic acid containing 12.5% 10 N hydrochloric acid) to the reacted mixture and incubated the solution at 37 °C for 20 min. Next, we measured the samples in a spectrophotometer (SP 2000 UV Spectrum) at 585 nm. DMSO was used as positive control due to its ability to completely inhibit de hyaluronidase enzyme. Propolis, a natural anti-inflammatory agent, was also included as a positive control. The results were expressed as the ability to inhibit the hyaluronidase enzyme, in percentage.

4.13 CYTOTOXICITY ASSESSMENT FOR TUMOR (CACO-2) AND NORMAL (MCF-10) CELLS BY COLORIMETRIC ASSAY MTT (1- (4,5-DIMETHYLTHIAZOL-2-YL) -2,5-DIPHENYL TETRAZOLIUM)

The cell lines CaCo-2, which is Colorectal Cancer, and MCF-10 A, which is a non-tumoral breast epithelial cell, were used to perform the assays with ethanol extracts from *R. oligosporus* NRRL 2710 fermentation of SB + BR. The ethanol extracts were first lyophilized for subsequent resuspension. Both cell lines were purchased FROM the Cell Bank of Rio de Janeiro, Brazil, passage 33. The lines were cultured in a 96-well plate in DMEM F12 medium with 1% penicillin/streptomycin for 24 h for cell adhesion. CaCo-2 cells were plated at 5×10^4 cells per well and MCF 10A cells at the concentration of 3×10^4 cells per well. After the 24 h incubation period, the medium was monitored by DMEM F12 with 10% penicillin/streptomycin and supplemented with 10% fetal bovine for both species. Plates were maintained in an incubator at 37 °C, 95% air atmosphere and 5% CO₂ for 48 h.

Cell viability was analyzed by colorimetric assay MTT (1- (4,5-dimethylthiazol-2-yl) -2,5-diphenyl tetrazolium), described by (MOSMANN, 1983). The MTT solution

(Sigma-Aldrich) was prepared at the concentration of 5 mg ml⁻¹ and added to the culture medium, where the final concentration in the well was 0.5 mg ml⁻¹. After 3 h of incubation at 37°C, 95% air atmosphere and 5% CO₂, the blue crystals formed were solubilized in DMSO (Panreac®), considered the most effective solvent to be capable of dissolve the formazan crystals easily. The reading was performed in a spectrophotometer (Biotek) at 595 nm.

4.14 DETERMINATION OF OXIDATIVE STABILITY

The thermogravimetric curves were obtained on a Perkin-Elmer thermogravimetric analyzer model TGA 4000 model. The temperature range was 30 °C to 800 °C and the heating rate was 10 °C min⁻¹. A dynamic atmosphere of synthetic air was used at a flow rate of 50 mL min⁻¹. Ceramic pots and initial sample masses ranging from about 8 to 10 mg were used in the apparatus.

4.15 STATISTICAL ANALYSIS

Analysis results were submitted to the statistical variance analysis and the post-test was chosen according to the normality of the data obtained such as Tukey Post-Test for data which follow normality. The program PRISMA® (GraphPad Prism 5 for Windows, version 5.4) was used. The averages of data are presented in graphs and the standard deviation shown by the error bars.

5 RESULTS AND DISCUSSION

In this section, the experimental results of *Rhizopus spp.* fermentation using different substrates as well as the effect of different extraction methods are presented. First, the influence of using different substrates on the alcoholic extract fermentation process is presented and the optimal point of bioactive compound production is evaluated by kinetics. The anti-inflammatory and anti-tumor potential of these extracts is also shown. The results of these studies are published in the journal *Biointerface Research in Applied Chemistry* (Topics 5.1 – 5.6).

Then, the experimental results are presented when the extraction methods applied were Soxhlet and supercritical CO₂ and the results when ethanol was used as a co-solvent in the process (scCO₂ + EtOH). These researches focus on these extractions with the aim of making a comparison between the results obtained and finding the best solvent for a given matrix. The results of this study are being published (Topics 5.6 – 5.11).

5.1 SUBSTRATE SELECTION FOR ANTIOXIDANT BIOSYNTHESIS

In Asian countries, the genus *Rhizopus spp.* is mentioned as one of the economically important molds due to its role as an inoculum source for the preparation of a traditional soy-based food, tempeh (Hesseltine, 1983; Astuti et al., 2000; Hartanti et al., 2015).

In preliminary studies *Rhizopus spp.* strains were grown on soybeans supplemented with different cereals (brown rice, wheat, corn and oat), aiming to achieve higher antioxidant activity. All supplementation processes were efficient in increasing the antioxidant power after 72h. Therefore, the best results ($p < 0.05$) were achieved when soybean (SB) was supplemented with brown rice (BR), resulting in the

greatest antioxidant potential for *R. oligosporus* NRRL 2710 (IC₅₀ 14.3 mg mL⁻¹) (Table 4) (PRADO *et al.*, 2021).

TABLE 3. IC₅₀ VALUES (MG ML⁻¹) DURING FERMENTATION IN SB ETHANOL EXTRACTS AND GRAIN MIXTURES AT THE STARTED POINT (T₀) AND AFTER 72 HOURS OF FERMENTATION

		IC ₅₀ (mg mL ⁻¹)	Phenolic compounds (mg GAE g ⁻¹)
SB	T ₀	105.01±0.62 ^{aa}	1.792
	72 h	20.69±0.27 ^{aa}	3.10
SB + BR	T ₀	374.5±0.61^{da}	0.697
	72 h	14.3±0.13^{aa}	6.447

* Means followed by the same letter did not differ statistically (p> 0.05).

SOURCE: The author (2022).

In the literature is vague a data on the actual increase in scavenger activities due to the fermentation process. (HUANG; LAI; CHOU, 2011) found 19.1 mg mL⁻¹ scavenging activity of fermented soybean + rice by *Aspergillus oryzae* after 16 days for sufu production, but the increment of the values in the course of the fermentation was not reported. The data of the present study clearly show the real increment in scavenging activities promoted by the fermentation process.

Also, soy supplementation with brown rice increased the concentration of phenolic compounds (0.697 to 6.447 mg GAE g⁻¹) significantly compared with the only use of soybean (1.792 to 3.10 mg GAE g⁻¹), using *R. oligosporus* NRRL 2710 (Table 4) (PRADO *et al.*, 2021). For this reason, the SB supplemented with BR and fermented by are selected for further studies (published data).

5.2 IDENTIFICATION AND QUANTIFICATION OF POLIPHENOLIC COMPOUNDS

The identification and respective concentrations of polyphenolic compounds synthesized by *R. oligosporus* NRRL 2710 under SSF of SB and SB+BR were shown in table 5, as quantified by RP- HPLC-DAD/UV. Compounds such as gallic acid, ferrulic acid, rutin, miricetin, caffeic acid, trans-cinnamic acid, quercetin, kaempferol, siringic acid and p-cumaric acid were detected. These compounds have been previously found in SSF by different fungi species (ALEJANDRA *et al.*, 2019; HIROTA *et al.*, 2005; OLMOS; GARRO, 2020; ZHANG *et al.*, 2012).

TABLE 4. IDENTIFICATION AND QUANTIFICATION IN GRAM DRY BASES (MG GDB⁻¹) OF POLYPHENOLIC AT THE ETHANOLIC EXTRACTS DURING SOLID STATE FERMENTATION OF SOYBEAN MIXTURE WITH BR SYNTHESIZED BY *R. OLIGOSPORUS* NRRL 2710.

Substrates	Concentrations ($\mu\text{g mL}^{-1}$)	Fermentation time (h)	
		0	72
SB	Gallic acid	0.2	0.0
	Ferrulic acid	0.3	0.3
	Rutin	29.3	8.7
	Miricetin	9.8	7.4
	Caffeic acid	0.0	0.0
	Trans-cinnamic acid	0.0	124.7
	Quercetin	0.0	0.2
	Kaempferol	0.0	0.1
SB + BR	Gallic acid	0.0	0.1
	Ferrulic acid	0.1	0.2
	Rutin	11.8	2.9
	Miricetin	6.5	7.4
	Caffeic acid	0.0	0.0
	Trans-cinnamic acid	19.8	103.3
	Quercetin	0.0	0.0
	Kaempferol	0.0	0.1
	Siringic acid	4.0	0.0
	p-cumaric acid	0.1	0.0

SOURCE: The author (2022).

Trans-cinnamic acid was the compound present in higher amounts, reaching $124.7 \mu\text{g mL}^{-1}$ ($3,197 \text{ mg gdb}^{-1}$) and $103.3 \mu\text{g mL}^{-1}$ ($2,649 \text{ mg gdb}^{-1}$) in SB and SB+BR fermented after 72 h, respectively. There are few reports on cinnamic acid production by fermentation process (NODA *et al.*, 2011). The cinammic acid or trans-3-

phenylacrylic acid is biosynthesized by the enzyme phenylalanine ammonia lyase (PAL), which converts the amino acid phenylalanine into cinnamic acid and is an important precursor of the biosynthesis of other important groups of polyphenolic compounds in the microorganism metabolism pathway. Cinnamic acid has been studied as an anti-tumoral compound, which induces tumor cell differentiation by modulating the expression of genes implicated in tumor metastasis and immunogenicity in cultured human melanoma cells. This work indicates a great potential for producing this compound due to the high accumulation in the fermented substrate. The separation and concentration of cinnamic acid from SB +BR fermented by *R. oligosporus* NRRL 2710 represents a great perspective for application in food and supplement products.

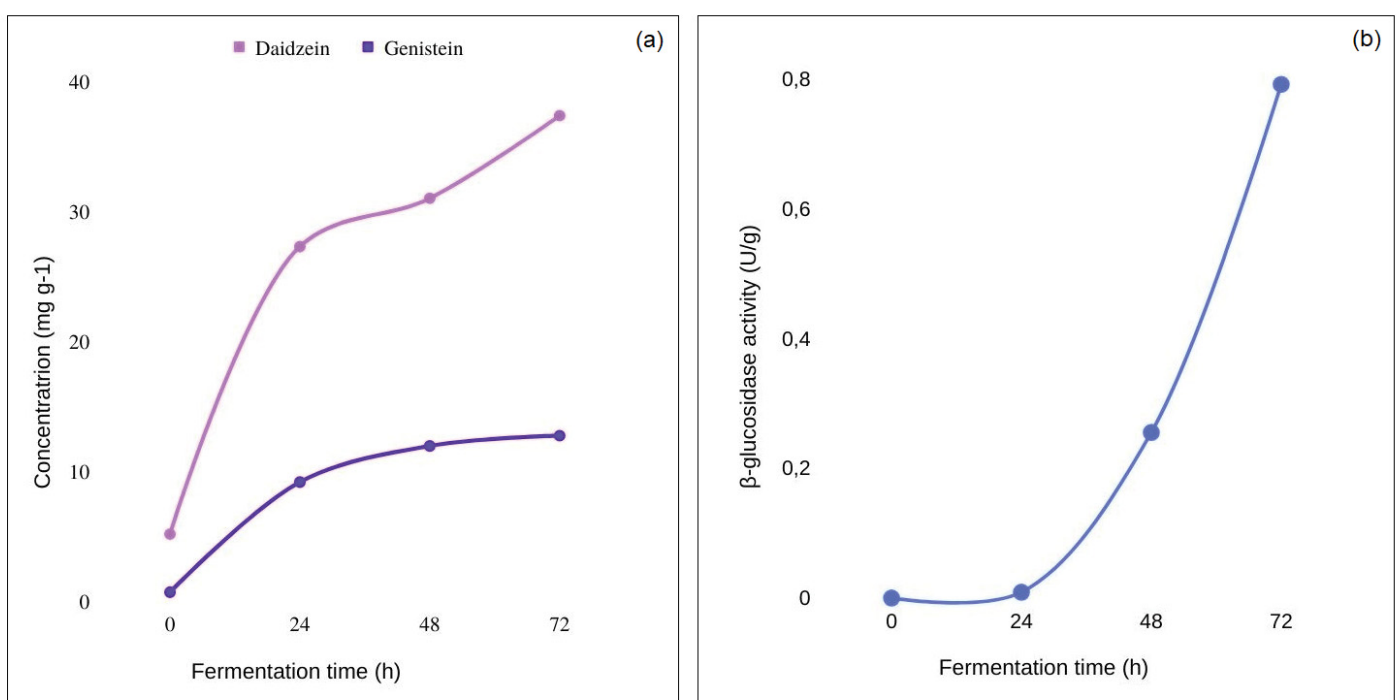
5.3 CHANGES IN THE ISOFLAVONE PROFILE DURING FERMENTATION

The isoflavones consist of a subclass of flavonoids with powerful antioxidant activity and linked to cancer prevention (BENEDETTI *et al.*, 2015). Soybeans is a known source of these compounds and isoflavones in the glycoside form (daidzin and genistin) are more present than aglycone form (daidzein and genistein). Fermentation of soybeans promotes the conversion of glycoside into aglycone isoflavones through the action of β -glucosidase produced by microorganism (DA SILVA; CELEGHINI; CHANG, 2011; MARAZZA *et al.*, 2012). All isoflavones are absorbed into the mucosa of the small intestine. However, the aglycone form is absorbed at a greater rate and has higher antioxidant activity than the glycoside form (HONG; MANDAL; LIM, 2012). In this study, the biotransformation of isoflavones during fermentation of SB+BR by *Rhizopus oligosporus* NRRL 2710 was studied using high-performance liquid Chromatography (HPLC) and β -glucosidase activity using spectrophotometry (Figure

5a). The concentrations of gaidzein and genistein (aglycone form) in the final fermented product ($18.65 \pm 0.08 \text{ mg g}^{-1}$ and $12.68 \pm 0.05 \text{ mg g}^{-1}$, respectively) were 3.75 and 20.10 fold higher, respectively, compared to non-fermented soybeans.

Simultaneously with the accumulation of aglycone isoflavones, β -glucosidase increased significantly after 24 h of fermentation (ranging from 0.01 to 0.81 U/g at the end of the process) (Figure 5b), being thus responsible for catalyzing aglycone isoflavones from glycine form. The values found in this study are significantly higher than those observed by (LEE *et al.*, 2019) (0.24 mg g^{-1} and 0.043 mg g^{-1} for daidzein and genistein, respectively) after 72 h of soybean fermentation using *Tricholoma matsutake*, and by (SANTOS *et al.*, 2018) (1.084 mg g^{-1} and 1.810 mg g^{-1} of daidzein and genistein, respectively) after 72 h of okara fermentation with *Saccharomyces cerevisiae*.

FIGURE 4 - B-GLUCOSIDASE ACTIVITY (U/G) AND ISOFLAVONES AGLYCONES CONTENT (MG G⁻¹) (DAIDZEIN AND GENISTEIN) DURING SOYBEANS PLUS BROWN RICE FERMENTATION



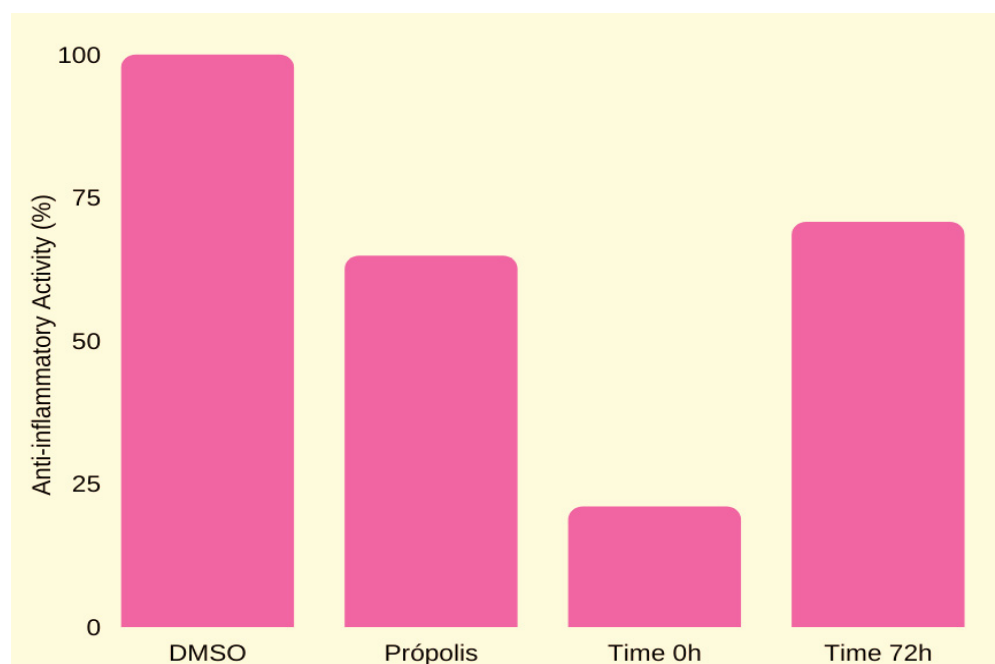
SOURCE: The autor (2022).

5.4 ANTI-INFLAMMATORY ACTIVITY

The anti-inflammatory activity of the *R. oligosporus* NRRL 2710 ethanolic extract was assessed by hyaluronidase inhibitory activity, these enzyme acts on the extracellular matrix of the tissues, increasing the permeability and facilitating the access of pathogens involved in inflammatory processes (EL-SAFORY; FAZARY; LEE, 2010; GIRISH; KEMPARAJU, 2007; SGARIGLIA *et al.*, 2013).

The results obtained shown the fermentation process increased hyaluronidase inhibitory activity 3.36 times when compared to unfermented soybeans (70.75% and 21.03% inhibition capacity for fermented and unfermented soybeans, respectively). In addition, showed significant higher activity than propolis, a commercial anti-inflammatory product(Figure 6).

FIGURE 5. ANTI-INFLAMMATORY ACTIVITY OF ALCOHOLIC EXTRACT OF THE *R. OLIGOSPORUS* NRRL 2710 CULTIVATED IN SOYBEAN SUPPLEMENTED WITH BROWN RICE BY 72 H.

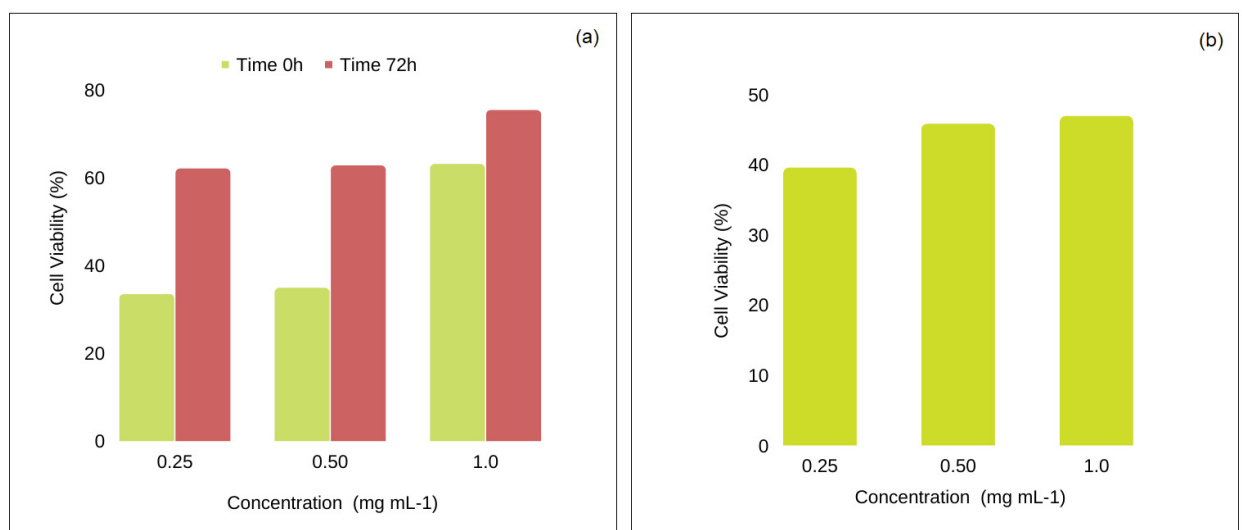


*DMSO: positive control capable of completely inhibiting the enzyme hyaluronidase (HIA);
 Propolis: commercial anti-inflammatory
 SOURCE: The autor (2022).

5.5 CYTOTOXICITY ASSESSMENT FOR TUMOR (CACO-2) AND NORMAL (MCF-10) CELLS

The decrease of viable cells at levels below 50% demonstrated by the values of cytotoxicity assessment for both unfermented soybeans (0h) and fermented substrate (72 h) showed cytotoxicity for normal cells (MCF-10). According to statistical analysis, these extracts presented no significant difference at the 95% level ($p < 0.05$). Considering this results, a cell viability study was carried out with CaCo-2 cancer cell to evaluate the cytotoxicity of the 72h *R. oligosporus* NRRL 2710 fermented ethanolic extract at different concentrations (Figure 7b). The extracts presented percentage of cellular viability between 39.53 and 46.89%.

FIGURE 6. CYTOTOXICITY ASSESSMENT OF ALCOHOLIC EXTRACT OF THE *R. OLIGOSPORUS* NRRL 2710 CULTIVATED IN SOYBEAN SUPPLEMENTED WITH BROWN RICE AGAINST (a) NORMAL CELLS (MCF-10) AND (b) CACO-2 TUMOR CELLS.



SOURCE: The autor (2022).

The inhibitory concentration values (IC_{50}) were 0.27 mg mL^{-1} for CaCo-2 cells and 0.79 mg mL^{-1} for normal MCF-10 cells, indicating a higher cytotoxic effect against carcinogenic cell. A lower percentage of cancer cell viability was observed at lower concentrations, showing that high concentrations are not required to attain cancer cell death. According to the statistical analysis, the extracts at different processing times showed a significant difference at the 95% level ($p < 0.05$), suggesting that the *R. oligosporus* NRRL 2710 SB+BR fermentation process was efficient; the lower value of cell viability of the cancer cells was obtained after 72 h of fermentation (39.53%).

Several studies show the inverse association between the consumption of soy products and the risk of degenerative diseases, such as cancer (SÁNCHEZ-CHINO *et al.*, 2015; ZHU *et al.*, 2015). The chemopreventive effects of soy are attributed to its bioactive molecules, such as isoflavones, which interact with the metabolic pathways responsible for controlling cell growth, proliferation and differentiation (DE MEJIA; DIA, 2010; DIA; DE MEJIA, 2011; MONTALES *et al.*, 2015; TSAI *et al.*, 2010).

5.6 SOXHLET EXTRACTION

The identification and quantification of bioactive compounds in natural sources can be performed by extraction with solvents of different polarities.

In this study, the results from soxhlet extraction with different organic solvents showed an increase in yield as the polarity of the solvent increased. The highest extraction yields were 44.10% and 45.24%, for ethanol and methanol, respectively, suggesting that the compounds present in the matrix studied have intermediate to high polarity (Table 6). Statistical data show that these values did not differ significantly ($p > 0.05$).

TABLE 5 - RESULTS OF EXTRACTION YIELD (%) OF THE SOXHLET EXTRACTION PROCESS WITH FERMENTED SOYBEAN DRIED AND WHOLE (WITHOUT DRYING) SAMPLES USING DIFFERENT SOLVENTS.

Run	Solvents	T (°C)*	Polarity	Extraction Yield (wt%)	
				Dry sample	Wet sample
S1	Hexane	68.0	0.0	15.56±4.02 ^c	-
S2	Petroleum ether	34.61	0.1	29.05±0.51 ^a	-
S3	Ethyl acetate	77.5	4.3	32.04±2.14 ^{ab}	-
S4	Acetone	56.0	5.4	34.07±0.68 ^{ab}	19.76±1.05 ^{ab}
S5	Ethanol	78.5	5.2	44.10±1.99 ^b	25.48±4.25 ^{bc}
S6	Methanol	64.7	6.6	45.24±3.70 ^b	32.84±3.70 ^c
S7	Water	100.0	10.2	23.66±4.32 ^a	14.20±2.82 ^a

* Boiling point of solvents. Means followed by the same letter do not differ statistically ($p > 0.05$)

SOURCE: The autor (2022).

The moisture levels in the raw material influence the extraction kinetics and yield since water is an interfering factor in the penetration of the solvent into the solid matrix and in the diffusion of the oil. Extractions with solvents from medium to high polarity were performed using dry sample; the results in Table 1 show yields of 25.48% and 32.84% for ethanol and methanol, respectively. It was observed that the drying process allowed a significant increase of up to 18.62% in extraction yield using ethanol. Previous studies also reported the optimization of extraction with lower moisture content, stating that the drying process facilitates the contact between the solvent and the solute to be extracted, resulting in higher yields (PIGHINELLI *et al.*, 2009; SANTOS *et al.*, 2013; TANGO; CARVALHO; SOARES, 2004).

The results in Table 6 show yields of 25.48% and 32.84% for ethanol and methanol, respectively. It was observed that the drying process allowed a significant increase of up to 18.62% in extraction yield using ethanol. Previous studies also reported the optimization of extraction with lower moisture content, stating that the drying process facilitates the contact between the solvent and the solute to be extracted, resulting in higher yields (PIGHINELLI *et al.*, 2009; SANTOS *et al.*, 2013; TANGO; CARVALHO; SOARES, 2004).

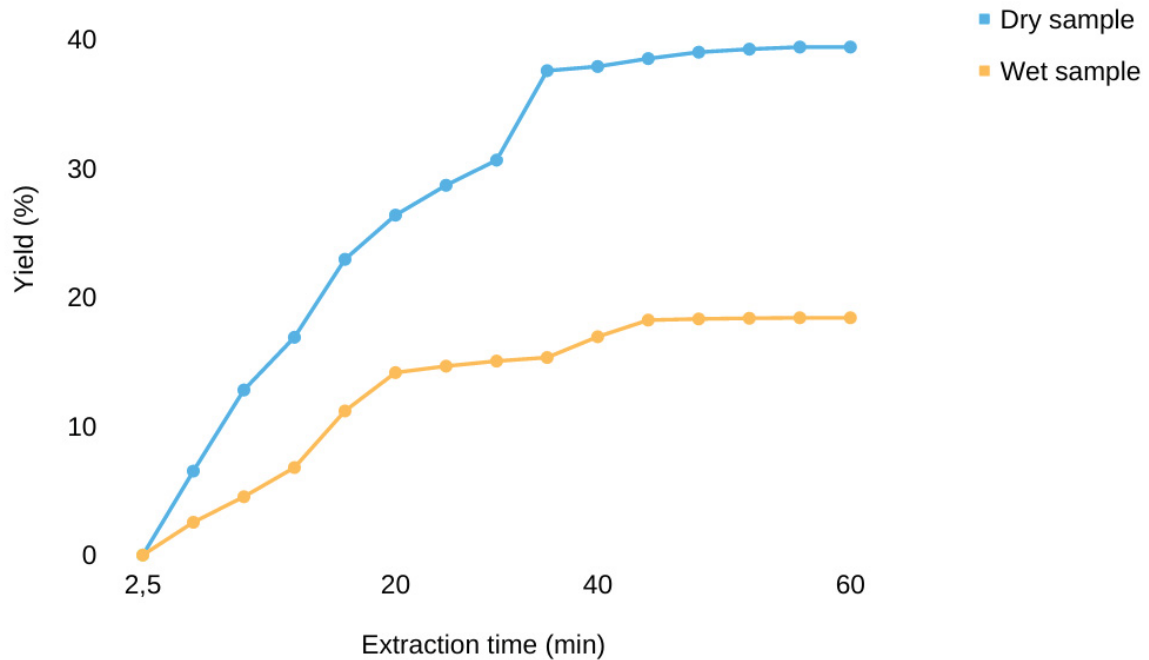
Water as a solvent in soxhlet extraction results in a higher yield. Although, the use of the high boiling temperature can lead to the degradation of bioactive compounds, such as polyphenols (BAHRIN *et al.*, 2018). Ethanol, besides providing greater extraction among the organic solvents due to its high polarity, also comprises low cost, lower toxicity and risks to human health and the environment. This way ethanol becomes more desirable than methanol for applications and was, therefore, selected as the liquid solvent for further extraction procedures.

5.7 EXTRACTION BY COMPRESSED SOLVENTS

For comparative purposes of the extraction techniques, supercritical CO₂ extractions were performed. Initially, an experiment was performed using supercritical CO₂ at 80 °C and 25 MPa, in which the extraction yield obtained was extremely low (<1%). Therefore, we chose to perform all additional extractions using the combined solvents scCO₂ + ethanol. For to promote and improve the contact between the solvent and the matrix before the compressed solvent extractions a preliminary pretreatment of the feedstock with ethanol was performed. After some tests, it was defined that 60-minute of contact was a suitable condition for this pretreatment. Raw material-ethanol contact can alter the matrix by swelling, thereby facilitating the analytes transport from interstitial pores to the surface and then to the bulk phase (TIM *et al.*, 1998).

A preliminary test using the described conditions was performed with dry and wet samples to evaluate the influence of humidity on scCO₂ + EtOH extraction. This same behavior was observed in the results obtained by Soxhlet extraction, in this case the drying process of the samples positively influenced the extraction yield resulting in an increase of 2.14 times in the procedure performed with dried samples.

FIGURE 7 - KINETICS OF DRY AND WET EXTRACTION OF FERMENTED SOYBEANS WITH CO₂ + ETOH AT A CONSTANT TEMPERATURE OF 80 °C AND PRESSURE OF 25 MPA EXPRESSED



SOURCE: The autor (2022).

A factorial design 2² with a central point was performed to verify the influence of the process parameters, such as pressure and temperature. The results obtained are shown in Table 7.

TABLE 6. EXPERIMENTAL CONDITIONS AND EXTRACTION YIELD USING SCCO₂ + ETOH AS SOLVENT CONSIDERING DRIED FERMENTED SOYBEAN RAW MATERIAL.

Run (M _{RM})	Solvent	P (MPa)	T (°C)	Time (min)	Extraction yield (wt%)
1	CO ₂ + EtOH	15	40	75	30.81
2	CO ₂ + EtOH	15	80	75	27.56
3	CO ₂ + EtOH	25	40	75	30.65
4	CO ₂ + EtOH	25	80	75	42.87
5	CO ₂ + EtOH	20	60	75	29.32±1.78

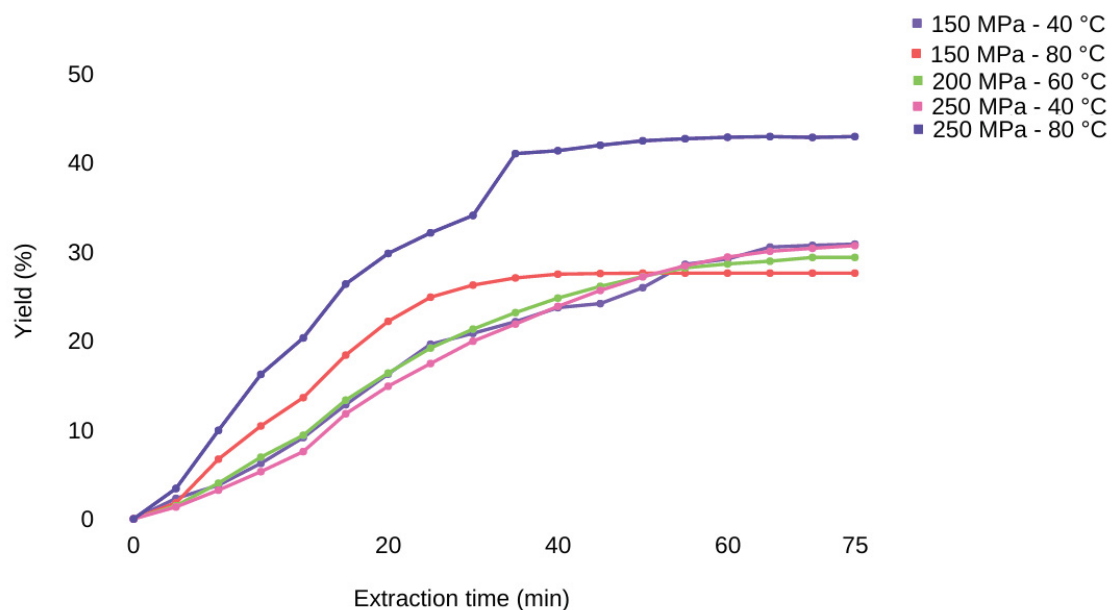
SOURCE: The autor (2022).

All scCO₂ + ethanol experiments were performed using static extraction time, called confinement period, a 60-minute. The highest extraction yield (42.87 wt%) was

obtained at the highest condition of pressure and temperature, 25 MPa and 80 °C. The comparison of runs 2 and 4 showed that changing the pressure, at 80°C, increased the yield to 15 percentual points (p.p.). Varying the temperature at a fixed pressure of 25 MPa, an increase in yield by 12 p.p. was observed. In general, the addition of ethanol as polarity modifier to scCO₂ showed to be an efficient approach to lead the extraction yields to levels compared to extraction in Soxhlet (Table 6).

The general extraction curves are provided and discussed in Figure 9 to better understanding the extraction using scCO₂ + EtOH. The operational parameters studied affect significantly the yields obtained. The parameters pressure and temperature had a positive effect on the yield because, at higher levels (80 °C and 25 MPa), they allowed higher yield values. The values obtained at different conditions presented a significant difference ($p > 0.05$).

FIGURE 8 - OVERALL EXTRACTION CURVES FOR DRIED FERMENTED SOYBEAN USING SCCO₂ + ETOH AS SOLVENT.



SOURCE: The autor (2022).

The lowest yield (27.56%) can be observed when using the lowest pressure (15 MPa) and highest temperature (80 °C). Pressure is the parameter that positively influences the yield of scCO₂ so that, its increasing reflects on the increase of density, viscosity and solvation power of CO₂ (AHANGARI; SARGOLZAEI, 2013; AKGÜN *et al.*, 2014). Therefore, the lower yield is probably a reflection of the low solubility of the extract since the lower CO₂ density results in low mass transfer rates during extraction (JOKIĆ *et al.*, 2011; MESOMO *et al.*, 2012).

The overall extraction curves in Figure 9, reveal that up to 40 minutes there were high initial extraction rates and a slope of the curve in the subsequent minutes. This indicates that there was first a period of dynamic extraction rate, where the outer surface of the particles is covered with the easily accessible and available solute. Then, the scCO₂ stops operating as a carrier and starts operating as an extraction agent and the mass transfer occurs by diffusion within the solute particles, resulting in the extraction rate dropping (JUCHEN *et al.*, 2019).

Among the methods mentioned, the Soxhlet extraction using ethanol and methanol as solvents showed a higher yield (44.10 and 45.24%, respectively) compared to scCO₂, where the maximum yield was obtained under different extraction conditions was 42.87%. However, scCO₂ extraction offers an advantage less time consuming than Soxhlet extraction.

5.8 FATTY ACIDS PROFILE

Table 8 shows the fatty acid (FA) profiles for the fermented soybean oil samples obtained from the different methods studied. The FA profiles have presented similarities, despite the different extraction conditions and solvents used.

TABLE 7 - FATTY ACID COMPOSITION OF FERMENTED SOYBEAN EXTRACTS OBTAINED FOR SOXHLET EXTRACTION USING DIFFERENT SOLVENTS AND SCCO₂+ETOH UNDER DIFFERENT CONDITIONS.

Run	Extraction condition	Composition (%)						
		Palmitic (C16:0)	Stearic (C18:0)	Oleic (C18:1)	α -Linolenic (C18:3)	Linoleic (C18:2)	Gondoic (C20:1)	Tricosanoic (C23:0)
S1	Hexane	11.35 ^d	4.33 ^b	31.86 ^e	5.47 ^c	46.32 ^f	-	0.67 ^a
S2	Petroleum ether	12.11 ^d	4.49 ^b	30.75 ^e	5.49 ^c	46.04 ^f	0.51 ^a	0.61 ^a
S3	Ethyl acetate	11.53 ^d	4.17 ^b	30.94 ^e	5.62 ^c	46.52 ^f	0.59 ^a	0.62 ^a
S4	Acetone	11.58 ^d	4.24 ^b	30.83 ^e	5.59 ^c	46.66 ^f	0.45 ^a	0.64 ^a
S5	Ethanol	13.27 ^d	4.88 ^b	29.96 ^e	5.23 ^c	45.22 ^f	0.62 ^a	0.81 ^a
S6	Methanol	12.37 ^d	4.50 ^b	30.21 ^e	5.64 ^c	46.14 ^f	0.45 ^a	0.69 ^a
S7	Water	14.68 ^a	-	33.41 ^b	-	51.92 ^c	-	-
scCO₂+EtOH								
1	15 MPa:40°C	11.27 ^d	4.57 ^c	37.34 ^e	4.27 ^b	41.45 ^f	0.51 ^a	0.59 ^a
2	15 MPa:80°C	11.28 ^d	4.69 ^c	37.24 ^e	4.17 ^b	41.38 ^f	0.59 ^a	0.64 ^a
3	25 MPa:40°C	10.98 ^d	4.55 ^c	37.25 ^e	4.26 ^b	41.93 ^f	0.50 ^a	0.54 ^a
4	25 MPa:80°C	11.31 ^d	4.63 ^c	37.17 ^e	4.23 ^b	41.42 ^f	0.55 ^a	0.68 ^a
5	20 MPa:60°C	11.36 ^d	4.71 ^c	37.13 ^e	4.27 ^b	41.37 ^f	0.56 ^a	0.61 ^a

Different superscripts indicate a significant difference ($p < 0.05$).

SOURCE: The autor (2022).

The results showed the main fatty acids present in all oil samples were linoleic acid and oleic acid. Ranged from 45.22 to 51.92%, and 29.96 to 33.41% respectively, in the extracts obtained by Soxhlet. The content of unsaturated fatty acids (C18:3, C18:1 and C18:2) account for about 82.70 of total fatty acids present in fermented soybean oil, such values suggest these as an attractive option in the functional food market.

Comparing the fatty acid profile of the oils obtained in this study with the fatty acid profile of the soybean oil reported *in nature*, an increase of 2.31% in oleic acid content and 7.7% in unsaturated fatty acid content is observed. Therefore, it is not

possible to state that the fermentative process had a positive influence on the composition of FA.

In a study by (MAN *et al.*, 2013), the fatty acid profile of soybean oil *in natura* of the analyzes performed by Soxhlet with *n*-hexane as solvent showed a predominant value of linoleic acid (range 49.56–55.83%), followed by oleic acid (21.77–26.55%) and palmitic acid (10.50–11.82%). In addition, the content of unsaturated fatty acids was approximately 75%. A comparison between soybean of different cultivars, tegument colors and years of harvest showed that the results in the fatty acid content did not present significant differences.

In a study that aimed to analyze fluctuations in the composition of soybean fatty acids over two years of storage at room temperature, the fatty acid profile obtained ranged from 49.2–56.8% for linoleic acid, 19.7–23.1% for oleic acid and 9.4–12.4% for palmitic acid, still presenting a total average content of unsaturated fatty acids > 75% (LEE; CHO, 2012).

(TIAN *et al.*, 2017) performed a comparative analysis of fatty acid composition between unfermented and fermented soybean extracts extracted by SC-CO₂. While the oleic acid content decreased from 22.35% to 18.59% after the fermentation process, the levels of palmitic acid and α -linolenic acid showed an increase of 12.13% to 15.41% and 7.30% to 8.43%, respectively. Comparing this mentioned study with the present work, it is not possible to state that the fermentation process and the addition of ethanol as a co-solvent are factors that affect the fatty acid profile of the extracts.

Fatty acids are key components of phospholipids in cell membranes playing important roles in various cellular functions, including metabolism and immune responses (Magnan, Levin & Luquet, 2015). The inclusion of oleic acid in the diet has been indicated because of its protective effects against cardiovascular and

neurodegenerative diseases. Linoleic acid, characteristic of soybeans and present in most of the oils obtained, has gained prominence in recent decades due to its biological and physiological benefits, including anti-carcinogenesis (MORAES *et al.*, 2017), anti-obesity (OLSON *et al.*, 2017), antidiabetic (YUCE *et al.*, 2016) as well as bone formation promoting properties (KIM; PARK; PARK, 2014).

5.9 TOTAL PHENOLIC CONTENT (TPC) AND ANTIOXIDANT POTENTIAL

Table 9 presents the total phenolic content (TPC) and antioxidant potential for both extraction techniques.

In Soxhlet extraction, the values ranged from $132.10 \pm 0.01 \mu\text{g GAE g}^{-1}$ to $1305.6 \pm 0.03 \mu\text{g GAE g}^{-1}$. The best results were obtained from fermented soybeans using solvents with higher polarity, as ethanol and water, $962.9 \mu\text{g GAE g}^{-1} \pm 0.02$ and $1305.6 \mu\text{g GAE g}^{-1} \pm 0.03$, respectively. The results highlight a significant difference ($p < 0.05$). This shows that the polarity of the solvent is a determining factor for obtaining phenolic compounds in the extract, pointing out that higher polarity extracts a greater amount of polar compounds.

The quantification of total phenolic compounds contained in the extracts obtained by $\text{scCO}_2 + \text{EtOH}$ ranged from $1058.2 \pm 0.01 \mu\text{g GAE g}^{-1}$ to $1391.9 \pm 0.09 \mu\text{g GAE g}^{-1}$ (Table 4), with statistical difference ($p > 0.05$).

High pressure and the temperature was the most efficient condition in the extraction of such compounds ($1391.9 \mu\text{g GAE g}^{-1}$), revealing the pressure had a positive influence on the extraction since a higher content of total phenolics is observed in the extracts of runs 3 and 4 (25MPa).

The obtained TPC values reveal the extracts as beneficial for human health since, according to the regulation established by EU no. 432/2012, the limit of 0.25 mg phenolics per g of oil is set to declare benefits (SQUEO *et al.*, 2019).

TABLE 8 - RESULTS OF PHENOLIC COMPOUNDS ($\mu\text{g GAE G}^{-1}$), ANTIOXIDANT ACTIVITY EXPRESSED BY INHIBITORY CONCENTRATION (IC_{50}), RADICAL SCAVENGING ACTIVITY BY DPPH (%) AND CAPACITY EQUIVALENT TO THE RADICAL TROLOX ($\mu\text{M TROLOX/G}$) OF THE SOXHLET AND $\text{scCO}_2 + \text{EtOH}$ EXTRACTION

Run	Extraction condition	Total Phenolic compounds ($\mu\text{g GAE g}^{-1}$)	Antioxidant Activity		
			IC_{50} (g g^{-1})	Inhibition DPPH (%)	$\mu\text{m Trolox/g}$
	Soxhlet				
S1	Hexane	132.1 \pm 0.02 ^a	2.91 \pm 0.47 ^a	18.25 \pm 0.99 ^a	*
S2	Petroleum ether	423.8 \pm 0.02 ^b	1.75 \pm 1.65 ^e	29.68 \pm 0.74 ^b	*
S3	Ethyl acetate	581.7 \pm 0.11 ^d	2.93 \pm 0.84 ^a	17.53 \pm 0.20 ^a	37.48 \pm 0.01 ^a
S4	Acetone	904.7 \pm 0.03 ^e	0.79 \pm 0.27 ^c	61.28 \pm 0.51 ^f	616.0 \pm 0.01 ^d
S5	Ethanol	962.9 \pm 0.023 ^f	1.57 \pm 1.20 ^d	38.97 \pm 0.83 ^c	200.44 \pm 0.02 ^b
S6	Methanol	446.0 \pm 0.06 ^c	1.82 \pm 0.76 ^f	43.12 \pm 0.92 ^d	303.40 \pm 0.03 ^c
S7	Water	1305.6 \pm 0.03 ^g	0.55 \pm 0.84 ^b	81.27 \pm 0.77 ^g	962.66 \pm 0.02 ^e
	scCO₂+EtOH				
1	15 MPa:40°C	1221.6 \pm 0.075 ^a	0.28 \pm 0.73 ^c	89.66 \pm 0.74 ^b	976.74 \pm 0.01 ^d
2	15 MPa:80°C	1158.2 \pm 0.01 ^{ab}	0.34 \pm 1.25 ^d	84.60 \pm 0.29 ^a	848.59 \pm 0.02 ^a
3	25 MPa:40°C	1249.0 \pm 0.01 ^a	0.18 \pm 2.31 ^b	93.62 \pm 0.37 ^c	903.41 \pm 0.01 ^c
4	25 MPa:80°C	1391.9 \pm 0.09 ^c	0.17 \pm 0.79 ^a	94.09 \pm 0.32 ^c	984.89 \pm 0.01 ^e
5	20 MPa:60°C	1058.9 \pm 0.02 ^a	0.34 \pm 1.20 ^d	89.94 \pm 0.24 ^b	884.89 \pm 0.01 ^b

Different superscripts indicate a significant difference ($p < 0.05$).

* The values found were outside the confidence interval studied, so they are not shown.

SOURCE: The autor (2022).

According to the values reported in Table 9, in Soxhlet extraction, values vary from $2.93 \pm 0.47 \text{ g g}^{-1}$ to $0.55 \pm 0.84 \text{ g g}^{-1}$. The extract with the highest antioxidant potential (water), showed a DPPH radical inhibition potential of 81.27% and $962.66 \mu\text{m Trolox g}^{-1}$. This behavior showed that the solvents of intermediate and high polarity used in the extractions favored the solubilization of compounds with antioxidant activity, detectable by the DPPH method.

The extracts obtained at high-temperature conditions (80 °C) and pressure (25 MPa) in $\text{scCO}_2 + \text{EtOH}$ showed highest antioxidant activity, corresponding to an IC_{50} of 0.17 g g^{-1} and $984 \mu\text{m Trolox g}^{-1}$, which were able to inhibit 94.09% of the DPPH

radical, significantly higher than other treatments ($p < 0.05$), confirming the correlation between the phenolic compounds and the antioxidant activity, already reported by other authors (VEBER *et al.*, 2015). Analyzing these results, it can be stated that no tendency was observed in the antioxidant activity concerning the pressure or temperature for any of the extracts obtained.

In terms of comparison among the different extractions techniques, $scCO_2 + EtOH$ demonstrated to be a viable approach, allowing the extraction of higher quality extracts when compared to those obtained by Soxhlet extraction with an organic solvent. Comparing the extracts obtained from fermented soybean using $scCO_2 + EtOH$, the antioxidant potential was 3.23 times higher than the extracts obtained by Soxhlet. Given these results, further analysis in this study was focused on the extracts obtained by $sCO_2 + EtOH$.

5.10 HIGH PERFORMANCE LIQUID CHROMATOGRAPHY OF PHENOL CONTENTS (HPLC)

The chromatographic analysis revealed the presence of four phenolic compounds, identified in all the extracts tested, being gallic acid (Benzyl 7-hydroxy-2,2-diphenyl-1,3-benzodioxole-5-carboxylate), transcinamic acid ((E)-3-phenylprop-2-enoic acid), genistein (5,7-dihydroxy-3-(4-hydroxyphenyl)chromen-4-one) and daidzein (7-hydroxy-3-(4-hydroxyphenyl)chromen-4-one), with variations in retention time (R_t) (Table 10).

TABLE 9 - RESULTS OF IDENTIFICATION OF PHENOLIC CONTENT BY HIGH PERFORMANCE LIQUID CHROMATOGRAPHY ON THE EXTRACTS OBTAINED BY $SCO_2 + EtOH$.

Phenolics compounds identified ($\mu g g^{-1}$)

Different superscripts indicate a significant difference ($p < 0.05$).

Run (M _{RM})	P (MPa)	T (°C)	Rt (min)	Gallic acid	Rt (min)	Transcinnamic acid	Rt (min)	Genistein	Rt (min)	Daidzein
1	15	40	3.46	416.3 ^a	25.46	518.3 ^b	26.06	593.1 ^a	24.59	606.1 ^c
2	15	80	3.42	449.5 ^b	25.37	521.4 ^c	25.94	625.3 ^d	24.50	605.0 ^c
3	25	40	3.42	414.5 ^a	25.36	507.0 ^a	25.94	596.5 ^b	24.50	577.3 ^a
4	25	80	3.42	457.2 ^c	25.38	518.1 ^b	25.95	621.7 ^c	24.50	603.9 ^c
5	20	60	3.42	417.7 ^a	25.38	518.7 ^b	25.95	597.6 ^b	24.51	565.9 ^b

SOURCE: The autor (2022).

Phenolic acids are non-flavonoid compounds that can be divided by their chemical structure (Sanja, Nikoli, Lukovi, Jovanovi, & Stefanovi, 2018). The chromatographic analysis revealed the presence of benzoic acid derivatives, such as gallic acid, and derivatives of cinnamic acid, such as transcinnamic acid. The values of gallic acid ranged from 414.5 to 457.2 $\mu\text{g g}^{-1}$ among the extracts obtained by different conditions, and transcinnamic acid ranged from 518.1 to 521.4 $\mu\text{g g}^{-1}$.

The presence of aglicone isoflavones, such as daidzein and genistein, has also been observed. The values obtained vary from 565.9 to 606.1 $\mu\text{g g}^{-1}$ of daidzein and 593.1 to 625.3 $\mu\text{g g}^{-1}$ of genistein. (Kim et al., 2016) a study of the presence of bioactive compounds in soybean reported daidzein values of 77.18 $\mu\text{g g}^{-1}$ and genistein 332.02 $\mu\text{g g}^{-1}$. (Santos et al., 2018) in studies using fermented soybean by-product (okara) by *Saccharomyces cerevisiae* also reported the presence of aglycone isoflavones. The presence of these compounds is expected since the fermentation of grains by microorganisms that produce β -glucosidase, such as *Rhizopus spp.*, promotes the biotransformation of glycosidic isoflavones.

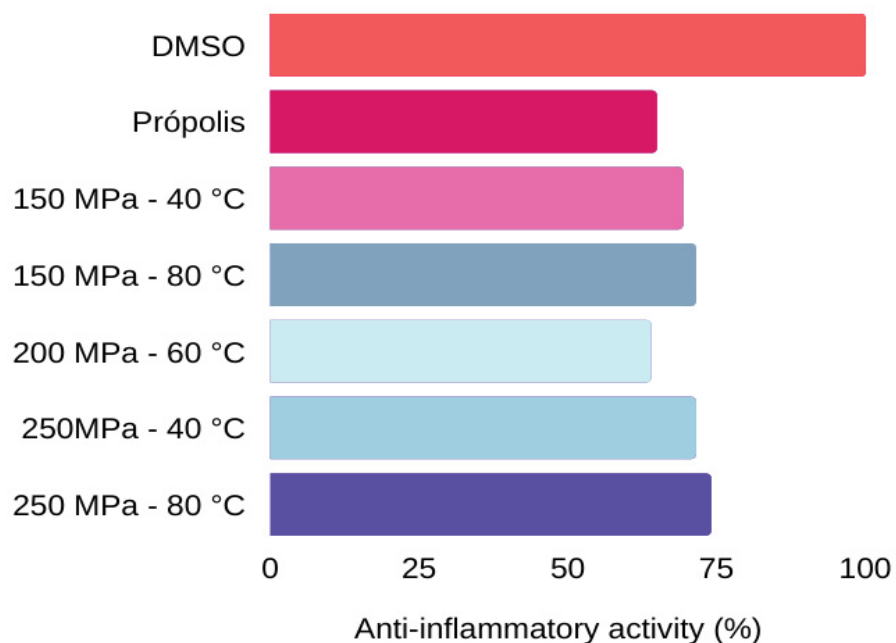
5.11 EVALUATION OF ANTI-INFLAMMATORY ACTIVITY IN VITRO

The anti-inflammatory potential of the extracts obtained by the scCO₂ + EtOH method under different conditions was evaluated by the inhibitory activity of the hyaluronidase enzyme (HIA), this is directly involved in inflammatory processes. HIA

acts on the extracellular matrix of the tissues, increasing permeability and facilitating the access of pathogens (GIRISH; KEMPARAJU, 2007).

The extracts showed potential for inhibition of the hyaluronidase enzyme in the range of 63.83% to 74.0%. These results were compared with the dimethyl sulfoxide control (DMSO), since it has a complete HIA inhibition capacity and with the propolis control, a commercial anti-inflammatory. The samples showed a significant difference ($p < 0.05$) (Figure 10). Comparing than the tested commercial control, which was able to inhibit 64.85%, the extracts obtained by $scCO_2 + EtOH$ showed 9.15% greater anti-inflammatory potential. It is possible to analyze that, as with the previous results, the extract with the highest inhibitory capacity of the hyaluronidase enzyme was obtained at conditions of high pressure and temperature (25 MPa and 80 °C).

FIGURE 9 - DETERMINATION OF THE ANTI-INFLAMMATORY ACTIVITY OF THE SAMPLES EXTRACTED BY $scCO_2 + EtOH$ UNDER DIFFERENT CONDITIONS OF TEMPERATURE AND



DMSO: positive control capable of completely inhibiting the enzyme hyaluronidase (HIA).
Própolis: commercial anti-inflammatory.

SOURCE: The autor (2022).

In an early study, mentioned in item 5.4, investigated the anti-inflammatory activity of fermented and unfermented soybean extracts. The results showed that the alcoholic extract obtained in the 72-hour fermentation process showed a maximum inhibition capacity of 70.75%, corresponding to an increase in activity of 3.36 times compared to the "in nature" extract (21.03%) (PRADO *et al.*, 2021). The extracts obtained by scCO₂ + EtOH have presented higher anti-inflammatory activity in 4%.

The anti-inflammatory potential of the tested extracts may be associated with the presence of total phenolic compounds in them. Several studies have demonstrated that plant-derived polyphenols, in particular, flavonoids have potential anti-inflammatory activity *in vitro* and *in vivo* (DOUBLE-BLIND *et al.*, 2015). Isoflavones, found mainly in soybean, are especially studied because they have acted as an anti-inflammatory agent due to the fact that genistein negatively regulates the signal transduction events induced by cytokines in the cells of the immune system (VERDRENGH *et al.*, 2003).

6 CONCLUSIONS

It can be concluded that in the interest of preventive and therapeutic strategies the future of fermented foods is quite optimistic. Natural antioxidants consist of one of the main components with beneficial effects in the prevention of many diseases caused by cellular oxidative processes, and reactive oxygen species. Oxidative stress (OS) is intimately related to a series of chronic diseases and metabolic imbalance. In this context, interest arises in fermented soy foods, which have bioactive compounds such as flavonoids, isoflavone, peptides, and soy proteins. Soy aglucone isoflavones are antioxidant compounds and their activities are associated with the ability to eliminate

reactive oxygen species (ROS). This study reported that soybean supplementation with brown rice is an important strategy to improve the biosynthesis of antioxidant compounds by SSF and the fermentation process was effective in the accumulation of myricetin, gallic acid, ferrulic acid, kaempferol and increased accumulation of trans-cinnamic acid. This fermentation process point out to accumulation of cinnamic acid, which can be used as a novel, alternative way for production of this antitumor compound. The results presented showed that the use of solvents with greater polarity culminates in higher extraction yields of bioactive compounds with antioxidant power, rich in phenolic compounds. The extraction efficiency using scCO₂ + EtOH was related to greater pressure and fixed temperature (25 MPa and 80 °C). The total phenolic content and the antioxidant capacity of the extracts showed present an important correlation. The extracts obtained by the supercritical fluid showed anti-inflammatory activity with a high potential for inhibition of the hyaluronidase enzyme (74%). In addition, important bioactive compounds were detected, including gallic acid and trans-cinnamic acid. Extraction with scCO₂ + EtOH allowed a significant improvement in soybean extractions fermented with *Rhizopus* spp., demonstrating the technical feasibility of the process and allowing the recovery of molecules with high bioactive potential for the pharmaceutical and food industries.

6.1 RECOMMENDATIONS FOR FUTURE STUDIES

As a suggestion for further studies, investigation of the actual compounds responsible for the anti-inflammatory and anti-tumor activities is needed. So is the purification of these compounds of interest. A thorough investigation of the other compounds present in fermented soy foods is needed, so as to be clear about their relationship with the bioactive properties.

Also, further in vivo studies should be conducted for the specification of these biologically active principles.

REFERENCES

- ABE, S.K. *et al.* Comparison between the impact of fermented and unfermented soy intake on the risk of liver cancer: the JPHC Study. **European Journal of Nutrition**, [s. l.], v. 60, n. 3, p. 1389–1401, 2021. Disponível em: <https://doi.org/10.1007/s00394-020-02335-9>.
- ADEM, Sevki *et al.* Identification of Potent COVID-19 Main Protease (Mpro) Inhibitors from Natural Polyphenols: An in Silico Strategy Unveils a Hope against CORONA. [s. l.], n. March, 2020. Disponível em: <https://doi.org/10.20944/preprints202003.0333.v1>.
- AHANGARI, Behdad; SARGOLZAEI, Javad. Extraction of lipids from spent coffee grounds using organic solvents and supercritical carbon dioxide. **Journal of Food Processing and Preservation**, [s. l.], v. 37, n. 5, p. 1014–1021, 2013.
- AHMAD, Aliya *et al.* Enhancement of β -secretase inhibition and antioxidant activities of tempeh, a fermented soybean cake through enrichment of bioactive aglycones. Enhancement of β -secretase inhibition and antioxidant activities of tempeh, a fermented soybean cake through. **Pharmaceutical Biology**, [s. l.], v. 53, n. 5, p. 758–766, 2015.
- AHMAD, Aliya *et al.* Total isoflavones from soybean and tempeh reversed scopolamine-induced amnesia, improved cholinergic activities and reduced neuroinflammation in brain. **FOOD AND CHEMICAL TOXICOLOGY**, [s. l.], v. 65, p. 120–128, 2014. Disponível em: <http://dx.doi.org/10.1016/j.fct.2013.12.025>.
- AHMED, Mohamed *et al.* Antioxidative and cytotoxic activity of essential oils and extracts of *Satureja montana* L., *Coriandrum sativum* L. and *Ocimum basilicum* L. obtained by supercritical fluid extraction. **The Journal of Supercritical Fluids**, [s. l.], v. 128, n. March, p. 128–137, 2017.
- AKGÜN, Nalan A. *et al.* Extraction behavior of lipids obtained from spent coffee grounds using supercritical carbon dioxide. **Chemical Engineering and Technology**, [s. l.], v. 37, n. 11, p. 1975–1981, 2014.
- ALEJANDRA, María *et al.* Solid state fermentation to obtain vegetable products bio-enriched with isoflavone aglycones using lactic cultures. **Revista Argentina de Microbiología**, [s. l.], v. 51, n. 3, p. 201–207, 2019. Disponível em: <https://doi.org/10.1016/j.ram.2018.04.006>.
- ALESSANDRA, Paula *et al.* Supercritical CO₂ oil extraction from *Bauhinia forficata* link subsp. *pruinosa* leaves: Composition, antioxidant activity and mathematical modeling. **The Journal of Supercritical Fluids**, [s. l.], v. 153, 2019.
- ALVAREZ, María V *et al.* Valorization of an agroindustrial soybean residue by supercritical fluid extraction of phytochemical compounds. **The Journal of Supercritical Fluids**, [s. l.], ano 143, n. March 2018, 2019. p. 90–96. Disponível em: <https://doi.org/10.1016/j.supflu.2018.07.012>.
- AMIN, Muhamad Nur Ghoyatul *et al.* Identification of a novel umami peptide in tempeh (Indonesian fermented soybean) and its binding mechanism to the umami receptor T1R. **Food Chemistry**, [s. l.], v. 333, n. July, p. 127411, 2020. Disponível em: <https://doi.org/10.1016/j.foodchem.2020.127411>.
- AOCS, D Firestone. Official methods and recommended practices of the American Oil Chemists' Society. **AOCS**, [s. l.], v. 5, p. 2–93, 1998.
- AOKI, Hideyuki *et al.* Comparison of bioactive components in tempeh produced by three different rhizopus starters and immunomodulatory effect of tempeh on atopic dermatitis mice. **Food Science and Technology Research**, [s. l.], v. 26, n. 5, p. 665–672, 2020.

- ASSEFA, Yared *et al.* Assessing Variation in US Soybean Seed Composition (Protein and Oil). **Frontiers in Plant Science**, [s. l.], v. 10, n. March, p. 298, 2019.
- ASSOCIATIONS BETWEEN INTAKE OF DIETARY FERMENTED SOY FOOD AND CONCENTRATIONS OF INFLAMMATORY MARKERS.PDF. [S. l.: s. n.], [s. d.].
- ASTUTI, Mary *et al.* Tempe, a nutritious and healthy food from Indonesia. **Asia Pacific Journal of Clinical Nutrition**, [s. l.], v. 9, n. 4, p. 322–325, 2000.
- BAHRIN, Nurlyana *et al.* Effect of Processing Temperature on Antioxidant Activity of Ficus carica Leaves Extract. **Journal of Science and Technology**, [s. l.], v. 10, n. 2, p. 99–103, 2018.
- BEHLOUL, Nouredine; WU, Guanzhong. Genistein : A promising therapeutic agent for obesity and diabetes treatment. **European Journal of Pharmacology**, [s. l.], v. 698, n. 1–3, p. 31–38, 2013.
- BELTRÁN-BARRIENTOS, L. M *et al.* Mechanistic Pathways Underlying the Antihypertensive Effect of Fermented Milk with Lactococcus lactis NRRL B-50571 in Spontaneously. **Nutrients**, [s. l.], v. 10, p. 262, 2018.
- BENEDETTI, Silvia *et al.* Antioxidant properties of tofu whey concentrate by freeze concentration and nanofiltration processes. **Journal of Food Engineering**, [s. l.], v. 160, p. 49–55, 2015. Disponível em: <http://dx.doi.org/10.1016/j.jfoodeng.2015.03.021>.
- BILIR, Birdal *et al.* Effects of genistein supplementation on genome - wide DNA methylation and gene expression in patients with localized prostate cancer. **international journal of oncology**, [s. l.], v. 51, p. 223–234, 2017.
- BOURDICHON, François *et al.* Food fermentations: Microorganisms with technological beneficial use. **International Journal of Food Microbiology**, [s. l.], v. 154, n. 3, p. 87–97, 2012. Disponível em: <http://dx.doi.org/10.1016/j.ijfoodmicro.2011.12.030>.
- BOUSQUET, Jean *et al.* Is diet partly responsible for differences in COVID - 19 death rates between and within countries ?. **Clin Transl Allergy**, [s. l.], v. 10, n. 1, p. 1–7, 2020.
- BRAND-WILLIAMS, W.; CUVELIER, M. E.; BERSET, C. Respostas Perceptivas E. **LWT - Food Science and Technology**, [s. l.], v. 28, n. 1, p. 25–30, 1995.
- CALDER, Philip C *et al.* Optimal nutritional status for a well-functioning immune system is an important factor to protect against viral infections. **Nutrients**, [s. l.], v. 12, n. 4, 2020. Disponível em: www.mdpi.com/journal/nutrients.
- CALVELLO, Rosa *et al.* Bovine and soybean milk bioactive compounds : Effects on inflammatory response of human intestinal Caco-2 cells. **Food Chemistry**, [s. l.], v. 210, p. 276–285, 2016. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2016.04.067>.
- CAO, Zhen-hui *et al.* Bioactivity of soy-based fermented foods : A review. **Biotechnology Advances**, [s. l.], v. 37, n. December 2018, p. 223–238, 2019.
- CAO, Xiao-Hong *et al.* Evaluation of a lipopeptide biosurfactant from Bacillus natto TK-1 as a potential source of anti-adhesive, antimicrobial and antitumor activities. **Brazilian Journal of Microbiology**, [s. l.], v. 40, n. 2, p. 373–379, 2009.
- CDC, Centers for Disease Control and Prevention. **Food and Coronavirus Disease 2019**. [S. l.], 2020. Disponível em: <https://www.cdc.gov/coronavirus2019-ncovdaily-life-copingfood-and-COVID-19.html>. Acesso em: 3 ago. 2021.
- CHA, Youn-soo *et al.* Visceral fat and body weight are reduced in overweight adults by the supplementation of Doenjang , a fermented soybean paste. **Nutrition Research and Practice**, [s. l.], v. 6, n. 6, p. 520–526, 2012.
- CHAN, E.W.C. *et al.* Natto and miso: an overview on their preparation, bioactive components and health-promoting effects. **Food Research**, [s. l.], v. 5, n. June, p. 446–452, 2021.

- CHAN, Yin-ching *et al.* Tempeh attenuates cognitive deficit, antioxidant imbalance, and amyloid β of senescence-accelerated mice by modulating Nrf2 expression via MAPK pathway. **Journal of Functional Foods**, [s. l.], v. 50, n. 200, p. 112–119, 2018. Disponível em: <https://doi.org/10.1016/j.jff.2018.09.023>.
- CHAN, Weng; LIU, Shao-quan. Biovalorisation of okara (soybean residue) for food and nutrition. **Trends in Food Science & Technology**, [s. l.], v. 52, p. 139–147, 2016. Disponível em: <http://dx.doi.org/10.1016/j.tifs.2016.04.011>.
- CHEN, Meinan *et al.* Association between soy isoflavone intake and breast cancer risk for pre- and post-menopausal women: A meta-analysis of epidemiological studies. **PLoS ONE**, [s. l.], v. 9, n. 2, 2014.
- CHEN, Lei *et al.* Intracellular signaling pathways of inflammation modulated by dietary flavonoids: The most recent evidence. **Critical Reviews in Food Science and Nutrition**, [s. l.], v. 0, n. 0, p. 1–17, 2018.
- CHEN, Tingtao *et al.* Molecular Identification of Microbial Community in Surface and Undersurface Douchi During Postfermentation. **Journal of Food Science**, [s. l.], v. 79, n. 4, p. 653–658, 2014.
- CHOI, Joo Hee *et al.* Cheonggukjang, a soybean paste fermented with *B. licheniformis*-67 prevents weight gain and improves glycemic control in high fat diet induced obese mice. **Journal of Clinical Biochemistry and Nutrition**, [s. l.], v. 59, n. 1, p. 31–38, 2016.
- CHOKSOMNGAM, Yanee *et al.* The metabolic role of spermidine in obesity: Evidence from cells to community. **Obesity Research and Clinical Practice**, [s. l.], v. 15, n. 4, p. 315–326, 2021. Disponível em: <https://doi.org/10.1016/j.orcp.2021.06.009>.
- CHOURASIA, Rounak *et al.* A Potential Peptide From Soy Cheese Produced Using *Lactobacillus delbrueckii* WS4 for Effective Inhibition of SARS-CoV-2 Main Protease and S1 Glycoprotein. **Frontiers in Molecular Biosciences**, [s. l.], v. 7, n. December, p. 1–12, 2020.
- CONAB. Acompanhamento da safra brasileira. [s. l.], v. 8, 2022. Disponível em: [file:///D:/fernanda agrop Prado/Downloads/E-book_BoletimZdeZSafraZ-Z3oZlevantamento \(1\).pdf](file:///D:/fernanda%20agrop Prado/Downloads/E-book_BoletimZdeZSafraZ-Z3oZlevantamento%20(1).pdf).
- CORREA, Madeline *et al.* Extraction of inflorescences of *Musa paradisiaca* L. using supercritical CO₂ and compressed propane. **The Journal of Supercritical Fluids**, [s. l.], v. 113, p. 128–135, 2016. Disponível em: <http://dx.doi.org/10.1016/j.supflu.2016.03.016>.
- COSTELA-RUIZ, Víctor J *et al.* SARS-CoV-2 infection: The role of cytokines in COVID-19 disease. **Cytokine and Growth Factor Reviews**, [s. l.], v. 54, n. June, p. 62–75, 2020. Disponível em: <https://doi.org/10.1016/j.cytogfr.2020.06.001>.
- CUI, Jingwen *et al.* A novel fermented soybean, inoculated with selected *Bacillus*, *Lactobacillus* and *Hansenula* strains, showed strong antioxidant and anti-fatigue potential activity. **Food Chemistry**, [s. l.], v. 333, n. July, 2020.
- CUI, Meizi *et al.* Antiobesity effects of kimchi in diet-induced obese mice. **Journal of Ethnic Foods**, [s. l.], v. 2, n. 3, p. 137–144, 2015. Disponível em: <http://dx.doi.org/10.1016/j.jef.2015.08.001>.
- DA SILVA, Leomar H.; CELEGHINI, Renata M.S.; CHANG, Yoon K. Effect of the fermentation of whole soybean flour on the conversion of isoflavones from glycosides to aglycones. **Food Chemistry**, [s. l.], v. 128, n. 3, p. 640–644, 2011. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2011.03.079>.
- DABBAGH, Fatemeh *et al.* Nattokinase: production and application. **Applied Microbiology and Biotechnology**, [s. l.], v. 98, n. 22, p. 9199–9206, 2014.
- DAI, Shuhong *et al.* Effects of Lactic Acid Bacteria-Fermented Soymilk on Isoflavone

- Metabolites and Short-Chain Fatty Acids Excretion and Their Modulating Effects on Gut Microbiota. **Journal of Food Science**, [s. l.], v. 84, n. 7, p. 1854–1863, 2019.
- DANDONA, Paresh; AJAY, Chaudhuri; SANDEEP, Dhindsa. Proinflammatory and Prothrombotic Effects. **Diabetes Care**, [s. l.], v. 33, n. 7, p. 1686–1687, 2010.
- DAS, Gitishree *et al.* Traditional fermented foods with anti-aging effect: A concentric review. **Food Research International**, [s. l.], v. 134, n. October 2019, p. 109269, 2020. Disponível em: <https://doi.org/10.1016/j.foodres.2020.109269>.
- DASTMALCHI, Narges *et al.* Antioxidants with two faces toward cancer. **Life Sciences**, [s. l.], v. 258, n. July, p. 118–186, 2020. Disponível em: <https://doi.org/10.1016/j.lfs.2020.118186>.
- DE LIMA, Daruichi Pereira *et al.* The impact of Chinese imports of soybean on port infrastructure in Brazil: A study based on the concept of the “Bullwhip Effect”. **Journal of Commodity Markets**, [s. l.], v. 9, n. October, p. 55–76, 2018. Disponível em: <https://doi.org/10.1016/j.jcomm.2017.11.001>.
- DE MEJIA, Elvira Gonzalez; DIA, Vermont P. The role of nutraceutical proteins and peptides in apoptosis, angiogenesis, and metastasis of cancer cells. **Cancer and Metastasis Reviews**, [s. l.], v. 29, n. 3, p. 511–528, 2010.
- DENG, Yi. Yu *et al.* Role of microglia in the process of inflammation in the hypoxic developing brain. **Frontiers in Bioscience**, [s. l.], v. 3, p. 884–900, 2011.
- DIA, Vermont P; BRINGE, Neal A; MEJIA, Elvira G De. Peptides in pepsin – pancreatin hydrolysates from commercially available soy products that inhibit lipopolysaccharide-induced inflammation in macrophages. **FOOD CHEMISTRY**, [s. l.], v. 152, p. 423–431, 2014. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2013.11.155>.
- DIA, Vermont P.; DE MEJIA, Elvira Gonzalez. Lunasin induces apoptosis and modifies the expression of genes associated with extracellular matrix and cell adhesion in human metastatic colon cancer cells. **Molecular Nutrition and Food Research**, [s. l.], v. 55, n. 4, p. 623–634, 2011.
- DIAS, Gisele Pereira *et al.* The Role of Dietary Polyphenols on Adult Hippocampal Neurogenesis: Molecular Mechanisms and Behavioural Effects on Depression and Anxiety. **Oxidative Medicine and Cellular Longevity**, [s. l.], v. 2012, 2012.
- DIFO, Harouna V *et al.* Chemical Changes during Open and Controlled Fermentation of Cowpea (*Vigna unguiculata*) Flour. **International Journal of Food Nutrition and Safety**, [s. l.], v. 5, n. 1, p. 1–10, 2014.
- DOUBLE-BLIND, Randomized *et al.* Supplementation of the Pure Flavonoids Epicatechin and Quercetin Affects Some Biomarkers of Endothelial Dysfunction and Inflammation in (Pre) Hypertensive Adults: A. [s. l.], n. C, p. 1–5, 2015.
- EISENBERG, Tobias *et al.* Induction of autophagy by spermidine promotes longevity. **Nature Cell Biology**, [s. l.], v. 11, n. 11, p. 1305–1314, 2009.
- EL-SAFORY, Nermeen S.; FAZARY, Ahmed E.; LEE, Cheng Kang. Hyaluronidases, a group of glycosidases: Current and future perspectives. **Carbohydrate Polymers**, [s. l.], v. 81, n. 2, p. 165–181, 2010. Disponível em: <http://dx.doi.org/10.1016/j.carbpol.2010.02.047>.
- FENG, Rong; CHEN, Liang; CHEN, Keping. Fermentation trip: amazing microbes, amazing metabolisms. [s. l.], p. 717–729, 2018.
- FLORINDO, Renata N *et al.* Structural insights into β -glucosidase transglycosylation based on biochemical, structural and computational analysis of two GH1 enzymes from *Trichoderma harzianum*. **New BIOTECHNOLOGY**, [s. l.], v. 40, n. November 2016, p. 218–227, 2018. Disponível em: <https://doi.org/10.1016/j.nbt.2017.08.012>.
- GAMAN, L; STOIAN, I; ATANASIU, V. Can ageing be slowed? Hormetic and redox perspectives. **Journal of Medicine and Life**, [s. l.], v. 4, n. 4, 2011.

- GARRETT, Rachael D; RAUSCH, Lisa L. Green for gold: social and ecological tradeoffs influencing the sustainability of the Brazilian soy industry. **The Journal of Peasant Studies**, [s. l.], v. 43, n. 2, p. 461–493, 2016.
- GIORDANO, Elena *et al.* Soy Isoflavones in Nutritionally Relevant Amounts Have Varied Nutrigenomic Effects on Adipose Tissue. **Molecules**, [s. l.], n. 1, p. 2310–2322, 2015.
- GIRISH, K. S.; KEMPARAJU, K. The magic glue hyaluronan and its eraser hyaluronidase: A biological overview. **Life Sciences**, [s. l.], v. 80, n. 21, p. 1921–1943, 2007.
- GOUDA, Ahmed S *et al.* Theoretical benefits of yogurt-derived bioactive peptides and probiotics in COVID-19 patients – A narrative review and hypotheses. **Saudi Journal of Biological Sciences**, [s. l.], n. xxxx, 2021. Disponível em: <https://doi.org/10.1016/j.sjbs.2021.06.046>.
- GRAJEDA-IGLESIAS, Claudia *et al.* Antioxidant activity of protocatechuates evaluated by DPPH, ORAC, and CAT methods. **FOOD CHEMISTRY**, [s. l.], ano 194, 2016. p. 749–757. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2015.07.119>.
- GRANATO, Daniel; KATAYAMA, Flávia Chizuko Uchida; DE CASTRO, Inar Alves. Phenolic composition of South American red wines classified according to their antioxidant activity, retail price and sensory quality. **Food Chemistry**, [s. l.], v. 129, n. 2, p. 366–373, 2011. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2011.04.085>.
- GRAY, Simon M; BLOCH, Michael H. Systematic Review of Proinflammatory Cytokines in Obsessive-Compulsive Disorder. [s. l.], p. 220–228, 2012.
- GUADAMURO, Lucía *et al.* Characterization of four β -glucosidases acting on iso flavone-glycosides from *Bifidobacterium pseudocatenulatum* IPLA 36007. **Food Research International**, [s. l.], v. 100, n. July, p. 522–528, 2017.
- GUO, Honghui; LING, Wenhua. The update of anthocyanins on obesity and type 2 diabetes: Experimental evidence and clinical perspectives. **Reviews in Endocrine and Metabolic Disorders**, [s. l.], v. 16, n. 1, p. 1–13, 2015.
- HAMINIUK, Charles Windson Isidoro *et al.* Extraction and quantification of phenolic acids and flavonols from *Eugenia pyriformis* using different solvents. **Journal of Food Science and Technology**, [s. l.], v. 51, n. 10, p. 2862–2866, 2014.
- HANDA, Cíntia Ladeira *et al.* Parameters of the fermentation of soybean flour by *Monascus purpureus* or *Aspergillus oryzae* on the production of bioactive compounds and antioxidant activity. **Food Chemistry**, [s. l.], v. 271, 2019.
- HARTANTI, Anastasia Tatik; RAHAYU, Gayuh; HIDAYAT, Iman. Rhizopus Species from Fresh Tempeh Collected from Several Regions in Indonesia. **HAYATI Journal of Biosciences**, [s. l.], v. 22, n. 3, p. 136–142, 2015. Disponível em: <http://dx.doi.org/10.1016/j.hjb.2015.10.004>.
- HE, Guiqiang *et al.* Comparing the differences of characteristic flavour between natural maturation and starter culture for Mucor-type Douchi. **International Journal Food Science and Technology**, [s. l.], v. 51, p. 1252–1259, 2016.
- HE, Ronghai *et al.* Modeling the QSAR of ACE-Inhibitory Peptides with ANN and Its Applied Illustration. **International Journal of Peptides**, [s. l.], v. 2012, 2012.
- HESELTIME, C W. Microbiology of Oriental Fermented Foods. **Annual Review of Microbiology**, [s. l.], v. 37, n. 1, p. 575–601, 1983. Disponível em: <http://www.annualreviews.org/doi/10.1146/annurev.mi.37.100183.003043>.
- HIROTA, Akira *et al.* 1,1-Diphenyl-2-picrylhydrazyl Radical-scavenging Compounds from Soybean Miso and Antiproliferative Activity of Isoflavones from Soybean Miso toward the Cancer Cell Lines. **Bioscience, Biotechnology, and Biochemistry**, [s. l.],

v. 64, n. 5, p. 1038–1040, 2005.

HIROTA, Akira *et al.* Isolation of 8-Hydroxyglycitein and 6-Hydroxydaidzein from Soybean Miso. **Bioscience, Biotechnology, and Biochemistry**, [s. l.], v. 68, n. 6, p. 1372–1374, 2004.

HITOSUGI, Masahito; HAMADA, Katsuo; MISAKA, Kazutaka. Effects of *Bacillus subtilis* var. natto products on symptoms caused by blood flow disturbance in female patients with lifestyle diseases. **International Journal of General Medicine**, [s. l.], v. 8, p. 41–46, 2015.

HONG, Go-Eun; MANDAL, Prabhat Kumar; LIM, Ki-Won Chi-Ho Lee. Fermentation Increases Isoflavone Aglycone Contents in Black Soybean Pulp. **Asian Journal of Animal and Veterinary Advances**, [s. l.], v. 7, n. 6, p. 502–511, 2012.

HSIAO, Yu Hsuan; HO, Chi Tang; PAN, Min Hsiung. Bioavailability and health benefits of major isoflavone aglycones and their metabolites. **Journal of Functional Foods**, [s. l.], v. 74, n. August, p. 0–8, 2020.

HUANG, Xiaoning *et al.* Bacterial community succession and metabolite changes during sufu fermentation. **LWT - Food Science and Technology**, [s. l.], v. 97, n. April, p. 537–545, 2018. Disponível em: <https://doi.org/10.1016/j.lwt.2018.07.041>.

HUANG, Chao *et al.* Soy isoflavones regulate lipid metabolism through an AKT/mTORC1 pathway in Diet-Induced Obesity (DIO) male rats. **Molecules**, [s. l.], v. 21, n. 5, p. 1–13, 2016.

HUANG, Q. *et al.* Targeting inflammation and cytokine storm in COVID-19. **Pharmacological Research journal**, [s. l.], v. 159, p. 3–5, 2020.

HUANG, Yung Hsin; LAI, Ying Jang; CHOU, Cheng Chun. Fermentation temperature affects the antioxidant activity of the enzyme-ripened sufu, an oriental traditional fermented product of soybean. **Journal of Bioscience and Bioengineering**, [s. l.], v. 112, n. 1, p. 49–53, 2011. Disponível em: <http://dx.doi.org/10.1016/j.jbiosc.2011.03.008>.

IBE, Sachie *et al.* Natto (Fermented Soybean) Extract Extends the Adult Lifespan of *Caenorhabditis elegans*. **Bioscience, Biotechnology, and Biochemistry**, [s. l.], v. 77, n. 2, p. 392–394, 2013.

INOMATA, Naoko; MIYAKAWA, Mami; AIHARA, Michiko. Surfing as a risk factor for sensitization to poly(γ -glutamic acid) in fermented soybeans, natto, allergy. **Allergology International**, [s. l.], v. 67, n. 3, p. 341–346, 2018. Disponível em: <https://doi.org/10.1016/j.alit.2017.11.001>.

INOUE, Yutaka *et al.* Analysis of the cooked aroma and odorants that contribute to umami aftertaste of soy miso (Japanese soybean paste). **Food Chemistry**, [s. l.], v. 213, p. 521–528, 2016. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2016.06.106>.

ITO, Koji. Review of the health benefits of habitual consumption of miso soup : focus on the effects on sympathetic nerve activity , blood pressure , and heart rate. **Environmental Health and Preventive Medicine**, [s. l.], v. 4, p. 1–9, 2020.

JASKULSKI, Stefanie *et al.* Genistein and enterolactone in relation to Ki-67 expression and HER2 status in postmenopausal breast cancer patients. **Molecular Nutrition and Food Research**, [s. l.], v. 61, n. 11, p. 1–10, 2017.

JEONG, Do Yeon *et al.* Short-Term Fermented Soybeans with *Bacillus amyloliquefaciens* Potentiated Insulin Secretion Capacity and Improved Gut Microbiome Diversity and Intestinal Integrity to Alleviate Asian Type 2 Diabetic Symptoms. **Journal of Agricultural and Food Chemistry**, [s. l.], v. 68, n. 46, p. 13168–13178, 2020.

JEONG, Do-Youn *et al.* γ -PGA-Rich Chungkookjang, Short-Term Fermented

- Soybeans: Prevents Memory Impairment by Modulating Brain Insulin Sensitivity, Neuro-Inflammation, and the Gut-Microbiome-Brain Axis. **Foods**, [s. l.], v. 10, p. 221, 2021. Disponível em: <https://doi.org/10.3390/foods10020221>. Acesso em: 7 jan. 2022.
- JIA, Fu *et al.* Soybean supply chain management and sustainability: A systematic literature review. **Journal of Cleaner Production**, [s. l.], v. 255, p. 120254, 2020. Disponível em: <https://doi.org/10.1016/j.jclepro.2020.120254>.
- JOKIĆ, Stela *et al.* Solubility and kinetics of soybean oil and fatty acids in supercritical CO₂. **European Journal of Lipid Science and Technology**, [s. l.], v. 113, n. 5, p. 644–651, 2011.
- JUCHEN, Patricia Trevisani *et al.* Extraction of parboiled rice bran oil with supercritical CO₂ and ethanol as co-solvent: Kinetics and characterization. **Industrial Crops & Products**, [s. l.], v. 139, n. February, p. 111506, 2019. Disponível em: <https://doi.org/10.1016/j.indcrop.2019.111506>.
- JUNG, Ji Young; LEE, Se Hee; JEON, Che Ok. Microbial community dynamics during fermentation of doenjang-meju, traditional Korean fermented soybean. **International Journal of Food Microbiology**, [s. l.], v. 185, p. 112–120, 2014. Disponível em: <http://dx.doi.org/10.1016/j.ijfoodmicro.2014.06.003>.
- JUNG, Keun Ok; PARK, Su Young; PARK, Kun Young. Longer aging time increases the anticancer and antimetastatic properties of doenjang. **Nutrition**, [s. l.], v. 22, n. 5, p. 539–545, 2006.
- KAMAO, Maya *et al.* Vitamin K Content of Foods and Dietary Vitamin K Intake in Japanese Young Women. **J Nutr Sci Vitaminol**, [s. l.], v. 53, p. 464–470, 2007.
- KANG, Su Jin *et al.* Antioxidant and Neuroprotective Effects of Doenjang Prepared with *Rhizopus*, *Pichia*, and *Bacillus*. **Prev. Nutr. Food Sci.**, [s. l.], v. 21, n. 3, p. 221–226, 2016.
- KATAGIRI, Ryoko *et al.* Association of soy and fermented soy product intake with total and cause specific mortality: prospective cohort study. **Bmj**, [s. l.], p. 368, 2020.
- KAYANO, Shin Ichi *et al.* Isoflavone C-glycosides isolated from the root of kudzu (*Pueraria lobata*) and their estrogenic activities. **Food Chemistry**, [s. l.], v. 134, n. 1, p. 282–287, 2012. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2012.02.137>.
- KHOSRAVI, Azin; RAZAVI, Seyed Hadi. Therapeutic effects of polyphenols in fermented soybean and black soybean products. **Journal of Functional Foods**, [s. l.], v. 81, n. March, p. 104467, 2021. Disponível em: <https://doi.org/10.1016/j.jff.2021.104467>.
- KIM, Il Sup *et al.* Current perspectives on the physiological activities of fermented soybean-derived Cheonggukjang. **International Journal of Molecular Sciences**, [s. l.], v. 22, n. 11, 2021.
- KIM, Min Young *et al.* Free and bound form bioactive compound profiles in germinated black soybean (*Glycine max* L.). **Food Sci. Biotechnol.**, [s. l.], v. 25, n. 6, p. 1551–1559, 2016.
- KIM, Jonggun; PARK, Yooheon; PARK, Yeonhwa. trans -10, cis -12 CLA promotes osteoblastogenesis via SMAD mediated mechanism in bone marrow mesenchymal stem cells. **Journal of Functional Foods**, [s. l.], v. 8, p. 367–376, 2014. Disponível em: <http://dx.doi.org/10.1016/j.jff.2014.04.006>.
- KIMURA, Tomomi. East meets west: Ethnic differences in prostate cancer epidemiology between East Asians and Caucasians. **Chinese Journal of Cancer**, [s. l.], v. 31, n. 9, p. 421–429, 2012.
- KOJIMA, Akane *et al.* Natto Intake is Inversely Associated with Osteoporotic Fracture Risk in Postmenopausal Bone mass. **The Journal of Nutrition**, [s. l.], n. 7, p. 599–605, 2019.

- KRAUS, Daniel *et al.* Nicotinamide N-methyltransferase knockdown protects against diet-induced obesity. **Nature**, [s. l.], v. 508, n. 7495, p. 258–262, 2014.
- KUBA, Keiji *et al.* A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus – induced lung injury. **Nature medicine**, [s. l.], v. 11, n. 8, p. 875–879, 2005.
- KUBA, Megumi *et al.* Angiotensin I-Converting Enzyme Inhibitory Peptides Isolated from Tofuyo Fermented Soybean Food. **Bioscience, Biotechnology, and Biochemistry**, [s. l.], v. 67, n. 6, p. 1278–1283, 2003.
- KULIGOWSKI, Maciej; PAWŁOWSKA, Katarzyna; JASIŃSKA-KULIGOWSKA, Iwona. Isoflavone composition , polyphenols content and antioxidative activity of soybean seeds during tempeh fermentation. **CyTA - Journal of Food**, [s. l.], v. 15, n. 1, p. 27–33, 2017. Disponível em: <http://dx.doi.org/10.1080/19476337.2016.1197316>.
- KUMAR, Amit *et al.* Production of bioactive protein hydrolysate using the yeasts isolated from soft chhurpi. **Bioresource Technology**, [s. l.], v. 219, p. 239–245, 2016. Disponível em: <http://dx.doi.org/10.1016/j.biortech.2016.07.129>.
- KUROSAWA, Yuko *et al.* A single-dose of oral nattokinase potentiates thrombolysis and anti- coagulation profiles. **Nature Publishing Group**, [s. l.], v. 5, p. 11601, 2015.
- LARA, Jose; MICHAEL J., Sherratt; MARGARET, Rees. Aging and anti-aging. **Maturitas**, [s. l.], v. 93, p. 1–3, 2016.
- LEE, Mi Hwa *et al.* Characterization of antimicrobial lipopeptides produced by *Bacillus* sp. LM7 isolated from chungkookjang, a Korean traditional fermented soybean food. **International Journal of Food Microbiology**, [s. l.], v. 221, p. 12–18, 2016. Disponível em: <http://dx.doi.org/10.1016/j.ijfoodmicro.2015.12.010>.
- LEE, Jin Hwan *et al.* Comparisons of nutritional constituents in soybeans during solid state fermentation times and screening for their glucosidase enzymes and antioxidant properties. **Food Chemistry**, [s. l.], v. 272, n. August 2018, p. 362–371, 2019. Disponível em: <https://doi.org/10.1016/j.foodchem.2018.08.052>.
- LEE, H. P. *et al.* Dietary effects on breast-cancer risk in Singapore. **The Lancet**, [s. l.], v. 337, n. 8751, p. 1197–1200, 1991.
- LEE, Jin Hwan; CHO, Kye Man. Changes occurring in compositional components of black soybeans maintained at room temperature for different storage periods. **Food Chemistry**, [s. l.], v. 131, n. 1, p. 161–169, 2012. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2011.08.052>.
- LEI, Hongwei *et al.* Genistein Reverses Free Fatty Acid-induced Insulin Resistance in HepG2 Hepatocytes through Targeting JNK. **Journal of Huazhong University of Science and Technology [Medical Sciences]**, [s. l.], v. 31, n. 2, p. 185–189, 2011.
- LEI, Chen *et al.* Modifications of dietary flavonoids towards improved bioactivity : An update on structure – activity relationship. **Critical Reviews in Food Science and Nutrition**, [s. l.], v. 58, n. 4, p. 513–527, 2018. Disponível em: <https://doi.org/10.1080/10408398.2016.1196334>.
- LEONARDI, Gislaine Ricci. An overview about oxidation in clinical practice of skin aging *. [s. l.], v. 92, n. 3, p. 367–374, 2017.
- LI-JUN, Yin *et al.* Changes in isoflavone contents and composition of sufu (fermented tofu) during manufacturing. **Food Chemistry**, [s. l.], v. 87, n. 4, p. 587–592, 2004.
- LI, Quan *et al.* Dynamic effects of fermentation on phytochemical composition and antioxidant properties of wampee (*Clausena lansium* (Lour .) Skeel) leaves. [s. l.], n. May 2018, p. 76–85, 2019.
- LI, Wei Fang *et al.* Genistein ameliorates Ischemia/Reperfusion-Induced renal injury in a SIRT1-Dependent manner. **Nutrients**, [s. l.], v. 9, n. 4, 2017.
- LI, Xiang *et al.* One-step coextraction method for flavouring soybean oil with the dried

- stipe of *Lentinus edodes* (Berk .) sing by supercritical CO₂ fluid extraction. **LWT - Food Science and Technology**, [s. l.], v. 120, n. October 2019, p. 108853, 2020. Disponível em: <https://doi.org/10.1016/j.lwt.2019.108853>.
- LIAO, Chien-li *et al.* Anti-inflammatory Activity of Soymilk and Fermented Soymilk Prepared with Lactic Acid Bacterium and Bifidobacterium Anti-inflammatory Activity of Soymilk and Fermented Soymilk Prepared with Lactic Acid Bacterium and Bifidobacterium. **Journal of Food and Drug Analysis**, [s. l.], v. 3, n. June, 2010.
- LOANE, David J; BYRNES, Kimberly R. Role of Microglia in Neurotrauma. **Neurotherapeutics**, [s. l.], v. 7, n. 4, p. 366–377, 2010.
- MA, Yanli; CHENG, Yongqiang; YIN, Lijun. Effects of Processing and NaCl on Angiotensin I-Converting Enzyme Inhibitory Activity and γ -Aminobutyric Acid Content During Sufu Manufacturing. **food bioprocess technology**, [s. l.], v. 6, p. 1782–1789, 2013.
- MAGALHÃES, João Pedro De; STEVENS, Michael; THORNTON, Daniel. The Business of Anti-Aging Science. **Trends in Biotechnonology**, [s. l.], v. 35, n. 11, p. 1062–1073, 2017.
- MAGNAN, Christophe; LEVIN, Barry E; LUQUET, Serge. Molecular and Cellular Endocrinology Brain lipid sensing and the neural control of energy balance. **Molecular and Cellular Endocrinology**, [s. l.], v. 418, p. 3–8, 2015. Disponível em: <http://dx.doi.org/10.1016/j.mce.2015.09.019>.
- MAH, Jae-hyung. Fermented Soybean Foods : Significance of Biogenic Amines. [s. l.], v. 3, n. 1, p. 1–3, 2015.
- MAHMUDPOUR, Mehdi *et al.* COVID-19 cytokine storm: The anger of inflammation. **Cytokine**, [s. l.], p. 155151, 2020. Disponível em: <https://doi.org/10.1016/j.cyto.2020.155151>.
- MAI, Chiang; TECHNOLOGY, Agricultural; RAI, Chiang. Analysis and characterisation of amino acid contents of thua nao , a traditionally fermented soybean food of Northern Thailand. **International Food Research Journal**, [s. l.], v. 599, p. 595–599, 2011.
- MAN, Kye *et al.* Changes of phytochemical constituents (isoflavones , flavanols , and phenolic acids) during cheonggukjang soybeans fermentation using potential probiotics *Bacillus subtilis* CS90. **Journal of Food Composition and Analysis**, [s. l.], v. 24, n. 3, p. 402–410, 2011. Disponível em: <http://dx.doi.org/10.1016/j.jfca.2010.12.015>.
- MAN, Kye *et al.* Soluble phenolics and antioxidant properties of soybean (*Glycine max* L .) cultivars with varying seed coat colours. **Journal of Functional Foods**, [s. l.], v. 5, n. 3, p. 1065–1076, 2013. Disponível em: <http://dx.doi.org/10.1016/j.jff.2013.03.002>.
- MANABE, Hisashi. D-Amino Acids in Viscous Parts of Natto. **Journal for the Integrated Study of Dietary Habits**, [s. l.], v. 14, n. 3, p. 200–206, 2011.
- MANI, V; MING, L C. **Tempeh and Other Fermented Soybean Products Rich in Isoflavones**. [S. l.]: Elsevier Inc., 2017. *E-book*. Disponível em: <http://dx.doi.org/10.1016/B978-0-12-802309-9.00019-4>.
- MARAZZA, A *et al.* Enhancement of the antioxidant capacity of soymilk by fermentation with *Lactobacillus rhamnosus*. [s. l.], v. 4, p. 0–7, 2012.
- MARCO, Maria L *et al.* Health benefits of fermented foods : microbiota and beyond Michael Ga. **Current Opinion in Biotechnology**, [s. l.], v. 44, p. 94–102, 2017.
- MEDIC, Jelena; ATKINSON, Christine; HURBURGH, Charles R. Current knowledge in soybean composition. **JAOCs, Journal of the American Oil Chemists' Society**, [s. l.], v. 91, n. 3, p. 363–384, 2014.
- MEINLSCHMIDT, Pia *et al.* Enzymatic treatment of soy protein isolates : effects on the potential allergenicity , technofunctionality , and sensory properties. [s. l.], 2015.

- MELO, Enayde de Almeida *et al.* Capacidade antioxidante de hortaliças usualmente consumidas. **Ciência e Tecnologia de Alimentos**, [s. l.], v. 26, n. 3, p. 639–644, 2006. Disponível em: http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0101-20612006000300024&lng=pt&nrm=iso&tlng=pt.
- MESOMO, Michele C *et al.* Ginger (*Zingiber officinale* R .) extracts obtained using supercritical CO₂ and compressed propane: Kinetics and antioxidant activity evaluation. **The Journal of Supercritical Fluids**, [s. l.], v. 71, p. 102–109, 2012. Disponível em: <http://dx.doi.org/10.1016/j.supflu.2012.08.001>.
- MISIHAIKABGWI, Jane; CHEIKHYOUSSEF, Ahmad. Traditional fermented foods and beverages of Namibia. **Journal of Ethnic Foods**, [s. l.], v. 4, n. 3, p. 145–153, 2017. Disponível em: <https://doi.org/10.1016/j.jef.2017.08.001>.
- MO, Haizhen *et al.* Effect of soybean processing on content and bioaccessibility of folate , vitamin B12 and isoflavones in tofu and tempe. **Food Chemistry**, [s. l.], v. 141, n. 3, p. 2418–2425, 2013. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2013.05.017>.
- MOHSENI, Houra *et al.* Are history of dietary intake and food habits of patients with clinical symptoms of COVID 19 different from healthy controls ? A case e control study. **Clinical Nutrition ESPEN**, [s. l.], v. 42, p. 280–285, 2021. Disponível em: <https://doi.org/10.1016/j.clnesp.2021.01.021>.
- MONTALES, Maria Theresa E. *et al.* Metformin and soybean-derived bioactive molecules attenuate the expansion of stem cell-like epithelial subpopulation and confer apoptotic sensitivity in human colon cancer cells. **Genes and Nutrition**, [s. l.], v. 10, n. 6, p. 1–14, 2015.
- MOORE, Thea R.; FRANKS, Rachel B.; FOX, Carol. Review of Efficacy of Complementary and Alternative Medicine Treatments for Menopausal Symptoms. **Journal of Midwifery and Women’s Health**, [s. l.], v. 62, n. 3, p. 286–297, 2017.
- MORAES, Camila De *et al.* Liver metabolic changes induced by conjugated linoleic acid in calorie-restricted rats. **Arch Endocrinol Metab**, [s. l.], n. 13, 2017.
- MOSMANN, Tim. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. **Journal of Immunological Methods**, [s. l.], v. 65, n. 1–2, p. 55–63, 1983.
- MUCHTARIDI, Muchtaridi *et al.* Natural Flavonoids as Potential for Anti-SARS-CoV-2. **Molecules**, [s. l.], v. 25, n. 3980, 2020.
- MUKHOPADHYAY, Asim Kumar. **Antioxidants-natural and Synthetic**. [S. l.]: Amani Int’l Publishers, 2006.
- MÜLLER, E. *et al.* Toll-Like Receptor Ligands and Interferon- γ Synergize for Induction of Antitumor M1 Macrophages.pdf. **Frontiers in immunology**, [s. l.], v. 8, p. 1383, 2017.
- MURRAY, Peter J; WYNN, Thomas A. Protective and pathogenic functions of macrophage subsets. **Nature Reviews Immunology**, [s. l.], v. 11, n. November, 2011.
- NAGATA, Chisato *et al.* Dietary soy and natto intake and cardiovascular disease mortality in Japanese adults : the Takayama study 1. **American Journal of Clinical Nutrition**, [s. l.], v. 105, p. 426–431, 2017.
- NODA, Shuhei *et al.* Cinnamic acid production using *Streptomyces lividans* expressing phenylalanine ammonia lyase. **Journal of Industrial Microbiology and Biotechnology**, [s. l.], v. 38, n. 5, p. 643–648, 2011.
- NURYANI, Hanifah *et al.* Taste and chemical characteristics of low molecular weight fractions from tofuyo – Japanese fermented soybean curd. **Food Chemistry**, [s. l.], v. 252, n. January, p. 265–270, 2018. Disponível em: <https://doi.org/10.1016/j.foodchem.2018.01.117>.

- OBA, Mami *et al.* Natto extract, a Japanese fermented soybean food, directly inhibits viral infections including SARS-CoV-2 in vitro. **Biochemical and Biophysical Research Communications**, [s. l.], v. 570, p. 21–25, 2021. Disponível em: <https://doi.org/10.1016/j.bbrc.2021.07.034>.
- OGASAWARA, Masashi; YAMADA, Yuki; EGI, Makoto. Taste enhancer from the long-term ripening of miso (soybean paste). **Food Chemistry**, [s. l.], v. 99, n. 4, p. 736–741, 2006.
- OLMOS, Antonieta Rodríguez De; GARRO, Marisa Selva. Metabolic profile of *Lactobacillus paracasei* subsp. *paracasei* CRL 207 in solid state fermentation using commercial soybean meal. **Food Bioscience**, [s. l.], v. 35, n. November 2018, p. 100584, 2020. Disponível em: <https://doi.org/10.1016/j.fbio.2020.100584>.
- OLSON, Jake M *et al.* A Comparison of the Anti - Inflammatory Effects of Cis - 9 , Trans - 11 Conjugated Linoleic Acid to Celecoxib in the Collagen - Induced Arthritis Model. **Lipids**, [s. l.], v. 52, n. 2, p. 151–159, 2017.
- OMIZU, Yohei *et al.* Determination of saponin contents in raw soybean and fermented soybean foods of India. **Journal of Scientific and Industrial Research**, [s. l.], v. 70, n. 7, p. 533–538, 2011.
- OPAS, Organização Pan-América da Saúde. **Histórico da pandemia de COVID-19**. [S. l.], 2021. Disponível em: <https://www.paho.org/pt/covid19/historico-da-pandemia-covid-19>. Acesso em: 2 ago. 2021.
- OVERK, Cassia R *et al.* Cortical M 1 receptor concentration increases without a concomitant change in function in Alzheimer ' s disease. **Journal of Chemical Neuroanatomy**, [s. l.], v. 40, n. 1, p. 63–70, 2010. Disponível em: <http://dx.doi.org/10.1016/j.jchemneu.2010.03.005>.
- PAGNONCELLI, M.G.B. *et al.* Nattokinase. *In*: CURRENT DEVELOPMENTS IN BIOTECHNOLOGY AND BIOENGINEERING - PRODUCTION, ISOLATION AND PURIFICATION OF INDUSTRIAL PRODUCTS. [S. l.: s. n.], 2017. p. 509–526.
- PAN, Min-hsiung *et al.* Molecular mechanisms for anti-aging by natural dietary compounds. **Molecular Nutrition and Food Research**, [s. l.], v. 56, p. 88–115, 2012.
- PARK, Kun Young *et al.* Antimutagenic effects of doenjang (Korean fermented soypaste) and its active compounds. **Mutation Research - Fundamental and Molecular Mechanisms of Mutagenesis**, [s. l.], v. 523–524, p. 43–53, 2003.
- PARK, Sunmin *et al.* Ischemic hippocampal cell death induces glucose dysregulation by attenuating glucose-stimulated insulin secretion which is exacerbated by a high fat diet. **Life Sciences**, [s. l.], v. 88, n. 17–18, p. 766–773, 2011. Disponível em: <http://dx.doi.org/10.1016/j.lfs.2011.02.021>.
- PARK, Young Kyoung; LEE, Jae Hoan; MAH, Jae-hyung. Occurrence and reduction of biogenic amines in traditional Asian fermented soybean foods : A review. **Food Chemistry**, [s. l.], v. 278, n. August 2018, p. 1–9, 2019. Disponível em: <https://doi.org/10.1016/j.foodchem.2018.11.045>.
- PARK, Young Kyoung; LEE, Jae Hoan; MAH, Jae Hyung. Occurrence and reduction of biogenic amines in traditional Asian fermented soybean foods: A review. **Food Chemistry**, [s. l.], v. 278, n. October 2018, p. 1–9, 2019. Disponível em: <https://doi.org/10.1016/j.foodchem.2018.11.045>.
- PENDYALAA, Brahmaiah; PATRASA, Ankit. In silico Screening of Food Bioactive Compounds to Predict Potential Inhibitors of COVID-19 Main protease (M. **ChemRxiv**, [s. l.], p. 11–44, 2020.
- PERKINS, Edward G. **Composition of Soybeans and Soybean Products**. [S. l.]: AOCS Press, 1995. v. I *E-book*. Disponível em: <http://dx.doi.org/10.1016/B978-0-935315-63-9.50006-1>.

- PESSOA, Fernando L. P. *et al.* Integrated Soybean Biorefinery. **Biorefinery Concepts**, [s. l.], 2019. Disponível em: <http://dx.doi.org/10.1039/C7RA00172J> <https://www.intechopen.com/books/advanced-biometric-technologies/liveness-detection-in-biometrics> <http://dx.doi.org/10.1016/j.colsurfa.2011.12.014>.
- PICARD, Martin *et al.* Mitochondrial morphology transitions and functions : implications for retrograde signaling ? . **American Journal of Physiology-Regulatory, Integrative and Comparative Physiology**, [s. l.], v. 304, n. 6, p. 396–406, 2013.
- PIGHINELLI, Anna L M T *et al.* Otimização da prensagem de grãos de girassol e sua caracterização Optimization of sunflower grain pressing and its characterization. [s. l.], n. 19, p. 63–67, 2009.
- PRADO, Fernanda Guilherme do Prado *et al.* Fungal-Mediated Biotransformation of Soybean Supplemented with Different Cereal Grains into a Functional Compound with Antioxidant , Anti- Inflammatory and Antitumoral Activities. **Biointerface Research in Applied Chemistry**, [s. l.], v. 11, n. 1, p. 8018–8033, 2021.
- PRIOR, Ronald L. *et al.* Antioxidant Capacity As Influenced by Total Phenolic and Anthocyanin Content, Maturity, and Variety of *Vaccinium* Species. **Journal of Agricultural and Food Chemistry**, [s. l.], v. 46, n. 7, p. 2686–2693, 1998. Disponível em: <https://pubs.acs.org/doi/10.1021/jf980145d>.
- RAI, Amit Kumar; KUMARASWAMY, Jeyaram. Health Benefits of Functional Proteins in Fermented Foods. **Health benefits of fermented foods and beverages**, [s. l.], v. 4, n. April, p. 455–474, 2015.
- RAI, Amit Kumar; SANJUKTA, Samurailatpam; JEYARAM, Kumaraswamy. Production of angiotensin I converting enzyme inhibitory (ACE-I) peptides during milk fermentation and their role in reducing hypertension. **Critical Reviews in Food Science and Nutrition**, [s. l.], v. 57, p. 2789–2800, 2017. Disponível em: <https://doi.org/10.1080/10408398.2015.1068736>.
- RAIMONDI, Stefano *et al.* Bioconversion of soy isoflavones daidzin and daidzein by Bifidobacterium strains. **Applied Microbial and Cell Physiology**, [s. l.], v. 81, p. 943–950, 2009.
- RAINS, Justin L; JAIN, Sushil K. Oxidative stress , insulin signaling , and diabetes. **Free Radical Biology and Medicine**, [s. l.], v. 50, n. 5, p. 567–575, 2011.
- RAMPAZZO, Valéria. **Avaliação da degradação térmica de óleos vegetais por termogravimetria, cromatografia gasosa e espectroscopia de infravermelho médio**. 2015. 93 f. - Universidade Federal do Paraná, [s. l.], 2015.
- RATNANINGRUM, Diah *et al.* The Production of Corn Kernel Miso Based on Rice-koji Fermented by *Aspergillus oryzae* and *Rhizopus oligosporus*. **Journal of Tropical Biodiversity and Biotechnology**, [s. l.], v. 3, n. 1, p. 8, 2018.
- REDDY, Krishna N.; DUKE, Stephen O. **Soybean Mineral Composition and Glyphosate Use**. [S. l.]: Elsevier Inc., 2015. *E-book*. Disponível em: <http://dx.doi.org/10.1016/B978-0-12-404699-3.00044-5>.
- REN, N. N. *et al.* A clinical study on the effect of nattokinase on carotid artery atherosclerosis and hyperlipidaemia. **Zhonghua yi xue za zhi**, [s. l.], v. 97, n. 26, p. 2038–2042, 2017.
- REVIEWS, I N Depth. **Biology of Healthy Aging and Longevity**. [s. l.], p. 7–16, 2016.
- ROMULO, Andreas; SURYA, Reggie. Tempe: A traditional fermented food of Indonesia and its health benefits. **International Journal of Gastronomy and Food Science**, [s. l.], v. 26, n. May, p. 100413, 2021. Disponível em: <https://doi.org/10.1016/j.ijgfs.2021.100413>.
- ROSAS-VILLEGAS, Adriana *et al.* Differential effect of sucrose and fructose in

- combination with a high fat diet on intestinal microbiota and kidney oxidative stress. **Nutrients**, [s. l.], v. 9, n. 4, 2017.
- SACHIE, I *et al.* Antihypertensive Effects of Natto , a Traditional Japanese Fermented Food , in Spontaneously Hypertensive Rats. **Food Science and Technology**, [s. l.], v. 15, n. 2, p. 199–202, 2009.
- SAGARA, Tatsuya *et al.* Soybean spermidine concentration: Genetic and environmental variation of a potential ‘ anti-aging ’ constituent. **Journal of Food Composition and Analysis**, [s. l.], v. 56, p. 11–17, 2017.
- SÁNCHEZ-CHINO, Xariss *et al.* Nutrient and nonnutrient components of legumes, and its chemopreventive activity: A review. **Nutrition and Cancer**, [s. l.], v. 67, n. 3, p. 401–410, 2015.
- SANJA, Đ *et al.* The impact of high-power ultrasound and microwave on the phenolic acid profile and antioxidant activity of the extract from yellow soybean seeds. **Industrial Crops & Products**, [s. l.], v. 122, n. May, p. 223–231, 2018.
- SANJUKTA, Samurailatpam *et al.* Enhancement of antioxidant properties of two soybean varieties of Sikkim Himalayan region by proteolytic *Bacillus subtilis* fermentation. **Journal of Functional Foods**, [s. l.], v. 14, p. 650–658, 2015. Disponível em: <http://dx.doi.org/10.1016/j.jff.2015.02.033>.
- SANJUKTA, Samurailatpam; RAI, Amit Kumar. Production of bioactive peptides during soybean fermentation and their potential health benefits. **Trends in Food Science & Technology**, [s. l.], v. 50, p. 1–10, 2016. Disponível em: <http://dx.doi.org/10.1016/j.tifs.2016.01.010>.
- SANJUKTA, Samurailatpam; RAI, Amit Kumar. Production of bioactive peptides during soybean fermentation and their potential health benefits. **Trends in Food Science and Technology**, [s. l.], v. 50, p. 1–10, 2016. Disponível em: <http://dx.doi.org/10.1016/j.tifs.2016.01.010>.
- SANTACROCE, Luigi *et al.* Potential beneficial role of probiotics on the outcome of COVID-19 patients: An evolving perspective. **Diabetes & Metabolic Syndrome: Clinical Research & Reviews**, [s. l.], v. 15, n. 1, p. 295–301, 2021. Disponível em: <https://doi.org/10.1016/j.dsx.2020.12.040>.
- SANTOS, Reginaldo Ferreira *et al.* Optimization of oil extraction from high energetic potential plants performed through drying and solvent extraction methods. **African Journal of Biotechnology**, [s. l.], v. 12, n. 48, p. 6761–6765, 2013.
- SANTOS, Vidiany A. Q. *et al.* Solid-state fermentation of soybean okara : Isoflavones biotransformation, antioxidant activity and enhancement of nutritional quality. **LWT**, [s. l.], v. 92, n. March, p. 509–515, 2018. Disponível em: <https://doi.org/10.1016/j.lwt.2018.02.067>.
- SANTOS, Ênio R M *et al.* Supercritical fluid extraction of *Rumex Acetosa* L . roots : Yield , composition , kinetics , bioactive evaluation and comparison with conventional techniques. **The Journal of Supercritical Fluids**, [s. l.], v. 122, p. 1–9, 2017. Disponível em: <http://dx.doi.org/10.1016/j.supflu.2016.11.019>.
- SAPBAMRER, Ratana; VISAVARUNGROJ, Nuwat; SUTTAJIT, Maitree. Effects of dietary traditional fermented soybean on reproductive hormones, lipids, and glucose among postmenopausal women in northern thailand. **Asia Pacific Journal of Clinical Nutrition**, [s. l.], v. 22, n. 2, p. 222–228, 2013.
- SATO, Kousuke *et al.* Isolation and characterization of peptides with dipeptidyl peptidase IV (DPPIV) inhibitory activity from natto using DPPIV from *Aspergillus oryzae*. **Food Chemistry**, [s. l.], v. 261, n. October 2017, p. 51–56, 2018. Disponível em: <https://doi.org/10.1016/j.foodchem.2018.04.029>.
- SEPTEMBRE-MALATERRE, Axelle; REMIZE, Fabienne; POUCHERET, Patrick.

- Fruits and vegetables , as a source of nutritional compounds and phytochemicals : Changes in bioactive compounds during lactic fermentation. **Food Research International**, [s. l.], v. 104, n. September 2017, p. 86–99, 2018. Disponível em: <https://doi.org/10.1016/j.foodres.2017.09.031>.
- SGARIGLIA, Melina A. *et al.* Anti-inflammatory properties of phenolic lactones isolated from *Caesalpinia paraguariensis* stem bark. **Journal of Ethnopharmacology**, [s. l.], v. 147, n. 1, p. 63–73, 2013.
- SHARMA, Amit; BALUJA, Zenis; PROFESSOR, Assistant. Therapeutic Effects of Glycine Max (Soybean): A Summary. **International Journal of Research in Pharmacy and Biosciences**, [s. l.], v. 2, n. 1, p. 22–27, 2015.
- SHIN, Donghwa; JEONG, Doyoun. Korean traditional fermented soybean products : Jang. **Journal of Ethnic Foods**, [s. l.], v. 2, n. 1, p. 2–7, 2015. Disponível em: <http://dx.doi.org/10.1016/j.jef.2015.02.002>.
- SHIRLEY STEFFANY MUÑOZ, Fernández; SANDRA MARIA LIMA, Ribeiro. Nutrition and Alzheimer Disease. **Clinics in Geriatric Medicine**, [s. l.], v. 34, n. 4, p. 677–697, 2018. Disponível em: <https://doi.org/10.1016/j.cger.2018.06.012>.
- SHUKLA, Shruti *et al.* Total phenolic content, antioxidant, tyrosinase and α -glucosidase inhibitory activities of water soluble extracts of noble starter culture Doenjang, a Korean fermented soybean sauce variety. **Food Control**, [s. l.], v. 59, p. 854–861, 2016. Disponível em: <http://dx.doi.org/10.1016/j.foodcont.2015.07.003>.
- SILVA, Erissandro dos Santos. **Avaliação de antioxidantes naturais na estabilidade de biodiesel de soja**. 2017. 72 f. - Inuversidade Federal de Campina Grande, [s. l.], 2017.
- SIMATENDE, Protus; HENRY, Tendekayi; JABULANI, Stanley. Methods of preparation of Swazi traditional fermented foods. **Journal of Ethnic Foods**, [s. l.], v. 2, n. 3, p. 119–125, 2015. Disponível em: <http://dx.doi.org/10.1016/j.jef.2015.08.008>.
- SINGH, Pal Brij; YADAV, Deepika; VIJ, Shilpa. Soybean Bioactive Molecules: Current Trend and Future Prospective. **Bioactive Molecules in Food**, [s. l.], p. 267–294, 2019.
- SIVAMARUTHI, Bhagavathi Sundaram; KESIKA, Periyana. Thai Fermented Foods as a Versatile Source of Bioactive Microorganisms — A Comprehensive Review. **Scientia Pharmaceutica**, [s. l.], v. 36, n. 3, p. 37, 2018.
- SODA, Kuniyasu; KANO, Yoshihiko; CHIBA, Fumihiko. Food Polyamine and Cardiovascular Disease -An Epidemiological Study-.pdf. **Global Journal of Health Science**, [s. l.], v. 4, n. 6, p. 170–178, 2012.
- SOLTANI, Saeede; MAZLOUMI, Seyed Hossein. Chemical Engineering Research and Design A new empirical model to correlate solute solubility in supercritical carbon dioxide in presence of co-solvent. **Chemical Engineering Research and Design**, [s. l.], v. 125, p. 79–87, 2017. Disponível em: <http://dx.doi.org/10.1016/j.cherd.2017.07.006>.
- SQUADRITO, Francesco *et al.* Genistein in the metabolic syndrome: Results of a randomized clinical trial. **Journal of Clinical Endocrinology and Metabolism**, [s. l.], v. 98, n. 8, p. 3366–3374, 2013.
- SQUEO, G. *et al.* Evaluation of total phenolic content in virgin olive oil using fluorescence excitation–emission spectroscopy coupled with chemometrics. **Journal of the Science of Food and Agriculture**, [s. l.], v. 99, p. 2513–2520, 2019.
- STEINKRAUS, K. H. *et al.* Studies on tempeh. An Indonesian fermented soybean food. **Food Research**, [s. l.], v. 25, n. 1176, 1959.
- STROMINGER, L; LELOIR, F; REISSIG, L. A modified colorimetric method for the estimatuon of N-acetylamino sugars. **Journal of Biological Chemistry**, [s. l.], v. 2, p. 959-966., 1955.

- SUNDARAM, Madhumitha Kedhari *et al.* Genistein Induces Alterations of Epigenetic Modulatory Signatures in Human Cervical Cancer Cells. **Anti-Cancer Agents in Medicinal Chemistry**, [s. l.], v. 18, p. 412–421, 2018.
- SURYA, Reggie; ROMULO, Andreas. Tempeh Extract Protects HepG2 Cells against Oxidative Stress-Induced Cell Death. **Journal of Physics: Conference Series**, [s. l.], v. 1655, n. 1, 2020.
- SWAIN, T; HILLS, W E. Determina-c* ao de fen! olicos totais. **J. Sci. Food Agric**, [s. l.], v. 10, p. 63–68, 1959.
- SZLISZKA, Ewelina *et al.* Inhibition of Inflammatory Mediators by Neobavaisoflavone in Activated RAW264.7 Macrophages. **Molecules**, [s. l.], v. 16, p. 3701–3712, 2011.
- TAKABAYASHI, Tetsuji *et al.* Nattokinase, profibrinolytic enzyme, effectively shrinks the nasal polyp tissue and decreases viscosity of mucus. **Allergology International**, [s. l.], v. 66, n. 4, p. 594–602, 2017. Disponível em: <https://doi.org/10.1016/j.alit.2017.03.007>.
- TAMANG, Jyoti P; WATANABE, Koichi; HOLZAPFEL, Wilhelm H. Review : Diversity of Microorganisms in Global Fermented Foods and Beverages. **Frontiers in Microbiology**, [s. l.], v. 7, n. March, 2016.
- TAN, Yue *et al.* Effect of different starter cultures on the control of biogenic amines and quality change of douchi by rapid fermentation. **LWT - Food Science and Technology**, [s. l.], v. 109, n. April, p. 395–405, 2019. Disponível em: <https://doi.org/10.1016/j.lwt.2019.04.041>.
- TANGO, João Shojiro; CARVALHO, Cássia Regina Limonta; SOARES, Nilberto Bernado. Physical and chemical characterization of avocado fruits aiming its potencial for oil extraction. **Rev. Bras. Frutic.**, [s. l.], v. 26, n. 1, p. 17–23, 2004.
- TIAN, Ying *et al.* Comparative analysis of lipid profile and in vitro cytotoxic activity of fermented and unfermented soybean extracted by supercritical CO₂ extraction. **Journal of Functional Foods**, [s. l.], v. 34, p. 369–378, 2017. Disponível em: <http://dx.doi.org/10.1016/j.jff.2017.05.006>.
- TIM, E *et al.* Organic compounds trapped in aqueous fluid inclusions. **Organic Geochemistry**, [s. l.], v. 29, n. 1, p. 195–205, 1998.
- TSAI, Cheng Yu *et al.* Effect of soy saponin on the growth of human colon cancer cells. **World Journal of Gastroenterology**, [s. l.], v. 16, n. 27, p. 3371–3376, 2010.
- TYUG, Tan Seok; PRASAD, K. Nagendra; ISMAIL, Amin. Antioxidant capacity, phenolics and isoflavones in soybean by-products. **Food Chemistry**, [s. l.], v. 123, n. 3, p. 583–589, 2010. Disponível em: <http://dx.doi.org/10.1016/j.foodchem.2010.04.074>.
- UBA, Megumi K *et al.* Angiotensin I-Converting Enzyme Inhibitory Peptides Isolated from Tofuyo Fermented Soybean Food. **Bioscience, Biotechnology, and Biochemistry**, [s. l.], v. 67, n. 6, p. 1278–1283, 2003.
- USDA. **United States Department of Agriculture**. [S. l.], 2022a. Disponível em: <https://usdabrazil.org.br/>. Acesso em: 2 fev. 2022.
- USDA. **United States Department of Agriculture**. [S. l.], 2022b. Disponível em: <https://www.fas.usda.gov/data/world-agricultural-production>.
- VEBER, J *et al.* Determinação dos compostos fenólicos e da capacidade antioxidante de extratos aquosos e etanólicos de Jambolão (*Syzygium cumini* L.). **Rev. bras. plantas med**, [s. l.], v. 17, n. 2, p. 267–273, 2015.
- VERDRENGH, M *et al.* Genistein as an anti-inflammatory agent. **Inflammation Research**, [s. l.], v. 52, p. 341–346, 2003.
- VIEIRA, Carla Paulo *et al.* *Lactococcus lactis* ssp. *cremoris* MRS47, a potential probiotic strain isolated from kefir grains, increases cis-9, trans-11-CLA and PUFA

- contents in fermented milk. **Journal of Functional Foods**, [s. l.], v. 31, p. 172–178, 2017.
- WANG, Daijie *et al.* A novel method for highly efficient biotransformation and separation of isoflavone aglycones from soybean with high-speed counter-current chromatography. **Industrial Crops & Products**, [s. l.], v. 129, n. November 2018, p. 224–230, 2019. Disponível em: <https://doi.org/10.1016/j.indcrop.2018.11.043>.
- WANG, Siyu *et al.* Anti-obesity molecular mechanism of soy isoflavones: Weaving the way to new therapeutic routes. **Food and Function**, [s. l.], v. 8, n. 11, p. 3831–3846, 2017.
- WANG, Yu-kun *et al.* Antioxidant property and their free, soluble conjugate and insoluble-bound phenolic. **Journal of Functional Foods**, [s. l.], v. 24, p. 359–372, 2016. Disponível em: <http://dx.doi.org/10.1016/j.jff.2016.04.026>.
- WANG, Shuo *et al.* Effect of the Chemical Composition of Miso (Japanese Fermented Soybean Paste) Upon the Sensory Evaluation. **Analytical Letters**, [s. l.], v. 52, n. 11, p. 1813–1827, 2019. Disponível em: <https://doi.org/10.1080/00032719.2019.1570244>.
- WANG, Dong *et al.* In vitro and in vivo studies on the antioxidant activities of the aqueous extracts of Douchi (a traditional Chinese salt-fermented soybean food). **Food Chemistry**, [s. l.], v. 107, n. 4, p. 1421–1428, 2008.
- WATANABE, Hiromitsu *et al.* Protective Effects of Japanese Soybean Paste (Miso) on Stroke in Stroke-Prone Spontaneously Hypertensive Rats (SHRSP). **American Journal of Hypertension**, [s. l.], v. 31, n. January, p. 43–47, 2018.
- WHO. **Coronavirus (COVID-19) Dashboard** World Health Organization. [S. l.], 2021. Disponível em: <https://covid19.who.int/>. Acesso em: 2 ago. 2021.
- WIPO. **World Intellectual Property Organization**. [S. l.], 2022.
- WOOD, B J B. **Fermentation , Origins and Applications**. 2. ed. [S. l.]: Elsevier Ltd., 2016. *E-book*. Disponível em: <http://dx.doi.org/10.1016/B978-0-08-100596-5.00135-9>.
- WU, Fei *et al.* Assessing sustainability of soybean supply in China: Evidence from provincial production and trade data. **Journal of Cleaner Production**, [s. l.], v. 244, p. 119006, 2020. Disponível em: <https://doi.org/10.1016/j.jclepro.2019.119006>.
- WU, Authors *et al.* The Soybean Aphid in China : A Historical Review. **Entomological Society of America**, [s. l.], v. 97, n. 2, p. 209–218, 2004.
- XIAO, Yu *et al.* Enhancement of the antioxidant capacity of soy whey by fermentation with *Lactobacillus plantarum* B1-6. **Journal of Functional Foods**, [s. l.], v. 12, p. 33–44, 2015.
- XU, L.; CAI, W. X.; XU, B. J. A systematic assesmet on vitamins (B2, B12) an GABA profiles in fermented soy products marketed in china. **Journal of Food Processing and Preservation**, [s. l.], v. 1, p. 1–12, 2017.
- XU, Lu; DU, Bin; XU, Baojun. A systematic, comparative study on the beneficial health components and antioxidant activities of commercially fermented soy products marketed in China. [s. l.], v. 174, p. 202–213, 2015.
- YANG, Huilin *et al.* Exploring functional core bacteria in fermentation of a traditional Chinese food, *Aspergillus*-type douchi. **PLoS ONE**, [s. l.], v. 14, n. 12, p. 1–14, 2019. Disponível em: <http://dx.doi.org/10.1371/journal.pone.0226965>.
- YANG, Bing; TAN, Yue; KAN, Jianquan. Determination and mitigation of chemical risks in sufu by NaCl and ethanol addition during fermentation. **Journal of Food Composition and Analysis**, [s. l.], v. 98, n. 2, 2021.
- YASUDA, Masaaki. Fermented Tofu, Tofuyo. *In*: SOYBEAN - BIOCHEMISTRY, CHEMISTRY AND PHYSIOLOGY. [S. l.: s. n.], 2011.
- YASUDA, Masaaki; TACHIBANA, Shinjiro; KUBA-MIYARA, Megumi. Biochemical aspects of red koji and tofuyo prepared using *Monascus* fungi. [s. l.], p. 49–60, 2012.

- YATAGAI, Chieko *et al.* Nattokinase-promoted tissue plasminogen activator release from human cells. **Pathophysiology of Haemostasis and Thrombosis**, [s. l.], v. 36, n. 5, p. 227–232, 2008.
- YEON, Do *et al.* Chungkookjang , a soy food , fermented with *Bacillus amyloliquefaciens* protects gerbils against ishcmcic stroke injury , and post-stroke hyperglycemia. **Food Research International**, [s. l.], v. 128, n. August 2019, p. 108769, 2020. Disponível em: <https://doi.org/10.1016/j.foodres.2019.108769>.
- YOUNG, Dae *et al.* Antidiabetic effects of fermented soybean products on type 2 diabetes. **Nutrition Research**, [s. l.], v. 30, n. 1, p. 1–13, 2010. Disponível em: <http://dx.doi.org/10.1016/j.nutres.2009.11.004>.
- YOUNG, Dae *et al.* Isoflavonoids and peptides from meju , long-term fermented soybeans , increase insulin sensitivity and exert insulinotropic effects in vitro. **Nutrition**, [s. l.], v. 27, n. 2, p. 244–252, 2011. Disponível em: <http://dx.doi.org/10.1016/j.nut.2010.02.004>.
- YUCE, Hatice Balci *et al.* The effect of commercial conjugated linoleic acid products on experimental periodontitis and diabetes mellitus in Wistar rats The effect of commercial conjugated linoleic acid products on experimental periodontitis and diabetes mellitus in Wistar rats Hat. **Acta Odontologica Scandinavica**, [s. l.], n. November, 2016.
- YUSOF, Hamidah Mohd *et al.* Anti-inflammatory , analgesic and acute toxicity effects of fermented soybean. [s. l.], v. 2, p. 1–7, 2019.
- ZENG, Furong *et al.* Association of inflammatory markers with the severity of COVID-19 : A meta-analysis. **International Journal of Infectious Diseases**, [s. l.], v. 96, p. 467–474, 2020. Disponível em: <https://doi.org/10.1016/j.ijid.2020.05.055>.
- ZHANG, Jian-hua *et al.* Angiotensin I-converting enzyme inhibitory peptides in douchi , a Chinese traditional fermented soybean product. **Food Chemistry**, [s. l.], v. 98, p. 551–557, 2006.
- ZHANG, Z. *et al.* Production of powerful antioxidant supplements via solid-state fermentation of wheat (*Triticum aestivum* Linn.) by *Cordyceps militaris*. **Food Technology and Biotechnology**, [s. l.], v. 50, n. 1, p. 32–39, 2012.
- ZHANG, Binle; YANG, Zixuan; HUANG, Weining. Isoflavone aglycones enrichment in soybean sourdough bread fermented by lactic acid bacteria strains isolated from traditional Qu starters : Effects on in vitro gastrointestinal digestion , nutritional , and baking properties. **Cereal Chemistry**, [s. l.], v. 96, n. May 2018, p. 129–141, 2019.
- ZHENG, L *et al.* Effects of *Bacillus* fermentation on the protein microstructure and anti-nutritional factors of soybean meal. [s. l.], 2017.
- ZHOU, Rui *et al.* Effective control of microbial spoilage in soybeans by water-soluble ZnO nanoparticles. **Food Chemistry**, [s. l.], v. 388, n. August 2021, p. 132994, 2022. Disponível em: <https://doi.org/10.1016/j.foodchem.2022.132994>.
- ZHU, Yun-yang *et al.* B-vitamin enriched fermented soymilk : A novel strategy for soy-based functional foods development. **Trends in Food Science & Technology**, [s. l.], v. 105, n. September, p. 43–55, 2020. Disponível em: <https://doi.org/10.1016/j.tifs.2020.08.019>.
- ZHU, Beibei *et al.* Dietary legume consumption reduces risk of colorectal cancer: Evidence from a meta-analysis of cohort studies. **Scientific Reports**, [s. l.], v. 5, p. 27–33, 2015.

ATTACHMENT 1 – PUBLICATION RESULTS ARTICLEDOI: [10.33263/BRIAC111.80188033](https://doi.org/10.33263/BRIAC111.80188033)

Biointerface Research in Applied Chemistry

Platinum Open Access Journal (ISSN: 2069-5837)

*Article***Volume 11, Issue 1, 2021, 8018 - 8033**<https://doi.org/10.33263/BRIAC111.80188033>**Fungal-Mediated Biotransformation of Soybean
Supplemented with Different Cereal Grains into a
Functional Compound with Antioxidant, Anti-
Inflammatory and Antitumoral Activities**

Fernanda Guilherme do Prado ¹, Mitiyo Fukuda Miyaoka ¹, Gilberto Vinicius de Melo Pereira ¹,
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ATTACHMENT 2 – PUBLICATION AS CO-AUTHORSHIP

<https://doi.org/10.1016/bs.afnr.2019.10.002>





Advances in Food and Nutrition Research



Volume 91, 2020, Pages 65-96



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