

UNIVERSIDADE FEDERAL DO PARANÁ
PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA E CONSERVAÇÃO

LARISSA FARIA



**AVALIAÇÃO DO POTENCIAL IMPACTO ECOLÓGICO CAUSADO PELA
INTRODUÇÃO DO BAGRE NÃO NATIVO *Ictalurus punctatus* (RAFINESQUE,
1818) EM UM RIO NEOTROPICAL**

CURITIBA

2018

LARISSA FARIA

**AVALIAÇÃO DO POTENCIAL IMPACTO ECOLÓGICO CAUSADO PELA
INTRODUÇÃO DO BAGRE NÃO NATIVO *Ictalurus punctatus* (RAFINESQUE,
1818) EM UM RIO NEOTROPICAL**

Dissertação apresentada como requisito parcial à obtenção do grau de Mestre em Ecologia e Conservação, no Curso de Pós-Graduação em Ecologia e Conservação, Setor de Ciências Biológicas, da Universidade Federal do Paraná.

Orientador: Prof. Dr. Jean Ricardo Simões Vitule

CURITIBA
2018

Universidade Federal do Paraná. Sistema de Bibliotecas.
Biblioteca de Ciências Biológicas.
(Telma Terezinha Stresser de Assis –CRB/9-944)

Faria, Larissa

Avaliação do potencial impacto ecológico causado pela introdução do bagre não nativo *Ictalurus punctatus* (Rafinesque, 1818) em um rio Neotropical. / Larissa Faria. – Curitiba, 2018.

55 f.: il. ; 30cm.

Orientador: Jean Ricardo Simões Vitule

Dissertação (Mestrado) - Universidade Federal do Paraná, Setor de Ciências Biológicas. Programa de Pós-Graduação em Ecologia e Conservação.

1. Água doce. 2. Conservação. 3. Bagre (Peixe). I. Título II. Vitule, Jean Ricardo Simões. III. Universidade Federal do Paraná. Setor de Ciências Biológicas. Programa de Pós-Graduação em Ecologia e Conservação.

CDD (20. ed.) 597.52



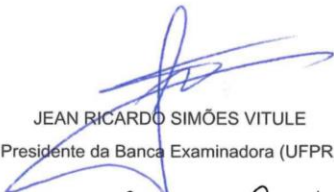
MINISTÉRIO DA EDUCAÇÃO
SETOR CIÊNCIAS BIOLÓGICAS
UNIVERSIDADE FEDERAL DO PARANÁ
PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO ECOLOGIA E
CONSERVAÇÃO

TERMO DE APROVAÇÃO


Os membros da Banca Examinadora designada pelo Colegiado do Programa de Pós-Graduação em ECOLOGIA E CONSERVAÇÃO da Universidade Federal do Paraná foram convocados para realizar a arguição da dissertação de Mestrado de **LARISSA FARIA** intitulada: **Avaliação do potencial impacto ecológico causado pela introdução do bagre não nativo *Ictalurus punctatus* (Rafinesque, 1818) em um rio Neotropical.**, após terem inquirido a aluna e realizado a avaliação do trabalho, são de parecer pela sua APROVAÇÃO no rito de defesa.

A outorga do título de mestre está sujeita à homologação pelo colegiado, ao atendimento de todas as indicações e correções solicitadas pela banca e ao pleno atendimento das demandas regimentais do Programa de Pós-Graduação.

Curitiba, 08 de Março de 2018.



JEAN RICARDO SIMÕES VITULE
Presidente da Banca Examinadora (UFPR)



MHAIRI E. ALEXANDER
Avaliador Externo (UWS)



ANDRE ANDRIAN PADIAL
Avaliador Interno (UFPR)

A todas as mulheres corajosas deste mundo.
Especialmente à minha avó Mari, *in memoriam*.

AGRADECIMENTOS

À Universidade Federal do Paraná, por me acolher por mais dois anos como sua aluna, e por ser a minha segunda casa.

Ao Programa de Pós-Graduação em Ecologia e Conservação, na figura de seus professores e funcionários sempre dispostos a ensinar e auxiliar.

À CAPES, pela bolsa de estudos sem a qual a realização desta empreitada pelo título de mestra não seria possível.

Ao meu orientador Jean Ricardo Simões Vitule, por toda sua dedicação e confiança no meu trabalho. Obrigada por acreditar em mim mais do que eu mesma e me inspirar a buscar o meu melhor sempre.

À banca, professor Dr. André Padial e professora Dra. Mhairi Alexander, por aceitarem o convite prontamente e disponibilizarem um pouco do seu tempo para contribuir com este trabalho.

Aos meus colegas do Laboratório de Ecologia e Conservação, que sempre estiveram a postos quando eu precisei e que tornaram o dia-a-dia mais divertido com sua companhia. Obrigada especialmente ao Thiago, ao Gustavo e ao Matheus por toda a ajuda!

Às aquiculturas que doaram os peixes para a realização deste trabalho, sem os quais nada disso seria possível.

À minha família, principalmente à minha mãe Adriane que esteve sempre comigo, me dando todo o apoio necessário para que eu conseguisse concluir mais essa etapa. Nem todas as palavras do mundo são suficientes para agradecer tudo que você faz por mim!

Aos meus amigos por todo o companheirismo, principalmente Carol e Ton, que compartilharam esse período comigo. Ter vocês ao meu lado durante o mestrado foi um verdadeiro presente.

E finalmente, ao Alexandre, que me encontrou no meio dessa jornada, tornando-a mais leve! Obrigada por estar sempre ao meu lado acreditando no meu sucesso.

RESUMO

Os ecossistemas aquáticos de água doce da região Neotropical apresentam uma enorme riqueza de espécies de peixes, abrigando inúmeras espécies endêmicas. O Rio Guaraguaçu está inserido nesta região, sendo o maior rio da planície litorânea do Paraná e um importante componente do complexo estuarino LAGAMAR, área chave para conservação da Mata Atlântica. Devido a alterações de habitat, poluição da água e introdução de espécies não nativas, a fauna de pequenos peixes deste rio encontra-se ameaçada. O bagre-do-canal (*Ictalurus punctatus*) é uma espécie de peixe não nativa presente no Rio Guaraguaçu e este trabalho buscou avaliar seu potencial impacto ecológico sobre as espécies nativas. Espécies não nativas são conhecidas por causar impacto através de consumo superior de recursos, ameaçando a fauna nativa devido à predação. Uma das formas de quantificar seu impacto é comparar a resposta funcional (RF) da espécie não nativa (combinada com sua abundância/biomassa na natureza) a uma espécie nativa análoga. Portanto, aplicou-se neste trabalho a abordagem de resposta funcional comparativa (RFC) e a métrica de Impacto Potencial Relativo (IPR) para identificar os potenciais impactos da introdução de *I. punctatus* no Rio Guaraguaçu. O bagre nativo jundiá (*Rhamdia quelen*) foi escolhido como espécie nativa análoga. A espécie de presa escolhida foi o peixe de pequeno porte *Mimagoniates microlepis* utilizada como um indicador do impacto de *I. punctatus* sobre uma espécie congênera, simpátrica e sintópica ameaçada de extinção (*Mimagoniates lateralis*) que ocorre no Rio Guaraguaçu. Experimentos de RF foram realizados utilizando cinco densidades iniciais de presa (2, 5, 10, 20, e 30 indivíduos). O tipo de RF e seus parâmetros associados, taxa de ataque (a) e tempo de manuseio (h), foram estimados e comparados entre as espécies. A métrica IPR foi calculada utilizando dados de amostragens no Rio Guaraguaçu (abundância total, biomassa e captura por unidade de esforço - CPUE) obtidos da literatura. Ambas as espécies apresentaram RF Tipo II. Os intervalos de confiança das curvas estimadas se sobrepõem, porém, *I. punctatus* apresentou um h menor e uma taxa máxima de consumo maior ($1/hT$). Considerando a abundância total no ecossistema de estudo, um $IPR < 1$ foi obtido, o que indica que o bagre não nativo não representa impacto superior às presas quando comparado com seu análogo nativo. Quando utilizando dados de CPUE, porém, foi obtido um $IPR > 1$ indicando o impacto superior de *I. punctatus*. Dessa forma é possível afirmar que *I. punctatus* representa um impacto superior às presas nativas quando comparado com o nativo análogo, devido à um consumo mais eficiente e CPUE maior em campo. *Ictalurus punctatus* pode atingir tamanho corporal superior ao de *R. quelen* quando adulto, logo, um aumento no seu efeito *per capita* é esperado, aumentando seu impacto. Fica claro, a partir destes resultados, que aquiculturas utilizando espécies não nativas devem ser banidas e que a abundância de *I. punctatus* controlada, para evitar futuros impactos negativos sobre espécies endêmicas do Rio Guaraguaçu.

Palavras-chave: Água doce. Conservação. Efeitos ecológicos. Invasões biológicas. Predador-presa. Siluriformes. Utilização de recursos.

ABSTRACT

Freshwater ecosystems of the Neotropical region present a huge richness of fish species sheltering many endemic species. The Guaraguaçu River is located in this region, being the largest river of the coastal plain of Paraná state and an important component of the LAGAMAR estuarine complex, a key area for conservation of Atlantic Forest. Due to habitat alterations, water pollution and introduction of non-native species, the fauna of small fish species of this river is threatened. The channel catfish (*Ictalurus punctatus*) is one of the non-native species present in the Guaraguaçu River, and this work aimed to evaluate its potential ecological impacts. Non-native species are known to cause impacts through superior consumption of resources, threatening the native fauna by predation. A way of quantifying their impact is to compare the functional response (FR) of the non-native species (combined with its abundance/biomass in the field) to an analogue native species. Therefore, the comparative functional response (CFR) approach and the Relative Impact Potential (RIP) metric were used here to identify the potential impacts of the introduction of *I. punctatus* in the Guaraguaçu River. The South American silver catfish (*Rhamdia quelen*) was selected as the native comparator. The prey species chosen was the blue tetra *Mimagoniates microlepis* used here as a proxy of the impact of *I. punctatus* on a sympatric and syntopic endangered congeneric species (*Mimagoniates lateralis*) that occurs in Guaraguaçu River. Functional response experiments were performed using five initial densities of prey (2, 5, 10, 20, and 30 individuals). The FR type and its associated parameters attack rate (a) and handling time (h) were estimated and compared between species. The RIP metric was calculated using data from sampling in the Guaraguaçu River (total abundance, biomass and capture per unit effort - CPUE), obtained from literature. Both species presented a Type II FR. The confidence intervals of the estimated FR curves overlapped, but *I. punctatus* had a lower h and a higher maximum feeding rate ($1/hT$). Using the total abundance a $RIP < 1$ was found, which indicates that the non-native species does not represent superior impact to the native prey when compared to its native analogue. However, using CPUE data, a $RIP > 1$ was obtained, indicating that *I. punctatus* has superior impact. In this regard is possible to state that *I. punctatus* represents a superior impact to native prey when compared with the native analogue species, due to more efficient consumption and higher CPUE in the field. *Ictalurus punctatus* can reach bigger sizes than *R. quelen* when adult, so an increase in its *per capita* effect is expected, increasing its impact. It is clear, from the results that aquacultures using non-native species should be banned and the abundance of *I. punctatus* must be controlled in order to avoid future impacts on endemic species of Guaraguaçu River.

Key-words: Biological invasions. Conservation. Ecological effects. Freshwater. Predator-prey. Resource use. Siluriformes.

SUMÁRIO

APRESENTAÇÃO	6
ASSESSING THE IMPACTS OF THE CHANNEL CATFISH (<i>Ictalurus punctatus</i>) INTRODUCTION IN A HIGHLY DIVERSE NEOTROPICAL RIVER USING THE COMPARATIVE FUNCTIONAL RESPONSE APPROACH.....	10
Abstract.....	11
1. INTRODUCTION.....	12
2. METHODS	15
2.1 Species used	15
2.2 Experimental design.....	16
2.3 Statistical analysis.....	16
3. RESULTS	18
4. DISCUSSION.....	19
REFERENCES	27
SUPPORTING INFORMATION.....	41
CONSIDERAÇÕES FINAIS	43
REFERÊNCIAS	44
ANEXO	58

APRESENTAÇÃO

Os ecossistemas aquáticos de água doce na Região Neotropical apresentam uma grande riqueza de espécies de peixes, com elevado endemismo devido à sua história de isolamento geológico e biótico (LUNDBERG et al., 2000; ABBEL et al., 2008; LÉVÊQUE et al., 2008; ALBERT; PETRY; REIS, 2011). Dentro desta região, a Floresta Atlântica é um importante *hotspot* de biodiversidade abrigando um grande número de peixes, especialmente de pequeno porte, que ocorre exclusivamente nas bacias costeiras deste bioma (MYERS et al., 2000; ABILHOA et al., 2011). Inserido na Floresta Atlântica, o Rio Guaraguaçu é o maior rio da planície litorânea do Paraná e um importante componente do complexo estuarino do LAGAMAR, apresentando uma imensa variedade de habitats e abrigando muitas espécies endêmicas de peixes de pequeno porte ameaçadas de extinção, tais como *Mimagoniates lateralis*, *Spintherobolus ankoseion* e *Scleromystax macropterus* (ABILHOA; DUBOC, 2004; MACHADO; DRUMMOND; PAGLIA, 2008; VITULE, 2008; ABILHOA et al., 2011). Impactos antropogênicos como a destruição e alteração de habitat, poluição da água e introdução de espécies não nativas estão entre as principais ameaças a estas espécies (DUDGEON et al., 2006; ABILHOA et al., 2011).

Dentro deste contexto, a introdução de espécies não nativas é uma das principais causas de perda de biodiversidade global e de importantes serviços ecossistêmicos, sendo responsável por grandes prejuízos econômicos e ecológicos (VITOUSEK et al., 1996; MACK et al., 2000; LOCKWOOD et al., 2007). Especificamente, a introdução de predadores pode devastar comunidades nativas, através da extinção de espécies de presas e, sendo conseqüentemente uma das causas da homogeneização biótica (MCKINNEY; LOCKWOOD, 1999; MACK et al., 2000; BELLARD et al., 2015). Dado a relevância de espécies não nativas para a biodiversidade e economia, um dos principais desafios em ecologia é a capacidade de prever os impactos do estabelecimento destas espécies em ecossistemas fora da sua área de distribuição (PARKER et al., 1999; MACK et al., 2000; RICCIARDI et al., 2013).

A maioria dos métodos utilizados para prever impactos é bastante subjetiva e pouco eficaz, pois se baseiam no histórico de invasão ou na análise de características funcionais, dados que muitas vezes são contexto-dependentes ou

não estão disponíveis para todas as espécies (MACK et al., 2000; RICCIARDI, 2003; GARCÍA-BERTHOU, 2007; LOCKWOOD; HOOPES; MARCHETTI, 2007; HAYES; BARRY, 2008; KULHANEK et al., 2011; VITULE et al., 2012). Informações úteis sobre o potencial impacto de espécies não nativas podem ser obtidas pela comparação da resposta funcional destas com a resposta funcional de espécies nativas de níveis tróficos similares e/ou análogos a elas, uma vez que uma propriedade comum de espécies não nativas é o consumo superior dos recursos disponíveis (PARKER et al., 1999; DICK et al., 2013; SALO et al., 2007; PAOLUCCI et al., 2013). Essa abordagem quantifica o impacto diferencial de espécies não nativas, sendo mais objetiva e preditiva quando comparada a outros métodos disponíveis (BOLLACHE et al., 2008; DICK et al., 2014; 2017a).

A resposta funcional (RF) é a relação entre a densidade de recurso e a taxa de consumo deste recurso por um consumidor (HOLLING, 1959a; JULIANO, 2001). Existem três tipos principais de RF (HOLLING, 1959a; FIGURA 1) e a forma e magnitude da RF podem informar se o predador irá regular, estabilizar ou desestabilizar a população de presas através do consumo (MURDOCH; OATEN, 1975; DICK et al., 2014).

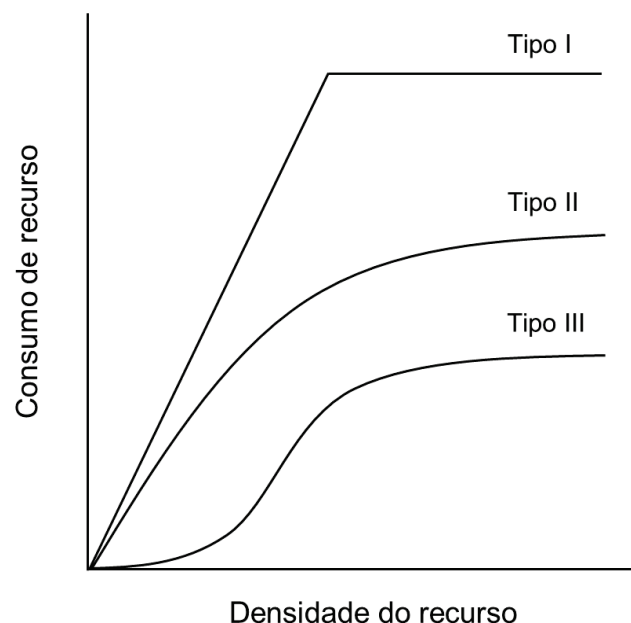


FIGURA 1. CURVAS REFERENTES AOS TRÊS TIPOS DE RESPOSTA FUNCIONAL IDENTIFICADOS POR HOLLING, 1959a.

Respostas funcionais do Tipo I são exclusivas de organismos filtradores que possuem um tempo de manuseio pequeno e uma taxa máxima de procura por

recurso constante em todas as densidades (HOLLING, 1959a; JESCHKE; KOPP; TOLLRIAN, 2004) não se aplicando, portanto a relações predador-presa. A RF do Tipo II é mais comum na natureza e tende a desestabilizar a população de presas, já que estas não conseguem “escapar” da predação mesmo em baixas densidades, e podem ser completamente consumidas, chegando à extinção (SINCLAIR et al., 1998; JULIANO, 2001). Já na RF Tipo III, as presas possuem um risco de mortalidade menor em baixa densidade, portanto, esse tipo de RF fornece estabilidade à dinâmica predador-presa, mantendo a população de presas viável abaixo de um limiar de densidade (MURDOCH; OATEN, 1975; JULIANO, 2001).

Se a espécie não nativa apresentar uma RF superior às espécies nativas análogas presentes na comunidade, é provável que a população de presas sofra uma grande redução devido ao aumento na taxa de predação, conseqüentemente refletindo-se em impacto ecológico (DICK et al., 2014). Portanto, a comparação da RF é um método eficaz para prever o potencial impacto de uma espécie não nativa na comunidade em que foi introduzida. O poder preditivo da RF pode ser aumentado considerando-se ainda a abundância e biomassa das espécies na natureza. Dick et al. (2017b) propuseram uma métrica, chamada Impacto Potencial Relativo (IPR), para avaliar o impacto de uma espécie não nativa em relação à uma espécie comparável nativa, utilizando a taxa máxima de consumo ($1/hT$) obtida experimentalmente e dados de abundância e biomassa coletados em campo.

Dado o potencial da metodologia descrita acima, em prever o impacto de espécies não nativas, ela foi aplicada neste trabalho para avaliar o impacto ecológico da introdução do bagre-do-canal (*Ictalurus punctatus*) no Rio Guaraguaçu. O bagre-do-canal é uma das espécies não nativas presentes neste rio há mais de 10 anos (VITULE; UMBRIA; ARANHA, 2005; VITULE, 2008). Esta espécie é onívora e generalista, e tem sua dieta baseada principalmente em invertebrados e peixes de pequeno porte, sendo considerada uma invasora de alto impacto (BAILEY; HARRISON JR., 1948; TYUS; NIKIRK, 1990; TOWNSEND; WINTERBOURN, 1992). O bagre-do-canal pode atingir tamanho corporal muito superior ao das espécies nativas presentes na região (JACKSON, 2004), tornando-se uma ameaça potencial às espécies de peixes ameaçadas de extinção que ocorrem no Rio Guaraguaçu.

Sendo assim, a presa nativa *Mimagoniates microlepis* que é bastante abundante na bacia do Rio Guaraguaçu e muito similar, simpátrica e sintópica à

outra espécie do mesmo gênero ameaçada de extinção (*M. lateralis*) (ABILHOA; DUBOC, 2004; BRAGA; ARANHA; VITULE, 2008; MACHADO; DRUMMOND; PAGLIA, 2008; MENEZES; WEITZMAN, 2009; VITULE 2008) foi utilizada para prever o impacto do bagre-do-canal sobre espécies de presa de pequeno porte. O objetivo é avaliar o impacto desta espécie para que novas estratégias de controle da mesma e de conservação das espécies ameaçadas presentes no rio sejam criadas. Este objetivo vem de encontro às diretrizes da Estratégia Nacional para Espécies Exóticas Invasoras no Brasil (CONABIO, 2009) e do Programa Estadual para Espécies Exóticas Invasoras do Estado do Paraná (IAP, 2008) no âmbito de geração de conhecimento científico para elaboração de planos ou medidas de ação para erradicação, contenção, controle e monitoramento de espécies não nativas.

Há uma grande lacuna de estudos sobre impactos de espécies não nativas na região tropical, principalmente no continente sul-americano, com poucos estudos utilizando abordagens experimentais (PYSEK et al., 2008; LÖVEI; LEWINSOHN, 2012; SPEZIALE et al., 2012; JUNQUEIRA, 2013; FREHSE et al., 2016). De fato, realizando uma busca nas bases Web of Science e Scopus, utilizando palavras-chave para resposta funcional, espécies não nativas e impacto ("functional response*" AND inva* OR introduc* OR "non\$native" OR alien OR exotic AND impact*) foram encontrados apenas seis estudos publicados na América do Sul e nenhum no Brasil, até Janeiro de 2018. Nós acreditamos que a abordagem utilizada aqui pode ser uma alternativa rápida e eficaz para avaliar o impacto potencial de espécies não nativas na região, e que pode ser amplamente replicada, diminuindo essa lacuna de conhecimento.

**ASSESSING THE IMPACTS OF THE CHANNEL CATFISH (*Ictalurus punctatus*)
INTRODUCTION IN A HIGHLY DIVERSE NEOTROPICAL RIVER USING THE
COMPARATIVE FUNCTIONAL RESPONSE APPROACH¹**

¹Formatado segundo as normas do periódico *Aquatic Conservation: Marine and Freshwater Ecosystems*

Assessing the impacts of the channel catfish (*Ictalurus punctatus*) introduction in a highly diverse Neotropical river using the Comparative Functional Response approach

Larissa Faria¹ & Jean R. S. Vitule²

¹Programa de Pós-Graduação em Ecologia e Conservação, Universidade Federal do Paraná, Curitiba, Brazil.

²Laboratório de Ecologia e Conservação, Departamento de Engenharia Ambiental, Universidade Federal do Paraná, Curitiba, Brazil.

Abstract

1. Non-native species are known to cause impacts through superior consumption of resources, threatening resident communities by predation. The functional response (FR) combined with the abundance/biomass of a non-native species can be compared to a native analogue and used to quantify its impact on native prey.

2. Here the comparative functional response approach (CFR) was applied to identify the potential impacts of the introduction of the non-native channel catfish (*Ictalurus punctatus*) in a Neotropical river. *Rhamdia quelen* was used as the native comparator and the prey species used was the small fish *Mimagoniates microlepis*, in five initial densities. The FR type and its associated parameters attack rate (a) and handling time (h) were estimated and compared between species. The Relative Impact Potential (RIP) metric was calculated using total abundance, biomass and capture per unit effort (CPUE) field data from literature.

3. Both species presented a Type II FR. The confidence intervals of the estimated FR curves overlapped, although h was lower for *I. punctatus* resulting in a higher maximum feeding rate ($1/hT$). Using CPUE data a $RIP > 1$ was calculated, which indicates that the non-native species represents a superior impact to prey when compared to its native analogue.

4. Despite the overlap of FR curves, *I. punctatus* represents a superior impact to native prey, due to a higher consumption efficiency and CPUE in the field. Additionally, *I. punctatus* can reach bigger sizes when adult compared to *R. quelen* which may increase its *per capita* effect. Finally, in a conservation perspective the

abundance of *I. punctatus* should be controlled and its cultivation should be banned in order to avoid future impacts on endemic species.

KEYWORDS

biological invasions, conservation, ecological effects, freshwater, predator-prey, resource use, Siluriformes

1. INTRODUCTION

Non-native species are considered the second major cause of global biodiversity loss, being one of the main drivers of species extinction (Bellard, Cassey, & Blackburn, 2016; Clavero & García-Berthou, 2005; Vitousek, D'Antonio, Loope, & Weestbrooks, 1996). Predation by non-native species, among other factors, can devastate native prey populations once non-native predators are known to have a superior consumption of available resources (Paolucci, Maclsaac, & Ricciardi, 2013; Salo, Korpimäki, Banks, Nordström, & Dickman, 2007; Simberloff & Vitule, 2014). Additionally, native prey frequently do not have effective anti-predator behaviour due to a lack of shared evolutionary history with the new predator (Cox & Lima, 2006; Simberloff & Vitule, 2014). These mechanisms can explain the establishment success of non-native predators and their negative impact in the recipient community, as declines in abundance of native populations and decreased diversity of the region (Gallardo, Clavero, Sánchez, & Vilà, 2016; Latini & Petrere-Jr, 2004; Sharpe, De León, González & Torchin, 2017; Simberloff et al., 2013; Trumpickas, Mandrak, & Ricciardi, 2011). In this regard, to predict and quantify impacts of non-native species is of great importance for conservation of biodiversity, although there is still no standard method for doing it (Kumschick et al., 2015; Parker et al., 1999; Ricciardi, Hoopes, Marchetti, & Lockwood, 2013).

Recently, Dick, Alexander, et al. (2017) claimed that the comparative functional response (CFR) approach “can unify invasion ecology” by providing a measurable trait of non-native species and their interactions with resources, that predicts its ecological impact. This framework is based on the comparison of the functional responses (FR) of non-native and trophically analogous native species, where the difference in the FR magnitude can predict and quantify the relative impacts of the non-native species (Dick et al., 2014). The functional response is the

function of consumption rate by resource density (Holling, 1959b). There are three types of FR: an increasing linear relationship between resource density and consumption until a threshold of satiation (Type I); a decelerating rate of consumption that reaches an asymptote at higher densities (Type II); and an S-shaped curve where the consumption rate first accelerates at low densities and then decelerates towards satiation (Type III) (Holling, 1959b). The FR type of the predator can have direct effects on the stability of the prey population; Type II FR has destabilizing effects with consumption constant even at low densities, whereas Type III has stabilizing effects with a refuge of predation at low densities (Oaten & Murdoch, 1975; Sinclair et al., 1998).

As FR measures the *per capita* effect of a non-native species, this measure can be combined with the abundance of the species in the field in a single metric called the Relative Impact Potential (RIP), in order to enhance the predictive power of the method (Dick, Laverty, et al., 2017). To enable a judicious selection of trophic links to assess the ecological impacts of introduced species, a framework based on foraging theory and CFR was proposed by Penk et al. (2017). This framework consists of three steps: (1) Map potential interaction partners of the focal species; (2) Screen interaction partners in the recipient community (*i.e.* abundant species, keystone species and/or species of conservation importance); and (3) Quantify interaction strengths using FR experiments (Penk et al., 2017).

These methods were employed to assess the potential impacts of a non-native species, the channel catfish (*Ictalurus punctatus*), which is found over 10 years in an Atlantic Forest basin (Occhi et al., unpublished data; Vitule, 2008; Vitule, Umbria, & Aranha, 2005). This species is native from North America and is currently introduced in South America, Europe and Asia for aquaculture and sport fishing purposes (Cruz-Spindler, Leal, Lehmann, & Schulz, 2012; Daga, Debona, Abilhoa, Gubiani, & Vitule, 2016; Elvira, 2001; Kottelat & Freyhof, 2007; Matsuzaki et al., 2011; Mejía Mojica, Paredes Lira, & Beltrán López, 2013; Zanatta, Ramos, da Silva, Langeani, & Carvalho, 2010). The species was first introduced in Brazil in 1971 and may have spread through accidental escapes from aquaculture cages (Orsi & Agostinho, 1999; Welcomme, 1988). *Ictalurus punctatus* is an omnivorous generalist species that preys mainly on vegetation, insects and fish, and a nocturnal forager that uses a variety of habitats with preference for shallow warm waters (Bailey &

Harrison Jr., 1948; Braun & Phelps, 2016; Hill, Duffy, & Thompson, 1995; Jordan, Neumann, & Schultz, 2004; Kottelat & Freyhof, 2007; Tyus & Nikirk, 1990). *Ictalurus punctatus* is monogamous and temperature is the primary factor that influences their spawning which occurs once a year, usually during the spring and early summer (Lang, Romaire, & Tiersch, 2003; Wellborn, 1988). They have high fecundity (approximately 10,000 - 20,000 eggs per female) and present parental care with the male guarding the nest (Tatarenkov, Barreto, Winkelman, & Avise, 2006; Wellborn, 1988). The species also has the ability to tolerate salinity and use estuaries as bridges for dispersion to adjacent basins (Gutierrez, Vitule, Freire, & Prodocimo, 2014). These characteristics place the *I. punctatus* as a potentially invasive species, being a threat to the native community through competition and predation of native species (Matsuzaki et al., 2011; Townsend & Winterbourn, 1992; Troca & Vieira, 2012).

The Guaraguaçu River is located in the Atlantic Forest which is one of the global hotspots of biodiversity and thus of great conservation value (Myers, Mittermeier, Mittermeier, da Fonseca, & Kent, 2000), especially for small-bodied fish species (Abilhoa, Braga, Bornatowski, & Vitule, 2011; Vitule, Braga, & Aranha, 2008). This river is the largest one of the Paraná coastal plain and shelters a high level of endemism, with many small fish species reported into the national and regional lists of threatened species such as *Mimagoniates lateralis*, *Spintherobolus ankoseion* and *Scleromystax macropterus* (Abilhoa & Duboc, 2004; Machado, Drummond, & Paglia, 2008). Therefore, the presence of *I. punctatus* in the Guaraguaçu River could increase the threat to these endangered species that already suffer the impacts of habitat alterations and water pollution in the region (Abilhoa et al., 2011; Vitule, Freire, & Simberloff, 2009; Vitule et al., 2005).

Knowing the FR type and the rate of predation is of fundamental importance to conservation management, once these data can be used to predict the degree of predator control needed to keep the endangered population of prey viable (Sinclair et al., 1998). Therefore, a prey of the resident community that is of great conservation importance was chosen in order to evaluate the potential impact of the *I. punctatus* in the Guaraguaçu River. In this regard, the potential negative impacts of *I. punctatus* predation on *M. lateralis* populations were assessed. A congeneric and sympatric species which is more abundant was used as a proxy prey species (*Mimagoniates*

microlepis) in the FR experiments in order to predict the impact of the *I. punctatus* predation on other endangered small-bodied fish.

Based on species traits and the available invasion history of *I. punctatus* (Townsend & Winterbourn, 1992), and considering its taxonomic distinctiveness within the recipient community (Ricciardi & Atkinson, 2004), we hypothesized that *I. punctatus* represents an impact to the community through superior consumption of small stream fishes. Given the research deficit about non-native species impacts in South American countries (Bellard & Jeschke, 2016; Speziale, Lambertucci, Carrete, & Tella, 2012; Vitule, Freire, Vazquez, Nuñez, & Simberloff, 2012), we aim to contribute to the understanding of the impacts of introduced predators in the Neotropical region, where biodiversity is rich and threatened; and therefore research and conservation measures are needed (Lowry et al., 2013; Myers et al., 2000).

2. METHODS

2.1 Species used

The channel catfish *I. punctatus* is an omnivorous species that feeds mainly on insects and small fishes (Bailey & Harrison Jr., 1948; Tyus & Nikirk, 1990). Thus, as a prey model for the experiments a small fish was selected, the blue tetra (*M. microlepis*) that is congeneric of *M. lateralis*, a threatened species according to the national and local lists of endangered species (Abilhoa & Duboc, 2004; Machado et al., 2008; MMA, 2014). Both *Mimagoniates* species occur in the Guaraguaçu River and are representatives of the subfamily Glandulocaudinae characterized by small stream fishes (Braga, Vitule, & Aranha, 2007; Braga, Aranha, & Vitule, 2008; Braga, Braga, & Vitule, 2013; Menezes & Weitzman, 2009; Vitule, 2008). Therefore, to avoid using an endangered species in the predation experiments, *M. microlepis* was chosen as a proxy for *M. lateralis*. Once these species are of the same subfamily, sympatric and syntopic they are very similar; and the experimental results can be extrapolated to the endangered species using the more abundant congeneric as a proxy.

FRs of the non-native *I. punctatus* were compared with a resident consumer, the South American silver catfish (*Rhamdia quelen*), popularly known in the region as 'jundiá'. This species is also a representative of the order Siluriformes, and is native in the region of interest. As others catfishes of the family, it is nocturnal and

omnivorous (Gomes, Golombieski, Gomes, & Baldisserotto, 2000). *Ictalurus punctatus* and *R. quelen* have many similarities in their anatomic digestive systems with the same food habits (Piedras, Pouey, & Moraes, 2006) which make of *R. quelen* the perfect resident comparator for *I. punctatus* in the FR experiments.

2.2 Experimental design

The experimental protocol (Fig. S1) was submitted and approved by the Ethics Commission on the Use of Animals of the Biological Sciences Department of the Federal University of Paraná (CEUA/BIO UFPR – Certificate nº1027). Juveniles of predator fish species (*I. punctatus* and *R. quelen*) were obtained in local aquaculture farms and stored in tanks of 300L in the lab. Each species was kept in a different tank with constant aeration and filtration for acclimation during 30 days. Fishes of the prey species (*M. microlepis*) were collected in the field using gill nets. They were kept in an 80L aquarium, with constant aeration and filtration, and were also acclimated for 30 days. The temperature in the lab was kept around 22-26°C with a natural light regime. During the acclimation period predators were fed every 48h with sausage to standardize prior experience.

Experiments were performed in 10L opaque plastic boxes (35 x 20 x 25 cm) with constant aeration. Individuals of predator species were randomly selected one day prior to use and placed in the experimental aquariums for 24h acclimation. They were reused in two more trials with different initial densities. Predators ($n = 9$ per species) were size-matched with respect to total length (*R. quelen*: 16.39 cm \pm 1.15 and *I. punctatus*: 16.83 cm \pm 2.14; t-test, $t = -0.55$, $df = 16$, $p = 0.59$). After the acclimation period, they were held without food for 72h to standardize hunger levels. Predators were then presented with the prey at five initial densities (2, 5, 10, 20, and 30 individuals), with at least five replicates per density. Preys were also size matched with respect to total length by visual selection (~3cm). Experiments were initiated at 14:00 and prey consumption was recorded after 24h. Controls were three replicates of each initial density of preys in the absence of predators.

2.3 Statistical analysis

The FR type was determined from a logistic regression of prey density by the proportion of consumed prey, as proposed by Juliano (2001). A negative first order term indicates a Type II FR, while a positive first order term followed by a negative

second order term indicates a Type III FR (Pritchard, Paterson, Bovy, & Barrios-O'Neill, 2017). Functional response curves and the parameters attack rate (a) and handling time (h) were modelled through Maximum Likelihood Estimation (Bolker, 2008) with the Rogers' random predator equation that considers the depletion of prey without replacement along the experimental period (Rogers, 1972):

$$N_e = N_0 \{1 - \exp [a (N_e h - T)]\} \quad (1)$$

where N_e is the number of prey consumed, N_0 is the initial density of prey, a is the attack rate, h is the handling time and T is the experimental period, given in days. The attack rate (a) is the instantaneous capture rate, *i.e.* the rate at which the consumer encounters resource items per unit of resource density; and the handling time (h) is the time spent capturing and consuming the prey item (Brose, 2010; Holling, 1959a; Jeschke, Kopp, & Tollrian, 2002). The h was used to estimate the maximum feeding rate ($1/hT$), which represents the FR curve asymptote. To compare the FR of each species, the parameters a and h were compared between predators using the indicator variables method, as the following equation:

$$0 = N_0 - N_0 \exp \{[a + Da(j)] [h + Dh(j)] (N_e) - T\} - N_e \quad (2)$$

where j is an indicator variable that takes value 0 for *R. quelen* and 1 for *I. punctatus*. The parameters Da and Dh estimate the differences between the predators in the value of the parameters a and h , respectively. If these parameters are significantly different from zero, thus the two species differ significantly in the corresponding parameters (Juliano 2001). To visualise the uncertainty around the fitted functional responses 95% confidence intervals were constructed by bootstrapping ($n=2000$) data around FR curves. If confidence intervals do not overlap, it can be stated that species FRs are different (Paterson et al., 2015). Analyses were carried in R v. 3.4.1 (R Core Team, 2015) using the 'frair' package (Pritchard et al., 2017) at 0.05 significance.

The Relative Impact Potential (RIP) metric was calculated using the following equation, according to proposed by Dick, Laverty, et al. (2017):

$$RIP = \left(\frac{FR_{non-native}}{FR_{native}} \right) \times \left(\frac{AB_{non-native}}{AB_{native}} \right) \quad (3)$$

where FR is the estimated maximum feeding rate ($1/hT$) and AB is the field abundance/biomass of the species. When $RIP < 1$, the non-native species is predicted to have less impact than the native equivalent; when $RIP = 1$, there is no impact above that driven by native equivalents; whereas $RIP > 1$ indicates a likely non-native ecological impact (Dick, Laverty, et al., 2017). Data of total abundance and biomass used are from samples collected in Guaraguaçu River using different fishing gears, and were obtained from literature (Vitule, 2008; Table S1). As data from different methods of capture and of different fishing efforts were compiled (Table S1), the capture per unit effort (CPUE) was calculated to standardize these differences, as the number of fishes captured divided by the number of fish hooks or area of gill nets multiplied by fishing effort in hours.

3. RESULTS

No prey deaths were recorded in control replicates, thus prey mortality in experimental trials was attributed to predation. Both species obtained a negative first-order term from logistic regressions, indicating a Type II FR (Table 1, Fig. 1). Parameter estimates of the FR model were all significant (Table 1). The attack rate did not differ between species ($Da = -0.646$, $z = -0.447$, $p = 0.655$), but the handling time was significantly lower for *I. punctatus* ($Dh = 0.053$, $z = 2.209$, $p < 0.05$), and thus a higher maximum feeding rate was obtained (Table 1).

Table 1. Linear coefficient (lc) from logistic regressions and functional response parameters estimates for both species preying on *Mimagoniates microlepis* (a attack rate, h handling time, $1/hT$ maximum feeding rate). SE standard error. * $p < 0.05$.

Predator	lc	a	SE	h	SE	$1/hT$
<i>Rhamdia quelen</i>	-0.0779*	2.809*	1.013	0.159*	0.021	6.3
<i>Ictalurus punctatus</i>	-0.0781*	3.456*	1.029	0.106*	0.012	9.4

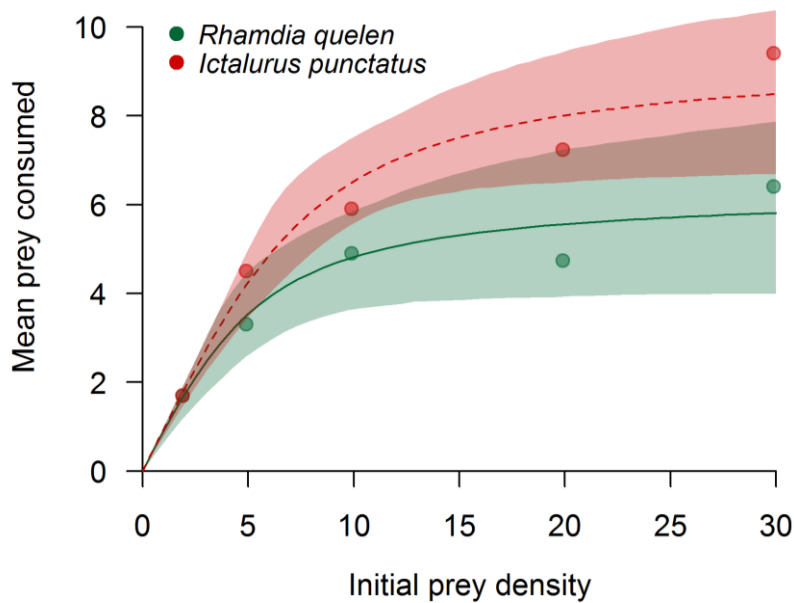


Figure 1. Type II Functional Response curves for *Ictalurus punctatus* (dashed line) and *Rhamdia quelen* (solid line) preying on *Mimagoniates microlepis*. Points indicate mean consumption per density. Shading represents bootstrapped (n=2000) 95% confidence intervals for each species.

The RIP calculated using field data on total abundance did not demonstrate superior impact of the non-native species (RIP < 1, Table 2). Considering biomass the impact of *I. punctatus* is considered similar to the native species *R. quelen* (RIP \approx 1, Table 2). However, using CPUE data, *I. punctatus* shows superior impact (RIP > 1, Table 2).

Table 2. Calculated Relative Impact Potential (RIP) for *Ictalurus punctatus* when compared with *Rhamdia quelen* preying on *Mimagoniates microlepis*. AB total abundance, CPUE capture per unit effort. Data of abundance and biomass from Vitule, 2008.

	AB	Biomass (g)	CPUE
<i>Ictalurus punctatus</i>	30	27692	0.05
<i>Rhamdia quelen</i>	112	38985	0.035
RIP	0.399	1.059	2.131

4. DISCUSSION

There is little information about the impacts of *I. punctatus* in regions where it is non-native despite it was widely introduced for aquaculture and sport fishing purposes worldwide (FAO, 2004; Vitule et al., 2009; Savini et al., 2010). The introduction of the *I. punctatus* in a highly diverse Neotropical river is cause of great

concern once this region is known as the richest realm regarding freshwater fishes, characterized by a great abundance of endemic and small species (Lindsey, 1966; Abell et al., 2008; Lévêque, Oberdorff, Paugy, Stiassny, & Tedesco, 2008). Besides threatened by habitat destruction and water pollution, these endemic species also face the danger of predation by introduced large-bodied fishes, such *I. punctatus* (MacRae & Jackson, 2001; Dudgeon et al., 2006; Vitule et al., 2009; Abilhoa et al., 2011).

To quantify and predict the impacts of non-native species is of fundamental importance for conservation, because such data is needed in order to create plans for management and control of non-native species. The CFR approach and the RIP metric have been proved to be a great tool for predicting the impact of existing and emerging high impact non-native species (Alexander, Dick, Weyl, Robinson, & Richardson, 2014; Dick, Alexander, et al., 2017; Dick, Laverty, et al., 2017; Laverty et al., 2017). Thus, in developing countries with deficient data about non-native species impacts, such as Brazil, the application of this approach is a good alternative (Lövei et al., 2012; Nuñez & Pauchard, 2010).

Using the CFR approach, evidences that the *I. punctatus* represents an impact to the native small fish prey were found. Despite the overlap of FR curves of *I. punctatus* and its native comparator (Fig. 1), it is apparent that *I. punctatus* preys more on *M. microlepis*, with a significant lower handling time, resulting in a higher maximum feeding rate (Table 1). Lower h and consequently higher $1/hT$ due to faster rates in processing prey can explain the superior impact of invaders (Alexander, Dick, et al., 2014). So, even if the superior impact of *I. punctatus* cannot be stated by differences in FR curve's magnitude, differences in their associated parameters suggest that *I. punctatus* inflict superior impact to the native community.

The native species *R. quelen* is more abundant in the Guaraguaçu River, which results in a lower relative impact of the non-native species (Table 2). However, using data standardized as CPUE a different result emerged, with *I. punctatus* presenting a higher CPUE, and thus a superior impact when compared to *R. quelen* (Table 2). To use only the FR gives a modest predictive power of non-native species impacts because a low FR can be compensated by higher abundance in the field, resulting in greater impact when compared with the native analogue; hence the combination of FR with field abundance yields more accurate predictions (Laverty et

al., 2017). The results obtained here, of a greater RIP using CPUE field data, are sufficient to justify the need of management and control of *I. punctatus* population in the Guaraguaçu River. As discussed by Simberloff (2003b) early interventions may be successful in avoiding non-native species impacts, and in this case there are enough evidences to motivate the immediate action to keep *I. punctatus* at low population levels.

Additionally, as the phenomenon of time lags in invasions is very common, it is not possible to be sure that a species that is causing no visible harm now will stay harmless in the future (Crooks, 2005; Essl et al., 2011; Strayer, Eviner, Jeschke, & Pace, 2006). That is why decisions about the control of non-native species based solely on its current perceived impact and not on its origin are very risky (Simberloff & Vitule, 2014). Furthermore, the impact of a non-native species is not always proportional to its density (Byers et al., 2002). Negative effects of *Pseudorasbora parva* on its prey were found even at low densities of *P. parva* (Jackson, Ruiz-Navarro, & Britton, 2015). So, caution must be taken when stating that the impact of a non-native species will be greater in higher abundance as approximations of density-impacts are frequently inaccurate and even at low densities the non-native species can have robust negative effects (Jackson et al., 2015; Thiele, Kollmann, Markussen, & Otte, 2010).

Relatedly, *I. punctatus* can reach bigger sizes when adult compared to *R. quelen*, so it is logical to anticipate an increased *per capita* effect with increased body size, and this feature may influence the FR results in practice. *Ictalurus punctatus* reaches its sexual maturity at 30 – 37.5 cm in length, whereas *R. quelen* generally matures at 13.4 – 17.5 cm length (Gomes et al., 2000; Jackson, 2004; Shephard & Jackson, 2005). Also, the maximum length reported for *R. quelen* was 47.4 cm and for *I. punctatus* 132 cm (Froese & Pauly, 2017), almost a three-fold difference. In the experiments this factor was controlled and juveniles of similar body size were used, but in a real situation in the field the scenario might be different. The body size seems to be more important for foraging behaviour than ontogeny in fishes and the stomach capacity of the *I. punctatus* increases in a great rate with increased body length (Gosch, Pope, & Michaletz, 2009; Miller, Crowder, Rice, & Binkowski, 1992). Predator body size influences most aspects of its feeding behaviour and there are allometric relationships between body size and the FR parameters, with increases in

a and decreases in h as predator size increases (Brose, 2010; González-Suárez et al., 2011; Miller et al., 1992; Vucic-Pestic, Rall, Kalinkat, & Brose, 2010). For instance, native and non-native predatory rates of size-matched amphipods (*Dikerogammarus villosus* and *Gammarus pulex*) were similar and the superior impact of the invasive *D. villosus* was only evident when using larger individuals in the experiments to represent natural sizes in the field (Taylor & Dunn, 2017). Therefore, size matters in the prediction of non-native species impacts and this is a factor that should always be taken into consideration (Taylor & Dunn, 2017). In the field different size classes are present, and the results using standardized lengths might be conservative (Dodd et al., 2014; Lavery et al., 2017). Taken this into account here, the different body sizes of adults of both species can be determinant to decouple the obtained FR curves; with *I. punctatus* likely having a superior response in comparison to *R. quelen*.

Differences in their biomass also corroborate the argument that the bigger sizes attained by *I. punctatus* will likely result in a superior impact. The mean biomass of *R. quelen* specimens captured in Guaraguaçu River is 348.1 g (total biomass divided by total abundance, Table 2) while for *I. punctatus* it is 923.1 g, more than twice of the native species. This difference, along with a RIP = 1 considering biomass, despite the lower total abundance of *I. punctatus* (Table 2), is evidence of *I. punctatus* bigger sizes in the field. Thus it can be expected that the *per capita* effect of *I. punctatus* is indeed superior to that of *R. quelen*. As discussed by Anderson et al. (2016) neither predator species nor size alone can predict consistently the FR type and its associated parameters, so both predator identity and size should be considered to fully assess the potential ecological impacts of a non-native species on invaded communities (Guo, Sheath, Trigo, & Britton, 2017).

Another thing that comes to attention is the similar responses of *I. punctatus* and *R. quelen* to the prey used, suggesting that these species may have niche overlap and might be competing or sharing resources in the field. The interspecific competition between native and non-native species can lead to changes in habitat selection, decreased growth rate and survival, with negative impacts for the native population (Blanchet, Loot, Grenouillet, & Brosse, 2007; Britton, Cucherousset, Grey, & Gozlan, 2011). Additionally, *I. punctatus* may be superior when competing for food with species that have a narrow diet breadth, impacting many native species through

competition (Matsuzaki et al., 2011). In the Hudson River (USA), outside its native range, *I. punctatus* seems to displace the native white catfish (*Ameiurus catus*) mainly because it has a great flexibility on habitat use according to feeding opportunity (Jordan et al., 2004). As highlighted by Dick, Alexander, et al. (2017), the CFR approach can, besides predicting the potential impact of non-native species on native prey, reveal some competitive interactions with the resident predators, and the competition between *R. quelen* and *I. punctatus* deserves further investigations.

Both species presented a Type II FR when exposed to *M. microlepis*, which means prey are consumed even at low densities leading to destabilizing effects on prey population (Holling 1959a; Murdoch & Oaten, 1975; Sinclair et al., 1998). This feature may be a matter of concern once a relevant species of prey, very similar to an endangered one was used, which classification is often based on population size (Sinclair et al., 1998; IUCN, 2012). It can be expected that the small-bodied endangered fish species present in this particular river may suffer predation pressure of *I. punctatus* in a similar manner of *M. microlepis*. Justification for conservation of smaller species that are non-commercial and not attractive to the angling community is difficult, increasing the challenge for conservation, although they are the most at risk of extinction (Cambray, 2003; Kopf, Shaw, & Humphries, 2017; Liu, Comte, & Olden, 2017; Olden, Hogan, & Zanden, 2007; Todd et al., 2017). Additionally, many tropical freshwater fish species are not evaluated on the official IUCN Red List, which would help to bring attention to their conservation (Arthington, Dulvy, Gladstone, & Winfield, 2016; Darwall & Freyhof, 2016; Vitule et al., 2017).

Furthermore, the historical and socio-cultural aspects of South American countries can contribute to a reduced concern about non-native species, impairing the conservation efforts for controlling them (Speziale et al., 2012). Also, species that do not present visible harm (mainly economic) are less prone to have their impacts studied and management plans applied (Junqueira, 2013; Simberloff, 2003a; Frehse, Braga, Nocera, & Vitule, 2016). Education and communication of scientific results about invasive species impacts are the key to solve this problem and increase the awareness about the issue, contributing to manage non-native species more effectively (Azevedo-Santos et al., 2015; Cambray, 2003; Gozlan, Burnard, Andreou, & Britton, 2013; Novoa, Dehnen-Schmutz, Fried, & Vimercati, 2017; Rodewald, 2012; Vitule, 2009). Eradication and control of non-native species are frequently costly and

impractical in larger areas, but an alternative is the construction of barriers to avoid the arrival of the species in certain areas (Cambray, 2003; Hulme, 2006; Leprieur et al., 2009; Vilà et al., 2010). In the case of Guaraguaçu River, and more broadly of the Neotropical region, education of local population combined with communication of research results to stakeholders are good starting points for reducing impacts of introduced species. Also areas where *I. punctatus* is still not present (or abundant) can be mapped and contained in order to preserve endangered small-bodied endemic fishes and the creation of new protected areas in the region is highly needed.

Along with that, the cultivation of the *I. punctatus* should be truly banned in the area, in order to reduce its spreading in natural water bodies. It is widely known that propagule pressure plays a fundamental role on the successful establishment and permanency of non-native species, contributing to its continuous impact (Kolar & Lodge, 2001; Ruesink, 2005). The juveniles of *I. punctatus* used in these experiments were obtained from local aquaculture. Aquaculture is recognized as the major pathway of introduction of freshwater invasive species (Naylor, Williams, & Strong, 2001; Casal, 2006; Gozlan, 2008) and in Brazil most of aquaculture production is based on non-native species (Lima Junior, Pelicice, Vitule, & Agostinho, 2012; Vitule, 2009). There are many fish-farming in Paraná state that cultivate and sell fry of *I. punctatus* despite a normative of the Environmental Institute of Paraná (IAP) that classified this species as Category I, which has its “*transportation, breeding, release or translocation, cultivation, propagation, commercialization, donation or intentional acquisition prohibited by any means*” (IAP, 2015). Besides that, the production of *I. punctatus* continues to increase with risk of escapes from aquaculture ponds being constant in Neotropical streams, representing a high propagule pressure (Forneck, Dutra, Zacarkim, & Cunico, 2016; Lima Junior et al., 2012; Vitule et al., 2009). This failed attempt to ban the introduction of *I. punctatus* in Paraná could be responsible for the worsened conservation status of many endemic fishes and is directly in conflict with the Aichi Targets of the Convention on Biological Diversity (Lima Junior et al., 2018; Pelicice et al., 2017; Vitule et al., 2009). Besides, the introduction of non-native catfishes in Brazil is paradoxical, once the Neotropical region has a great richness of Siluriformes that are valued for food and sport fishing (Abilhoa et al., 2011; Lévêque et al., 2008; Ota et al. 2015; Vitule, Skóra & Abilhoa,

2012). These native Siluriformes could readily replace non-native species and should have their production promoted (Cambray, 2003; Vitule et al., 2009).

As with any experimental study, there are some limitations that may be considered. A possible bias in the FR experiment results is that omnivores tend to present a Type II FR when exposed to a single type of prey (Murdoch & Oaten, 1975). For example, the presence of an alternative food changed the FR results of amphipods *D. villosus* and *G. pulex* and the superior consumption of the invasive *D. villosus* disappeared when it was presented with an alternative non-animal food (Médoc, Thuillier, & Spataro, 2017). So, for opportunistic omnivores, such as *I. punctatus*, the presence of an alternative food item might change its FR (Médoc et al., 2017) and this is a feature that should be tested in the future. Another limitation is the consideration of only variable densities of prey, instead of also considering predator-density dependence (Médoc & Spataro, 2015; Skalski & Gilliam, 2001). If conspecific predators compete interfering in each other's consumption, or if they cooperate, may be reflected in their real FR; in the first case, the impacts predicted by CFR may be overestimated, while in the latter they may be underestimated (Médoc & Spataro, 2015). The predator dependence importance on FR also remains to be tested for *I. punctatus*.

There are other context dependencies that may influence in the FR of non-native predators. The synergistic effects between climate change and invasions have been studied in the last years, revealing some insightful consequences of warming climate and altered precipitation patterns over the dispersion of non-natives (Diez et al., 2012; Dukes & Mooney, 1999; Walther et al., 2009). However, it is known that warmer temperatures can also increase the consumption rate of ectothermic species such as fishes, magnifying their impact on native preys (Rahel & Olden, 2008). Indeed, many studies revealed that the temperature directly changes the FR and its associated parameters, reflecting in lower h and higher a as the temperature increases (Pellan, Médoc, Renault, Spataro, & Piscart, 2016; Rall et al., 2012; South, Dick, McCard, Barrios-O'Neill, & Anton, 2017). Therefore, temperature is another factor that should be considered in future research about impacts of non-native species in a changing climate reality.

A different context that may alter FR results, through altered predator behaviour, is habitat complexity (Alexander, Dick, O'Connor, Haddaway, &

Farnsworth, 2012; Barrios-O'Neill, Dick, Emmerson, Ricciardi, & MacIsaac, 2015). In some cases, habitat complexity can change the FR from Type II to Type III, if the complexity acts as a refuge for preys in low density, and habitat simplification is expected to increase the impact of a non-native species (Alexander, Kaiser, Weyl, & Dick, 2015). However, the opposite may also be true if habitat complexity contributes to predation of ambush predators, resulting in a higher consumption rate in the presence of habitat structuring (Santos, García-Berthou, Hayashi, & Santos, 2013). This context-dependency factor may also be an important feature to be tested in the FR of *I. punctatus*, once the Guaraguaçu river is invaded by the African signalgrass (*Urochloa arrecta*), a macrophyte that increases habitat complexity but also promotes homogenization due to the displacement of native macrophytes (Michelan, Thomaz, Mormul, & Carvalho, 2010; Thomaz, Ribeiro, & Cunha, 2010; Vitule, Umbria, & Aranha, 2006).

The present study is important once there are few research publications about the impact of non-native species in megadiverse developing countries; and in Brazil few studies about it are performed in an experimental fashion (Bellard et al., 2016; Frehse et al., 2016; Junqueira, 2013; Lövei & Lewinsohn, 2012; Nuñez & Pauchard, 2010; Pyšek et al., 2008). To our knowledge, this is the first study using the CFR approach to evaluate the impacts of a non-native species in Brazil. The CFR approach provides a valuable assessment of the potential impacts of non-native species and can be performed elsewhere in a very simple fashion (Dick et al., 2014, Dick, Alexander, et al., 2017). We expect that this work stimulate other Brazilian researchers to use this framework in their studies about non-native species.

Negative impacts from aquatic invaders are in general difficult to detect, especially because aquatic species are not easily observed, often experience time lags and interact synergistically with other disturbances, having indirect effects on the native biodiversity and ecosystem services (Crooks, 2005; Dudgeon et al., 2006; Gallardo et al., 2016; Lima Junior et al., 2018; Vitule, 2009; Vitule, Freire, et al., 2012). Therefore, we cannot wait for the impacts to be visible in order to start confronting them and the CFR approach can be a useful tool in this endeavour (Alexander, Dick, et al., 2014; Dick et al., 2014; Simberloff & Vitule, 2014). *Ictalurus punctatus* represents a negative impact in a highly diverse Neotropical river and

measures should be taken if we want to preserve and discover all the diversity that remains to be known.

REFERENCES

- Abell, R., Thieme, M. L., Revenga, C., Bryer, M., Kottelat, M., Bogutskaya, N., ... Petry, P. (2008). Freshwater Ecoregions of the World: A New Map of Biogeographic Units for Freshwater Biodiversity Conservation. *BioScience*, 58(5), 403. Available at: <http://www.intechopen.com/books/changing-diversity-in-changing-environment/fishes-of-the-atlantic-rain-forest-streams-ecological-patterns-and-conservation>
- Abilhoa, V., Braga, R. R., Bornatowski, H., & Vitule, J. R. S. (2011). Fishes of the Atlantic Rain Forest Streams: Ecological Patterns and Conservation. In O. Grillo (Ed.), *Changing Diversity in Changing Environment* (pp. 259–282). InTech. <https://doi.org/www.intechopen.com>
- Abilhoa, V., & Duboc, L. F. (2004). Peixes. In S. B. Mikich & R. S. Bérnils (Eds.), *Livro vermelho dos animais ameaçados de extinção no Estado do Paraná* (pp. 581–677). Curitiba: Mater Natura e Instituto Ambiental do Paraná.
- Alexander, M. E., Dick, J. T. A., O'Connor, N. E., Haddaway, N. R., & Farnsworth, K. D. (2012). Functional responses of the intertidal amphipod *Echinogammarus marinus*: Effects of prey supply, model selection and habitat complexity. *Marine Ecology Progress Series*, 468, 191–202. <https://doi.org/10.3354/meps09978>
- Alexander, M. E., Dick, J. T. A., Weyl, O. L. F., Robinson, T. B., & Richardson, D. M. (2014). Existing and emerging high impact invasive species are characterized by higher functional responses than natives. *Biology Letters*, 10, 2–6. <https://doi.org/http://dx.doi.org/10.1098/rsbl.2013.0946>
- Alexander, M. E., Kaiser, H., Weyl, O. L. F., & Dick, J. T. A. (2015). Habitat simplification increases the impact of a freshwater invasive fish. *Environmental Biology of Fishes*, 98(2), 477–486. <https://doi.org/10.1007/s10641-014-0278-z>
- Anderson, T. L., Linares, C., Dodson, K. N., & Semlitsch, R. D. (2016). Variability in functional response curves among larval salamanders: Comparisons across species and size classes. *Canadian Journal of Zoology*, 94, 23–30. <https://doi.org/10.1139/cjz-2015-0149>
- Arthington, A. H., Dulvy, N. K., Gladstone, W., & Winfield, I. J. (2016). Fish conservation in freshwater and marine realms: Status, threats and management. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 26(5), 838–857. <https://doi.org/10.1002/aqc.2712>
- Azevedo-Santos, V. M., Pelicice, F. M., Lima-Junior, D. P., Magalhães, A. L. B., Orsi, M. L., Vitule, J. R. S., & Agostinho, A. A. (2015). How to avoid fish introductions in Brazil: Education and information as alternatives. *Natureza & Conservação*, 13, 123–132. <https://doi.org/10.1016/j.ncon.2015.06.002>

- Bailey, R. M., & Harrison Jr., H. M. J. (1948). Food Habits of the Southern Channel Catfish (*Ictalurus lacustris punctatus*) in the Des Moines River, Iowa. *Transactions of the American Fisheries Society*, 75(1), 110–138. [https://doi.org/10.1577/1548-8659\(1948\)75\[110:FHOTSC\]2.0.CO;2](https://doi.org/10.1577/1548-8659(1948)75[110:FHOTSC]2.0.CO;2)
- Barrios-O'Neill, D., Dick, J. T. A., Emmerson, M. C., Ricciardi, A., & MacIsaac, H. J. (2015). Predator-free space, functional responses and biological invasions. *Functional Ecology*, 29(3), 377–384. <https://doi.org/10.1111/1365-2435.12347>
- Bellard, C., Cassey, P., & Blackburn, T. M. (2016). Alien species as a driver of recent extinctions. *Biology Letters*, 12, 1–4. <https://doi.org/10.1098/rsbl.2015.0623>
- Bellard, C., & Jeschke, J. (2016). A spatial mismatch between invader impacts and research publications. *Conservation Biology*, 30(1), 230–232. <https://doi.org/10.1111/cobi.12611>
- Blanchet, S., Loot, G., Grenouillet, G., & Brosse, S. (2007). Competitive interactions between native and exotic salmonids: A combined field and laboratory demonstration. *Ecology of Freshwater Fish*, 16, 133–143. <https://doi.org/10.1111/j.1600-0633.2007.00205.x>
- Bolker, B. M. (2008). *Ecological Models and Data in R*. Princeton: Princeton University Press.
- Braga, M. R., Aranha, J. M. R., & Vitule, J. R. S. (2008). Reproduction period of *Mimagoniates microlepis*, from an Atlantic Forest stream in Southern Brazil. *Brazilian Archives of Biology and Technology*, 51(2), 345–351. <https://doi.org/10.1590/S1516-89132008000200014>
- Braga, M. R., Vitule, J. R. S., & Aranha, J. M. R. (2007). Population structure of *Mimagoniates microlepis* (Steindachner, 1876) (Characidae, Glandulocaudinae) from an Atlantic Forest stream, Paraná (Brazil). *Acta Biológica Paranaense*, 36(1–2), 67–81. <https://doi.org/10.1590/S1516-89132008000200014>
- Braga, R. R., Braga, M. R., & Vitule, J. R. S. (2013). Population structure and reproduction of *Mimagoniates microlepis* with a new hypothesis of ontogenetic migration: Implications for stream fish conservation in the Neotropics. *Environmental Biology of Fishes*, 96, 21–31. <https://doi.org/10.1007/s10641-012-0018-1>
- Braun, A. P., & Phelps, Q. E. (2016). Channel Catfish Habitat Use and Diet in the Middle Mississippi River. *The American Midland Naturalist*, 175, 47–54. <https://doi.org/10.1674/amid-175-01-47-54.1>
- Britton, J. R., Cucherousset, J., Grey, J., & Gozlan, R. E. (2011). Determining the strength of exploitative competition from an introduced fish: Roles of density, biomass and body size. *Ecology of Freshwater Fish*, 20, 74–79. <https://doi.org/10.1111/j.1600-0633.2010.00460.x>
- Brose, U. (2010). Body-mass constraints on foraging behaviour determine population and food-web dynamics. *Functional Ecology*, 24, 28–34.

<https://doi.org/10.1111/j.1365-2435.2009.01618.x>

- Byers, J. E., Reichard, S., Randall, J. M., Parker, I. M., Carey, S., Lonsdale, W., ... Hayes, D. (2002). Directing Research to Reduce the Impacts of Nonindigenous Species. *Conservation Biology*, *16*(3), 630–640. <https://doi.org/10.1046/j.1523-1739.2002.01057.x>
- Cambray, J. A. (2003). Impact on indigenous species biodiversity caused by the globalisation of alien recreational freshwater fisheries. *Hydrobiologia*, *500*, 217–230. <https://doi.org/10.1023/A:1024648719995>
- Casal, C. M. V. (2006). Global documentation of fish introductions: The growing crisis and recommendations for action. *Biological Invasions*, *8*(1), 3–11. <https://doi.org/10.1007/s10530-005-0231-3>
- Clavero, M., & García-Berthou, E. (2005). Invasive species are a leading cause of animal extinctions. *Trends in Ecology and Evolution*, *20*(3), 110. <https://doi.org/10.1016/j.tree.2005.01.003>
- Cox, J. G., & Lima, S. L. (2006). Naiveté and an aquatic-terrestrial dichotomy in the effects of introduced predators. *Trends in Ecology and Evolution*, *21*(12), 674–680. <https://doi.org/10.1016/j.tree.2006.07.011>
- Crooks, J. A. (2005). Lag times and exotic species: The ecology and management of biological invasions in slow-motion. *Ecoscience*, *12*(3), 316–329. <https://doi.org/10.2980/i1195-6860-12-3-316.1>
- Cruz-Spindler, S., Leal, M. E., Lehmann, P. C., & Schulz, U. H. (2012). First record of the exotic channel catfish *Ictalurus punctatus* (Rafinesque 1818) (Siluriformes: Ictaluridae) in the Rio dos Sinos basin, RS, Brazil. *Biota Neotropica*, *12*(3), 64–67. <https://doi.org/10.1590/S1676-06032012000300005>
- Daga, V. S., Debona, T., Abilhoa, V., Gubiani, E. A., & Vitule, J. R. S. (2016). Non-native fish invasions of a Neotropical ecoregion with high endemism: A review of the Iguazu River. *Aquatic Invasions*, *11*(2), 209–223. <https://doi.org/10.3391/ai.2016.11.2.10>
- Darwall, W. R. T., & Freyhof, J. (2016). Lost fishes, who is counting? The extent of the threat to freshwater fish biodiversity. In G. P. Closs, M. Krkosek, & J. D. Olden (Eds.), *Conservation of Freshwater Fishes* (p. 1-36). Cambridge: Cambridge University Press. <https://doi.org/10.1017/CBO9781139627085>
- Dick, J. T. A., Alexander, M. E., Jeschke, J., Ricciardi, A., Maclsaac, H. J., Robinson, T. B., ... Richardson, D. M. (2014). Advancing impact prediction and hypothesis testing in invasion ecology using a comparative functional response approach. *Biological Invasions*, *16*(4), 735–753. <https://doi.org/10.1007/s10530-013-0550-8>
- Dick, J. T. A., Alexander, M. E., Ricciardi, A., Laverty, C., Downey, P. O., Xu, M., ... Shaw, R. H. (2017). Functional responses can unify invasion ecology. *Biological Invasions*, *19*(5), 1667–1672. <https://doi.org/10.1007/s10530-016-1355-3>

- Dick, J. T. A., Laverty, C., Lennon, J. J., Barrios-O'Neill, D., Mensink, P. J., Britton, J. R., ... Caffrey, J. M. (2017). Invader Relative Impact Potential: A new metric to understand and predict the ecological impacts of existing, emerging and future invasive alien species. *Journal of Applied Ecology*, *54*(4), 1259–1267. <https://doi.org/10.1111/1365-2664.12849>
- Diez, J. M., D'Antonio, C., Dukes, J. S., Grosholz, E. D., Olden, J. D., Sorte, C. J. B., ... Miller, L. P. (2012). Will extreme climatic events facilitate biological invasions? *Frontiers in Ecology and the Environment*, *10*(5), 249–257. <https://doi.org/10.1890/110137>
- Dodd, J. A., Dick, J. T. A., Alexander, M. E., MacNeil, C., Dunn, A. M., & Aldridge, D. C. (2014). Predicting the ecological impacts of a new freshwater invader: Functional responses and prey selectivity of the “killer shrimp”, *Dikerogammarus villosus*, compared to the native *Gammarus pulex*. *Freshwater Biology*, *59*, 337–352. <https://doi.org/10.1111/fwb.12268>
- Dudgeon, D., Arthington, A. H., Gessner, M. O., Kawabata, Z.-I., Knowler, D. J., Lévêque, C., ... Sullivan, C. A. (2006). Freshwater biodiversity: Importance, threats, status and conservation challenges. *Biological Reviews*, *81*, 163–182. <https://doi.org/10.1017/S1464793105006950>
- Dukes, J. S., & Mooney, H. (1999). Does global change increase the success of biological invaders? *Trends in Ecology and Evolution*, *14*(4), 135–139. [https://doi.org/10.1016/S0169-5347\(98\)01554-7](https://doi.org/10.1016/S0169-5347(98)01554-7)
- Elvira, B. (2001). Identification of non-native freshwater fishes established in Europe and assessment of their potential threats to the biological diversity. In *Convention on the Conservation of European Wildlife and Natural Habitats* (p. 35). Strasbourg.
- Essl, F., Dullinger, S., Rabitsch, W., Hulme, P. E., Hülber, K., Jarošík, V., ... Pyšek, P. (2011). Socioeconomic legacy yields an invasion debt. *Proceedings of the National Academy of Sciences of the United States of America*, *108*(1), 203–207. <https://doi.org/10.1073/pnas.1011728108>
- FAO. (2004). *Ictalurus punctatus*. In R. R. Stickney (Ed.), *Cultured Aquatic Species Information Programme*. Rome: FAO Fisheries and Aquaculture Department [online].
- Forneck, S. C., Dutra, F. M., Zacarkim, C. E., & Cunico, A. M. (2016). Invasion risks by non-native freshwater fishes due to aquaculture activity in a Neotropical stream. *Hydrobiologia*, *773*, 193–205. <https://doi.org/10.1007/s10750-016-2699-5>
- Frehse, F. A., Braga, R. R., Nocera, G. A., & Vitule, J. R. S. (2016). Non-native species and invasion biology in a megadiverse country: Scientometric analysis and ecological interactions in Brazil. *Biological Invasions*, *18*(12), 3713–3725. <https://doi.org/10.1007/s10530-016-1260-9>
- Froese, R., & Pauly, D. (2017). FishBase. Retrieved October 1, 2017, from

www.fishbase.org

- Gallardo, B., Clavero, M., Sánchez, M. I., & Vilà, M. (2016). Global ecological impacts of invasive species in aquatic ecosystems. *Global Change Biology*, 22(1), 151–163. <https://doi.org/10.1111/gcb.13004>
- Gomes, L. C., Golombieski, J. I., Gomes, A. R. C., & Baldisserotto, B. (2000). Biology of *Rhamdia quelen* (Teleostei, Pimelodidae). *Ciência Rural*, 30(1), 179–185. <https://doi.org/10.1590/S0103-84782000000100029>
- González-Suárez, M., Mugabo, M., Decencièrre, B., Perret, S., Claessen, D., & Le Galliard, J. F. (2011). Disentangling the effects of predator body size and prey density on prey consumption in a lizard. *Functional Ecology*, 25, 158–165. <https://doi.org/10.1111/j.1365-2435.2010.01776.x>
- Gosch, N. J. C., Pope, K. L., & Michaletz, P. H. (2009). Stomach capacities of six freshwater fishes. *Journal of Freshwater Ecology*, 24(4), 645–649. <https://doi.org/10.1080/02705060.2009.9664342>
- Gozlan, R. E. (2008). Introduction of non-naive freshwater fish: is it all bad? *Fish and Fisheries*, 9, 106–115. <https://doi.org/10.1111/j.1467-2979.2007.00267.x>
- Gozlan, R. E., Burnard, D., Andreou, D., & Britton, J. R. (2013). Understanding the Threats Posed by Non-Native Species: Public vs. Conservation Managers. *PLoS ONE*, 8(1), 1–10. <https://doi.org/10.1371/journal.pone.0053200>
- Guo, Z., Sheath, D., Trigo, F. A., & Britton, J. R. (2017). Comparative functional responses of native and high-impacting invasive fishes: Impact predictions for native prey populations. *Ecology of Freshwater Fish*, 26(4), 533–540. <https://doi.org/10.1111/eff.12297>
- Gutierrez, S. M. M., Vitule, J. R. S., Freire, C. A., & Prodocimo, V. (2014). Physiological tools to predict invasiveness and spread via estuarine bridges: Tolerance of Brazilian native and worldwide introduced freshwater fishes to increased salinity. *Marine and Freshwater Research*, 65, 425–436. <https://doi.org/10.1071/MF13161>
- Hill, T. D., Duffy, W. G., & Thompson, M. R. (1995). Food-Habits of Channel Catfish in Lake Oahe, South-Dakota. *Journal of Freshwater Ecology*, 10(4), 319–323. <https://doi.org/10.1080/02705060.1995.9663454>
- Holling, C. S. (1959a). Some Characteristics of Simple Types of Predation and Parasitism. *The Canadian Entomologist*, 91(7), 385–398. <https://doi.org/10.4039/Ent91385-7>
- Holling, C. S. (1959b). The Components of Predation as Revealed by a Study of Small-Mammal Predation of the European Pine Sawfly. *The Canadian Entomologist*, 91(5), 234–261. <https://doi.org/10.4039/Ent91293-5>
- Hulme, P. E. (2006). Beyond control: Wider implications for the management of biological invasions. *Journal of Applied Ecology*, 43, 835–847.

<https://doi.org/10.1111/j.1365-2664.2006.01227.x>

- IAP. (2015). Portaria IAP n° 059, de 15 de abril de 2015. Curitiba: Instituto Ambiental do Paraná.
- IUCN. 2012. *IUCN Red List Categories and Criteria: Version 3.1*. Gland, Switzerland and Cambridge, UK.
- Jackson, D. C. (2004). Natural History and Fisheries. In C. S. Tucker & J. A. Hargreaves (Eds.), *Biology and Culture of Channel Catfish*. Amsterdam: Elsevier.
- Jackson, M. C., Ruiz-Navarro, A., & Britton, J. R. (2015). Population density modifies the ecological impacts of invasive species. *Oikos*, 124(7), 880–887. <https://doi.org/10.1111/oik.01661>
- Jeschke, J., Kopp, M., & Tollrian, R. (2002). Predator Functional Responses: Discriminating Between Handling and Digesting Prey. *Ecological Monographs*, 72(1), 95–112. [https://doi.org/10.1890/0012-9615\(2002\)072\[0095:PFRDBH\]2.0.CO;2](https://doi.org/10.1890/0012-9615(2002)072[0095:PFRDBH]2.0.CO;2)
- Jordan, S. M., Neumann, R. M., & Schultz, E. T. (2004). Distribution, habitat use, growth, and condition of a native and an introduced catfish species in the Hudson River estuary. *Journal of Freshwater Ecology*, 19(1), 59–67. <https://doi.org/10.1080/02705060.2004.9664513>
- Juliano, S. A. (2001). Nonlinear curve fitting. In S.M.Scheiner & J.Gurevitch (Eds.), *Design and Analysis of Ecological Experiments* (pp. 178–196). Oxford: Oxford University Press.
- Junqueira, A. O. R. (2013). Guilty or innocent? The need to improve the assessment of impacts caused by non-native species. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 23, 641–645. <https://doi.org/10.1002/aqc.2392>
- Kolar, C. S., & Lodge, D. M. (2001). Progress in invasion biology: Predicting invaders. *Trends in Ecology and Evolution*, 16(4), 199–204. [https://doi.org/10.1016/S0169-5347\(01\)02101-2](https://doi.org/10.1016/S0169-5347(01)02101-2)
- Kopf, R. K., Shaw, C., & Humphries, P. (2017). Trait-based prediction of extinction risk of small-bodied freshwater fishes. *Conservation Biology*, 31(3), 581–591. <https://doi.org/10.1111/cobi.12882>
- Kottelat, M., & Freyhof, J. (2007). *Handbook of European freshwater fishes*. Berlin: Kottelat, Cornol, Switzerland and Freyhof.
- Kumschick, S., Gaertner, M., Vilà, M., Essl, F., Jeschke, J., Pyšek, P., ... Winter, M. (2015). Ecological impacts of alien species: Quantification, Scope, Caveats, and Recommendations. *BioScience*, 65(1), 55–63. <https://doi.org/10.1093/biosci/biul93>
- Lang, R. P., Romaine, R. P., & Tiersch, Y. R. (2003). Induction of Early Spawning of Channel Catfish in Heated Earthen Ponds. *North American Journal of*

- Aquaculture*, 65, 73–81. [https://doi.org/10.1577/1548-8454\(2003\)65](https://doi.org/10.1577/1548-8454(2003)65)
- Latini, A. O., & Petreire Jr., M. (2004). Reduction of a native fish fauna by alien species: an example from Brazilian freshwater tropical lakes. *Fisheries Management and Ecology*, 11, 71–79. <https://doi.org/10.1046/j.1365-2400.2003.00372.x>
- Lavery, C., Green, K. D., Dick, J. T. A., Barrios-O'Neill, D., Mensink, P. J., Médoc, V., ... Gallagher, C. (2017). Assessing the ecological impacts of invasive species based on their functional responses and abundances. *Biological Invasions*, 19(5), 1653–1665. <https://doi.org/10.1007/s10530-017-1378-4>
- Leprieur, F., Brosse, S., García-Berthou, E., Oberdorff, T., Olden, J. D., & Townsend, C. R. (2009). Scientific uncertainty and the assessment of risks posed by non-native freshwater fishes. *Fish and Fisheries*, 10, 88–97. <https://doi.org/10.1111/j.1467-2979.2008.00314.x>
- Lévêque, C., Oberdorff, T., Paugy, D., Stiassny, M. L. J., & Tedesco, P. A. (2008). Global diversity of fish (Pisces) in freshwater. *Hydrobiologia*, 595, 545–567. <https://doi.org/10.1007/s10750-007-9034-0>
- Lima Junior, D. P., Magalhães, A. L. B., Pelicice, F. M., Vitule, J. R. S., Azevedo-Santos, V. M., Orsi, M. L., ... Agostinho, A. A. (2018). Aquaculture expansion in Brazilian freshwaters against the Aichi Biodiversity Targets. *Ambio*, 1–14. <https://doi.org/10.1007/s13280-017-1001-z>
- Lima Junior, D. P., Pelicice, F. M., Vitule, J. R. S., & Agostinho, A. A. (2012). Aquaculture, Politics and Environment in Brazil: New Proposals and Old Misconceptions. *Natureza & Conservação*, 10(1), 88–91. <https://doi.org/10.4322/natcon.2012.015>
- Lindsey, C. C. (1966). Body Sizes of Poikilotherm Vertebrates at Different Latitudes. *Evolution*, 20, 456–465. <https://doi.org/10.2307/2406584>
- Liu, C., Comte, L., & Olden, J. D. (2017). Heads you win, tails you lose: Life-history traits predict invasion and extinction risk of the world's freshwater fishes. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 27, 773–779. <https://doi.org/10.1002/aqc.2740>
- Lövei, G. L., Lewinsohn, T. M., & the Biological Invasions in Megadiverse Regions Network. (2012). Megadiverse developing countries face huge risks from invasives. *Trends in Ecology and Evolution*, 27(1), 2–3. <https://doi.org/10.1016/j.tree.2011.10.002>
- Lowry, E., Rollinson, E. J., Laybourn, A. J., Scott, T. E., Aiello-Lammens, M. E., Gray, S. M., ... Gurevitch, J. (2013). Biological invasions: A field synopsis, systematic review, and database of the literature. *Ecology and Evolution*, 3(1), 182–196. <https://doi.org/10.1002/ece3.431>
- Machado, A. B. M., Drummond, G. M., & Paglia, A. P. (2008). *Livro Vermelho da Fauna Brasileira Ameaçada de Extinção* (1st ed., Vol. II). Belo Horizonte:

Fundação Biodiversitas.

- MacRae, P. S. D., & Jackson, D. A. (2001). The influence of smallmouth bass (*Micropterus dolomieu*) predation and habitat complexity on the structure of littoral zone fish assemblages. *Canadian Journal of Fisheries and Aquatic Sciences*, *58*, 342–351. <https://doi.org/10.1139/cjfas-58-2-342>
- Matsuzaki, S. S., Takamura, N., Arayama, K., Tominaga, A., Iwasaki, J., & Washitani, I. (2011). Potential impacts of non-native channel catfish on commercially important species in a Japanese lake, as inferred from long-term monitoring data. *Aquatic Conservation: Marine and Freshwater Ecosystems*, *21*, 348–358. <https://doi.org/10.1002/aqc.1198>
- Médoc, V., & Spataro, T. (2015). Predicting the impact of invasive species: A look forward on the comparative functional response approach. *Revue d'Ecologie (Terre et Vie)*, *70*(12), 114–126.
- Médoc, V., Thuillier, L., & Spataro, T. (2017). Opportunistic omnivory impairs our ability to predict invasive species impacts from functional response comparisons. *Biological Invasions*. <https://doi.org/10.1007/s10530-017-1628-5>
- Mejía Mojica, H., Paredes Lira, M. E., & Beltrán López, R. G. (2013). First record and establishment of the Channel Catfish *Ictalurus punctatus* (Siluriformes: Ictaluridae) in a Tributary of the Balsas River, Mexico. *Hidrobiologica*, *23*(3), 456–459.
- Menezes, N. A., & Weitzman, S. H. (2009). Systematics of the Neotropical fish subfamily Glandulocaudinae (Teleostei: Characiformes: Characidae). *Neotropical Ichthyology*, *7*(3), 295–370. <https://doi.org/10.1590/S1679-62252009000300002>
- Michelan, T. S., Thomaz, S. M., Mormul, R. P., & Carvalho, P. (2010). Effects of an exotic invasive macrophyte (tropical signalgrass) on native plant community composition, species richness and functional diversity. *Freshwater Biology*, *55*(6), 1315–1326. <https://doi.org/10.1111/j.1365-2427.2009.02355.x>
- Miller, T. J., Crowder, L. B., Rice, J. A., & Binkowski, F. P. (1992). Body size and the ontogeny of the functional response in fishes. *Canadian Journal of Fisheries and Aquatic Sciences*, *49*, 805–812. <https://doi.org/10.1139/f92-091>
- MMA. (2014). Portaria nº 445, de 17 de dezembro de 2014. Peixes e Invertebrados Aquáticos Ameaçados. Ministério do Meio Ambiente.
- Murdoch, W. W., & Oaten, A. (1975). Predation and Population Stability. *Advances in Ecological Research*, *9*, 1–131. [https://doi.org/10.1016/S0065-2504\(08\)60288-3](https://doi.org/10.1016/S0065-2504(08)60288-3)
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A. B., & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, *403*, 853–858. <https://doi.org/10.1038/35002501>
- Naylor, R. L., Williams, S. L., & Strong, D. R. (2001). A Gateway for Exotic Species.

- Science*, 294, 1655–1656. <https://doi.org/10.1126/science.1064875>
- Novoa, A., Dehnen-Schmutz, K., Fried, J., & Vimercati, G. (2017). Does public awareness increase support for invasive species management? Promising evidence across taxa and landscape types. *Biological Invasions*, 19, 3691–3705. <https://doi.org/10.1007/s10530-017-1592-0>
- Núñez, M. A., & Pauchard, A. (2010). Biological invasions in developing and developed countries: Does one model fit all? *Biological Invasions*, 12(4), 707–714. <https://doi.org/10.1007/s10530-009-9517-1>
- Oaten, A., & Murdoch, W. W. (1975). Functional response and stability in predator-prey systems. *The American Naturalist*, 109(967), 289–298. <https://doi.org/10.1086/282998>
- Olden, J. D., Hogan, Z. S., & Zanden, M. J. Vander. (2007). Small fish, big fish, red fish, blue fish: Size-biased extinction risk of the world's freshwater and marine fishes. *Global Ecology and Biogeography*, 16, 694–701. <https://doi.org/10.1111/j.1466-8238.2007.00337.x>
- Orsi, M. L., & Agostinho, A. A. (1999). Fish species introduction by accidental escape from aquaculture in the high Paraná River Basin. *Revista Brasileira de Zoologia*, 16(2), 557–560. <https://doi.org/10.1590/S0101-81751999000200020>
- Ota, R. R., Message, H. J., Graça, W. J., & Pavanelli, C. S. (2015). Neotropical Siluriformes as a Model for Insights on Determining Biodiversity of Animal Groups. *PLoS ONE*, 10(7), 1–13. <https://doi.org/10.1371/journal.pone.0132913>
- Paolucci, E. M., Maclsaac, H. J., & Ricciardi, A. (2013). Origin matters: Alien consumers inflict greater damage on prey populations than do native consumers. *Diversity and Distributions*, 19, 988–995. <https://doi.org/10.1111/ddi.12073>
- Parker, I. M., Simberloff, D., Lonsdale, W., Goodell, K., Wonham, M., Kareiva, P., ... Goldwasser, L. (1999). Impact: Toward a framework for understanding the ecological effects of invaders. *Biological Invasions*, 1, 3–19. <https://doi.org/10.1023/A:1010034312781>
- Paterson, R. A., Dick, J. T. A., Pritchard, D. W., Ennis, M., Hatcher, M. J., & Dunn, A. M. (2015). Predicting invasive species impacts: A community module functional response approach reveals context dependencies. *Journal of Animal Ecology*, 84, 453–463. <https://doi.org/10.1111/1365-2656.12292>
- Pellicice, F. M., Azevedo-Santos, V. M., Vitule, J. R. S., Orsi, M. L., Lima Junior, D. P., Magalhães, A. L. B., ... Agostinho, A. A. (2017). Neotropical freshwater fishes imperilled by unsustainable policies. *Fish and Fisheries*, 18(6), 1119–1133. <https://doi.org/10.1111/faf.12228>
- Pellan, L., Médoc, V., Renault, D., Spataro, T., & Piscart, C. (2016). Feeding choice and predation pressure of two invasive gammarids, *Gammarus tigrinus* and *Dikerogammarus villosus*, under increasing temperature. *Hydrobiologia*, 781,

43–54. <https://doi.org/10.1007/s10750-015-2312-3>

- Penk, M., Saul, W. C., Dick, J. T. A., Donohue, I., Alexander, M. E., Linzmaier, S., & Jeschke, J. M. (2017). A trophic interaction framework for identifying the invasive capacity of novel organisms. *Methods in Ecology and Evolution*, 8(12), 1786–1794. <https://doi.org/10.1111/2041-210X.12817>
- Piedras, S. R. N., Pouey, J. L. O. F., & Moraes, P. R. R. (2006). Alimentary and reproductive behavior of cultivated exotic and native fish in the Southern region of Rio Grande do Sul. *Revista Brasileira de Agrociência*, 12(3), 341–344.
- Pritchard, D. W., Paterson, R. A., Bovy, H. C., & Barrios-O'Neill, D. (2017). frair: An R package for fitting and comparing consumer functional responses. *Methods in Ecology and Evolution*, 8(11), 1528–1534. <https://doi.org/10.1111/2041-210X.12784>
- Pyšek, P., Richardson, D. M., Pergl, J., Jarošík, V., Sixtová, Z., & Weber, E. (2008). Geographical and taxonomic biases in invasion ecology. *Trends in Ecology and Evolution*, 23(5), 237–244. <https://doi.org/10.1016/j.tree.2008.02.002>
- R Core Team (2015). R: A language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. Retrieved from <https://www.r-project.org/>
- Rahel, F. J., & Olden, J. D. (2008). Assessing the effects of climate change on aquatic invasive species. *Conservation Biology*, 22(3), 521–533. <https://doi.org/10.1111/j.1523-1739.2008.00950.x>
- Rall, B. C., Brose, U., Hartvig, M., Kalinkat, G., Schwarzmüller, F., Vucic-Pestic, O., ... Petchey, O. L. (2012). Universal temperature and body-mass scaling of feeding rates. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 367(1605), 2923–2934. <https://doi.org/10.1098/rstb.2012.0242>
- Ricciardi, A., & Atkinson, S. K. (2004). Distinctiveness magnifies the impact of biological invaders in aquatic ecosystems. *Ecology Letters*, 7, 781–784. <https://doi.org/10.1111/j.1461-0248.2004.00642.x>
- Ricciardi, A., Hoopes, M., Marchetti, M. P., & Lockwood, J. L. (2013). Progress toward understanding the ecological impacts of non-native species. *Ecological Monographs*, 83(3), 263–282. <https://doi.org/10.1890/13-0183.1>
- Rodewald, A. D. (2012). Spreading messages about invasives. *Diversity and Distributions*, 18, 97–99. <https://doi.org/10.1111/j.1472-4642.2011.00817.x>
- Rogers, D. (1972). Random Search and Insect Population Models. *Journal of Animal Ecology*, 41(2), 369–383. <https://doi.org/10.2307/3474>
- Ruesink, J. L. (2005). Global analysis of factors affecting the outcome of freshwater fish introductions. *Conservation Biology*, 19(6), 1883–1893. <https://doi.org/10.1111/j.1523-1739.2005.00289.x>

- Salo, P., Korpimäki, E., Banks, P. B., Nordström, M., & Dickman, C. R. (2007). Alien predators are more dangerous than native predators to prey populations. *Proceedings of the Royal Society B*, 274, 1237–1243. <https://doi.org/10.1098/rspb.2006.0444>
- Santos, A. F. G. N., García-Berthou, E., Hayashi, C., & Santos, L. N. (2013). When habitat complexity increases predation risk: Experiments with invasive and neotropical native fishes. *Marine and Freshwater Research*, 64(8), 752–760. <https://doi.org/10.1071/MF12264>
- Savini, B. D., Occhipinti-Ambrogi, A., Marchini, A., Tricarico, E., Gherardi, F., Olenin, S., & Gollasch, S. (2010). The top 27 animal alien species introduced into Europe for aquaculture and related activities. *Journal of Applied Ichthyology*, 26, 1–7. <https://doi.org/10.1111/j.1439-0426.2010.01503.x>
- Sharpe, D. M. T., De León, L. F., González, R., & Torchin, M. E. (2017). Tropical fish community does not recover 45 years after predator introduction. *Ecology*, 98(2), 412–424. <https://doi.org/10.1002/ecy.1648>
- Shephard, S., & Jackson, D. C. (2005). Channel Catfish Maturation in Mississippi Streams. *North American Journal of Fisheries Management*, 25, 1467–1475. <https://doi.org/10.1577/M04-139.1>
- Simberloff, D. (2003a). Confronting introduced species: A form of xenophobia? *Biological Invasions*, 5, 179–192. <https://doi.org/10.1023/A:1026164419010>
- Simberloff, D. (2003b). How much information on population biology is needed to manage introduced species? *Conservation Biology*, 17(1), 83–92. <https://doi.org/10.1046/j.1523-1739.2003.02028.x>
- Simberloff, D., Martin, J. L., Genovesi, P., Maris, V., Wardle, D. A., Aronson, J., ... Vilà, M. (2013). Impacts of biological invasions: What's what and the way forward. *Trends in Ecology and Evolution*, 28(1), 58–66. <https://doi.org/10.1016/j.tree.2012.07.013>
- Simberloff, D., & Vitule, J. R. S. (2014). A call for an end to calls for the end of invasion biology. *Oikos*, 123, 408–413. <https://doi.org/10.1111/j.1600-0706.2013.01228.x>
- Sinclair, A. R. E., Pech, R. P., Dickman, C. R., Hik, D., Mahon, P., & Newsome, A. E. (1998). Predicting effects of predation on conservation of endangered prey. *Conservation Biology*, 12(3), 564–575. <https://doi.org/10.1046/j.1523-1739.1998.97030.x>
- Skalski, G. T., & Gilliam, J. F. (2001). Functional responses with predator interference: Viable alternatives to the Holling Type II model. *Ecology*, 82(11), 3083–3092. [https://doi.org/10.1890/00129658\(2001\)082\[3083:FRWPIV\]2.0.CO;2](https://doi.org/10.1890/00129658(2001)082[3083:FRWPIV]2.0.CO;2)
- South, J., Dick, J. T. A., McCard, M., Barrios-O'Neill, D., & Anton, A. (2017). Predicting predatory impact of juvenile invasive lionfish (*Pterois volitans*) on a crustacean prey using functional response analysis: Effects of temperature,

- habitat complexity and light regimes. *Environmental Biology of Fishes*, 100, 1155–1165. <https://doi.org/10.1007/s10641-017-0633-y>
- Speziale, K. L., Lambertucci, S. A., Carrete, M., & Tella, J. L. (2012). Dealing with non-native species: What makes the difference in South America? *Biological Invasions*, 14, 1609–1621. <https://doi.org/10.1007/s10530-011-0162-0>
- Strayer, D. L., Eviner, V. T., Jeschke, J. M., & Pace, M. L. (2006). Understanding the long-term effects of species invasions. *Trends in Ecology and Evolution*, 21(11), 645–651. <https://doi.org/10.1016/j.tree.2006.07.007>
- Tatarenkov, A., Barreto, F., Winkelman, D. L., & Avise, J. C. (2006). Genetic Monogamy in the Channel Catfish, *Ictalurus punctatus*, a Species with Uniparental Nest Guarding. *Copeia*, 2006(4), 735–741. [https://doi.org/10.1643/0045-8511\(2006\)6\[735:GMITCC\]2.0.CO;2](https://doi.org/10.1643/0045-8511(2006)6[735:GMITCC]2.0.CO;2)
- Taylor, N. G., & Dunn, A. M. (2017). Size matters: Predation of fish eggs and larvae by native and invasive amphipods. *Biological Invasions*, 19(1), 89–107. <https://doi.org/10.1007/s10530-016-1265-4>
- Thiele, J., Kollmann, J., Markussen, B., & Otte, A. (2010). Impact assessment revisited - improving the theoretical basis for management of invasive alien species. *Biological Invasions*, 12(7), 2025–2035. <https://doi.org/10.1007/s10530-009-9605-2>
- Thomaz, S. M., & Cunha, E. R. (2010). The role of macrophytes in habitat structuring in aquatic ecosystems: Methods of measurement, causes and consequences on animal assemblages' composition and biodiversity. *Acta Limnologica Brasiliensia*, 22(2), 218–236. <https://doi.org/10.4322/actalb.02202011>
- Todd, C. R., Koehn, J. D., Pearce, L., Dodd, L., Humphries, P., & Morrongiello, J. R. (2017). Forgotten fishes: What is the future for small threatened freshwater fish? Population risk assessment for southern pygmy perch, *Nannoperca australis*. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 27, 1290–1300. <https://doi.org/10.1002/aqc.2808>
- Townsend, C. R., & Winterbourn, M. J. (1992). Assessment of the environmental risk posed by an exotic fish: The proposed introduction of Channel Catfish (*Ictalurus punctatus*) to New Zealand. *Conservation Biology*, 6(2), 273–282. <https://doi.org/10.1046/j.1523-1739.1992.620273.x>
- Troca, D. F. A., & Vieira, J. P. (2012). Potential invasive non-native fish farmed in the coastal region of Rio Grande do Sul, Brazil. *Boletim do Instituto de Pesca*, 38(2), 109–120.
- Trumpickas, J., Mandrak, N. E., & Ricciardi, A. (2011). Nearshore fish assemblages associated with introduced predatory fishes in lakes. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 21, 338–347. <https://doi.org/10.1002/aqc.1192>
- Tyus, H. M., & Nikirk, N. J. (1990). Abundance, Growth, and Diet of Channel Catfish,

- Ictalurus punctatus*, in the Green and Yampa Rivers, Colorado and Utah. *The Southwestern Naturalist*, 35(2), 188–198. <https://doi.org/10.2307/3671541>
- Vilà, M., Basnou, C., Pyšek, P., Josefsson, M., Genovesi, P., Gollasch, S., ... DAISIE. (2010). How well do we understand the impacts of alien species on ecosystem services? A pan-European, cross-taxa assessment. *Frontiers in Ecology and the Environment*, 8(3), 135–144. <https://doi.org/10.1890/080083>
- Vitousek, P. M., D'Antonio, C., Loope, L. L., & Westbrooks, R. (1996). Biological invasions as global environmental change. *American Scientist*, 84, 468–478.
- Vitule, J. R. S. (2008). *Distribuição, Abundância e Estrutura Populacional de Peixes Introduzidos no Rio Guaraguaçu, Paranaguá, Paraná, Brasil*. (PhD thesis). Universidade Federal do Paraná, Brazil. <https://doi.org/10.1017/CBO9781107415324.004>
- Vitule, J. R. S. (2009). Introduction of fishes in Brazilian continental ecosystems: Review, comments and suggestions for actions against the almost invisible enemy. *Neotropical Biology and Conservation*, 4(2), 111–122. <https://doi.org/10.4013/nbc.2009.42.07>
- Vitule, J. R. S., Agostinho, A. A., Azevedo-Santos, V. M., Daga, V. S., Darwall, W. R. T., Fitzgerald, D. B., ... Winemiller, K. O. (2017). We need better understanding about functional diversity and vulnerability of tropical freshwater fishes. *Biodiversity and Conservation*, 26, 757–762. <https://doi.org/10.1007/s10531-016-1258-8>
- Vitule, J. R. S., Braga, M. R., & Aranha, J. M. R. (2008). Ontogenetic, spatial and temporal variations in the feeding ecology of *Deuterodon langei* Travassos, 1957 (Teleostei: Characidae) in a Neotropical stream from the Atlantic rainforest, southern Brazil. *Neotropical Ichthyology*, 6(2), 211–222. <https://doi.org/10.1590/S1679-62252008000200008>
- Vitule, J. R. S., Freire, C. A., & Simberloff, D. (2009). Introduction of non-native freshwater fish can certainly be bad. *Fish and Fisheries*, 10(1), 98–108. <https://doi.org/10.1111/j.1467-2979.2008.00312.x>
- Vitule, J. R. S., Freire, C. A., Vazquez, D. P., Nuñez, M. A., & Simberloff, D. (2012). Revisiting the Potential Conservation Value of Non-Native Species. *Conservation Biology*, 26(6), 1153–1155. <https://doi.org/10.1111/j.1523-1739.2012.01950.x>
- Vitule, J. R. S., Skóra, F., & Abilhoa, V. (2012). Homogenization of freshwater fish faunas after the elimination of a natural barrier by a dam in Neotropics. *Diversity and Distributions*, 18, 111–120. <https://doi.org/10.1111/j.1472-4642.2011.00821.x>
- Vitule, J. R. S., Umbria, S. C., & Aranha, J. M. R. (2005). Registro de Ocorrência do Bagre do Canal *Ictalurus punctatus* (Rafinesque, 1818) em uma Importante Bacia Hidrográfica da Floresta Atlântica Paranaense. In *I Simpósio Brasileiro sobre Espécies Exóticas Invasoras*. Brasília: Ministério do Meio Ambiente.

Retrieved from <http://www.mma.gov.br/port/sbf/invasoras/index.cfm>

- Vitule, J. R. S., Umbria, S. C., & Aranha, J. M. R. (2006). Introduction of the African catfish *Clarias gariepinus* (Burchell, 1822) into Southern Brazil. *Biological Invasions*, 8, 677–681. <https://doi.org/10.1007/s10530-005-2535-8>
- Vucic-Pestic, O., Rall, B. C., Kalinkat, G., & Brose, U. (2010). Allometric functional response model: Body masses constrain interaction strengths. *Journal of Animal Ecology*, 79, 249–256. <https://doi.org/10.1111/j.1365-2656.2009.01622.x>
- Walther, G. R., Roques, A., Hulme, P. E., Sykes, M. T., Pyšek, P., Kühn, I., ... Settele, J. (2009). Alien species in a warmer world: Risks and opportunities. *Trends in Ecology and Evolution*, 24(12), 686–693. <https://doi.org/10.1016/j.tree.2009.06.008>
- Wellborn, T. L. (1988). Channel Catfish Life History and Biology. Southern Regional Aquaculture Center, *SRAC Publication*, 180, 1–4. Retrieved from <http://www2.ca.uky.edu/wkrec/CatfishBiologyHistory.pdf>
- Welcomme, R. L. (1988). *International introductions of inland aquatic species*. FAO Fisheries Technical Paper.
- Zanatta, A. S., Ramos, I. P., da Silva, R. J., Langeani, F., & Carvalho, E. D. (2010). Pisces, Siluriformes, Ictaluridae, *Ictalurus punctatus* (Rafinesque, 1818): First record in middle Paranapanema river reservoir, aquaculture and exotic species dispersion. *Check List*, 6(4), 589–591.

SUPPORTING INFORMATION

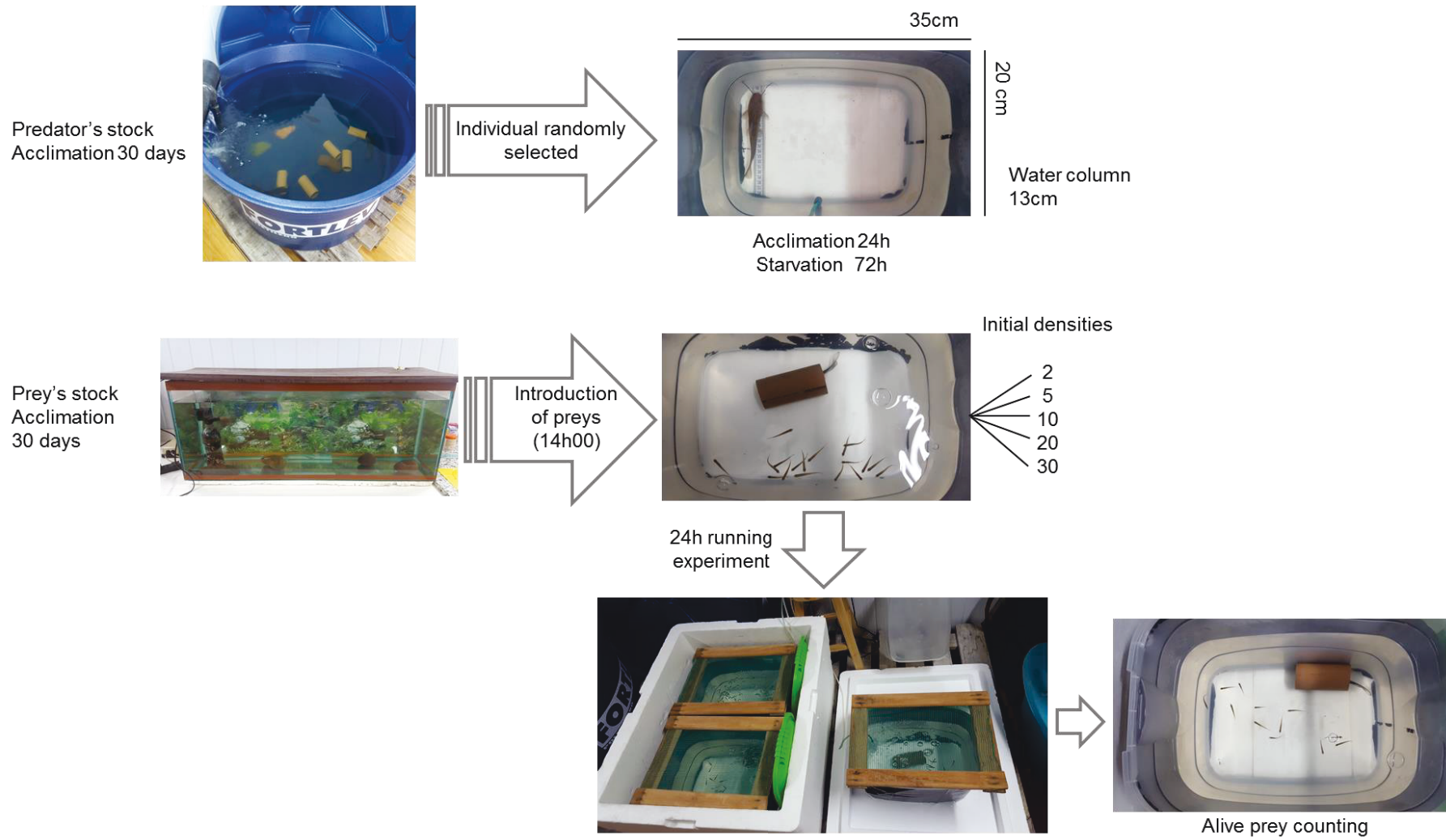


Figure S1. Experimental protocol used to obtain FR curves of *Ictalurus punctatus* and *Rhamdia quelen* preying on *Mimagoniates microlepis*.

Table S1. Field data on total abundance (n), capture per unit effort (CPUE) and total biomass (in grams) of *Ictalurus punctatus* and *Rhamdia quelen* captured in 17 standardized samples, between 2005 and 2007, using different fishing gears. Data from Vitule, 2008.

Fishing gear	Effort	n	CPUE	Biomass (g)
<i>Ictalurus punctatus</i>				
Longline	3 longlines of 10m with 10 fish hooks, 1500hrs	8	0.000178	9312
Gill net 20mm mesh size	Gill net of 30x1.70m, 1500hrs	5	6.54E-05	1069
Gill net 40mm mesh size	Gill net of 30x1.70m, 1500hrs	4	5.23E-05	2075
Gill net 60mm mesh size	Gill net of 30x1.70m, 1500hrs	1	1.31E-05	740
Fishing rod	4 anglers, aprox. 60hrs	12	0.05	14496
<i>Rhamdia quelen</i>				
Longline	3 longlines of 10m with 10 fish hooks, 1500hrs	40	0.000889	14106
Gill net 20mm mesh size	Gill net of 30x1.70m, 1500hrs	31	0.000405	8133
Gill net 40mm mesh size	Gill net of 30x1.70m, 1500hrs	33	0.000431	14768
Fishing rod	4 anglers, aprox. 60hrs	8	0.033333	1978

CONSIDERAÇÕES FINAIS

O Rio Guaraguaçu é um importante rio da costa brasileira, abrigando uma diversidade enorme de espécies de peixe de pequeno porte, porém, a introdução de espécies não nativas de predadores de grande porte, como o bagre-do-canal, ameaça esta diversidade. Acreditamos que os resultados obtidos aqui são suficientes para demonstrar a necessidade de monitoramento e controle desta espécie devido ao seu potencial impacto ecológico sobre as populações de espécies ameaçadas de extinção presentes no rio.

Avaliar e prever o impacto de espécies não nativas não é uma tarefa trivial em ecologia. A metodologia da Resposta Funcional Comparativa vem se mostrando uma boa ferramenta para isso, dentro é claro, de algumas limitações. Em um país de dimensões continentais como o Brasil, e com um alto número de espécies não nativas introduzidas, esta abordagem pode ser bastante útil para avaliar rapidamente o impacto de espécies não nativas em diferentes ecossistemas. Considerando os resultados obtidos neste trabalho, percebe-se a necessidade de estudar de maneira mais aprofundada os impactos ecológicos da introdução de espécies em ambientes aquáticos continentais, dentro de todos os contextos possíveis. Este trabalho serve de pontapé inicial para esta tarefa e ainda há muito a ser feito neste campo, porém, espera-se que ele sirva de exemplo para próximas pesquisas.

A conservação de espécies no Brasil continua sendo um grande desafio e para enfrentá-lo a comunicação de resultados científicos deve ser amplamente divulgada para que seja aplicada de maneira eficaz. Este trabalho chama a atenção para o problema do cultivo de espécies não nativas no país, o que certamente traz mais impactos negativos à biodiversidade do que positivos à economia. Esperamos que através da comunicação com tomadores de decisão e educação da população consigamos conscientizar sobre os problemas causados por espécies não nativas para que estes atores se tornem aliados no combate à introdução de espécies e à degradação da diversidade local.

REFERÊNCIAS

- ABELL, R.; THIEME, M. L.; REVENGA, C.; et al. Freshwater Ecoregions of the World: A New Map of Biogeographic Units for Freshwater Biodiversity Conservation. **BioScience**, v. 58, n. 5, p. 403, 2008.
- ABILHOA, V.; BRAGA, R. R.; BORNATOWSKI, H.; VITULE, J. R. S. Fishes of the Atlantic Rain Forest Streams: Ecological Patterns and Conservation. In: GRILLO, O. (Ed.); **Changing Diversity in Changing Environment**. InTech, 2011. p.259–282. Disponível em: <<http://www.intechopen.com/books/changing-diversity-in-changing-environment/fishes-of-the-atlantic-rain-forest-streams-ecological-patterns-and-conservation>>.
- ABILHOA, V.; DUBOC, L. F. Peixes. In: MIKICH, S. B.; BÉRNILS, R. S. (Eds.); **Livro vermelho dos animais ameaçados de extinção no Estado do Paraná**. Curitiba: Mater Natura e Instituto Ambiental do Paraná, 2004. p.581–677.
- ALBERT, J. S.; PETRY, P.; REIS, R. E. Major Biogeographic and Phylogenetic Patterns. In: ALBERT, J. S., REIS, R. E. (Eds.); **Historical Biogeography of Neotropical Freshwater Fishes** Bekerley and Los Angeles: University of California Press, 2011. p. 20-57.
- ALEXANDER, M. E.; DICK, J. T. A.; O'CONNOR, N. E.; HADDAWAY, N. R.; FARNSWORTH, K. D. Functional responses of the intertidal amphipod *Echinogammarus marinus*: Effects of prey supply, model selection and habitat complexity. **Marine Ecology Progress Series**, v. 468, p. 191–202, 2012.
- ALEXANDER, M. E.; DICK, J. T. A.; WEYL, O. L. F.; ROBINSON, T. B.; RICHARDSON, D. M. Existing and emerging high impact invasive species are characterized by higher functional responses than natives. **Biology Letters**, v. 10, p. 2–6, 2014.
- ALEXANDER, M. E.; KAISER, H.; WEYL, O. L. F.; DICK, J. T. A. Habitat simplification increases the impact of a freshwater invasive fish. **Environmental Biology of Fishes**, v. 98, n. 2, p. 477–486, 2015.
- ANDERSON, T. L.; LINARES, C.; DODSON, K. N.; SEMLITSCH, R. D. Variability in functional response curves among larval salamanders: Comparisons across species and size classes. **Canadian Journal of Zoology**, v. 94, p. 23–30, 2016.
- ARTHINGTON, A. H.; DULVY, N. K.; GLADSTONE, W.; WINFIELD, I. J. Fish conservation in freshwater and marine realms: status, threats and management. **Aquatic Conservation: Marine and Freshwater Ecosystems**, v. 26, n. 5, p. 838–857, 2016.
- AZEVEDO-SANTOS, V. M.; PELICICE, F. M.; LIMA-JUNIOR, D. P.; et al. How to avoid fish introductions in Brazil: Education and information as alternatives. **Natureza & Conservação**, v. 13, p. 123–132, 2015.

BAILEY, R. M.; HARRISON JR., H. M. J. Food Habits of the Southern Channel Catfish (*Ictalurus lacustris punctatus*) in the Des Moines River, Iowa. **Transactions of the American Fisheries Society**, v. 75, n. 1, p. 110–138, 1948.

BARRIOS-O'NEILL, D.; DICK, J. T. A.; EMMERSON, M. C.; RICCIARDI, A.; MACISAAC, H. J. Predator-free space, functional responses and biological invasions. **Functional Ecology**, v. 29, n. 3, p. 377–384, 2015.

BELLARD, C.; CASSEY, P.; BLACKBURN, T. M. Alien species as a driver of recent extinctions. **Biology Letters**, v. 12, p. 1–4, 2016.

BELLARD, C.; JESCHKE, J. A spatial mismatch between invader impacts and research publications. **Conservation Biology**, v. 30, n. 1, p. 230–232, 2016.

BLANCHET, S.; LOOT, G.; GRENOUILLET, G.; BROSSE, S. Competitive interactions between native and exotic salmonids: A combined field and laboratory demonstration. **Ecology of Freshwater Fish**, v. 16, p. 133–143, 2007.

BOLKER, B. M. **Ecological Models and Data in R**. Princeton: Princeton University Press, 2008.

BOLLACHE, L.; DICK, J. T. A.; FARNSWORTH, K. D.; MONTGOMERY, W. I. Comparison of the functional responses of invasive and native amphipods. **Biology Letters**, v. 4, n. 2, p. 166–169, 2008.

BRAGA, M. R.; ARANHA, J. M. R.; VITULE, J. R. S. Reproduction period of *Mimagoniates microlepis*, from an Atlantic Forest stream in Southern Brazil. **Brazilian Archives of Biology and Technology**, v. 51, n. 2, p. 345–351, 2008.

BRAGA, M. R.; VITULE, J. R. S.; ARANHA, J. M. R. Estrutura populacional de *Mimagoniates microlepis* (Steindachner, 1876)(Characidae, Glandulocaudinae) em um riacho de Floresta Atlântica, Paraná (Brasil). **Acta Biológica Paranaense**, v. 36, n. 1–2, p. 67–81, 2007.

BRAGA, R. R.; BRAGA, M. R.; VITULE, J. R. S. Population structure and reproduction of *Mimagoniates microlepis* with a new hypothesis of ontogenetic migration: Implications for stream fish conservation in the Neotropics. **Environmental Biology of Fishes**, v. 96, p. 21–31, 2013.

BRAUN, A. P.; PHELPS, Q. E. Channel Catfish Habitat Use and Diet in the Middle Mississippi River. **The American Midland Naturalist**, v. 175, p. 47–54, 2016.

BRITTON, J. R.; CUCHEROUSSET, J.; GREY, J.; GOZLAN, R. E. Determining the strength of exploitative competition from an introduced fish: Roles of density, biomass and body size. **Ecology of Freshwater Fish**, v. 20, p. 74–79, 2011.

BROSE, U. Body-mass constraints on foraging behaviour determine population and food-web dynamics. **Functional Ecology**, v. 24, p. 28–34, 2010.

BYERS, J. E.; REICHARD, S.; RANDALL, J. M.; et al. Directing Research to Reduce

the Impacts of Nonindigenous Species. **Conservation Biology**, v. 16, n. 3, p. 630–640, 2002.

CAMBRAY, J. A. Impact on indigenous species biodiversity caused by the globalisation of alien recreational freshwater fisheries. **Hydrobiologia**, v. 500, p. 217–230, 2003.

CASAL, C. M. V. Global documentation of fish introductions: The growing crisis and recommendations for action. **Biological Invasions**, v. 8, n. 1, p. 3–11, 2006.

CLAVERO, M.; GARCÍA-BERTHOU, E. Invasive species are a leading cause of animal extinctions. **Trends in Ecology and Evolution**, v. 20, n. 3, p. 110, 2005.

CONABIO. Resolução CONABIO nº05, de 21 de outubro de 2009. Dispõe sobre a Estratégia Nacional sobre Espécies Exóticas Invasoras. **Conselho Nacional da Biodiversidade**, Brasília, DF.

COX, J. G.; LIMA, S. L. Naiveté and an aquatic-terrestrial dichotomy in the effects of introduced predators. **Trends in Ecology and Evolution**, v. 21, n. 12, p. 674–680, 2006.

CROOKS, J. A. Lag times and exotic species: The ecology and management of biological invasions in slow-motion. **Ecoscience**, v. 12, n. 3, p. 316–329, 2005.

CRUZ-SPINDLER, S.; LEAL, M. E.; LEHMANN, P. C.; SCHULZ, U. H. First record of the exotic channel catfish *Ictalurus punctatus* (Rafinesque 1818) (Siluriformes: Ictaluridae) in the Rio dos Sinos basin, RS, Brazil. **Biota Neotropica**, v. 12, n. 3, p. 64–67, 2012.

DAGA, V. S.; DEBONA, T.; ABILHOA, V.; GUBIANI, E. A.; VITULE, J. R. S. Non-native fish invasions of a Neotropical ecoregion with high endemism: A review of the Iguaçu River. **Aquatic Invasions**, v. 11, n. 2, p. 209–223, 2016.

DARWALL, W. R. T.; FREYHOF, J. Lost fishes, who is counting? The extent of the threat to freshwater fish biodiversity. In: CLOSS; G. KRKOSEK; P.; M.; OLDEN, J. D. (Eds.); **Conservation of Freshwater Fishes**. Cambridge: Cambridge University Press, 2016. p. 1-36.

DICK, J. T. A.; ALEXANDER, M. E.; RICCIARDI, A.; et al. Functional responses can unify invasion ecology. **Biological Invasions**, v. 19, n. 5, p. 1667–1672, 2017a.

DICK, J. T. A.; GALLAGHER, K.; AVLIJAS, S.; et al. Ecological impacts of an invasive predator explained and predicted by comparative functional responses. **Biological Invasions**, v. 15, n. 4, p. 837–846, 2013.

DICK, J. T. A.; LAVERTY, C.; LENNON, J. J.; et al. Invader Relative Impact Potential: A new metric to understand and predict the ecological impacts of existing, emerging and future invasive alien species. **Journal of Applied Ecology**, v. 54, n. 4, p. 1259–1267, 2017b.

DICK, J. T. A.; ALEXANDER, M. E.; JESCHKE, J.; et al. Advancing impact prediction and hypothesis testing in invasion ecology using a comparative functional response approach. **Biological Invasions**, v. 16, n. 4, p. 735–753, 2014.

DIEZ, J. M.; D'ANTONIO, C.; DUKES, J. S.; et al. Will extreme climatic events facilitate biological invasions? **Frontiers in Ecology and the Environment**, v. 10, n. 5, p. 249–257, 2012.

DODD, J. A.; DICK, J. T. A.; ALEXANDER, M. E.; et al. Predicting the ecological impacts of a new freshwater invader: Functional responses and prey selectivity of the “killer shrimp”, *Dikerogammarus villosus*, compared to the native *Gammarus pulex*. **Freshwater Biology**, v. 59, p. 337–352, 2014.

DUDGEON, D.; ARTHINGTON, A. H.; GESSNER, M. O.; et al. Freshwater biodiversity: Importance, threats, status and conservation challenges. **Biological Reviews**, v. 81, p. 163–182, 2006.

DUKES, J. S.; MOONEY, H. Does global change increase the success of biological invaders? **Trends in Ecology and Evolution**, v. 14, n. 4, p. 135–139, 1999.

ELVIRA, B. Identification of non-native freshwater fishes established in Europe and assessment of their potential threats to the biological diversity. **Convention On The Conservation Of European Wildlife And Natural Habitats**. Strasbourg, 2001. 35p.

ESSL, F.; DULLINGER, S.; RABITSCH, W.; et al. Socioeconomic legacy yields an invasion debt. **Proceedings of the National Academy of Sciences of the United States of America**, v. 108, n. 1, p. 203–207, 2011.

FAO. *Ictalurus punctatus*. In: STICKNEY, R. R. (Ed.); **Cultured Aquatic Species Information Programme**. Rome: FAO Fisheries and Aquaculture Department [online], 2004.

FORNECK, S. C.; DUTRA, F. M.; ZACARKIM, C. E.; CUNICO, A. M. Invasion risks by non-native freshwater fishes due to aquaculture activity in a Neotropical stream. **Hydrobiologia**, v. 773, p. 193–205, 2016.

FREHSE, F. A.; BRAGA, R. R.; NOCERA, G. A.; VITULE, J. R. S. Non-native species and invasion biology in a megadiverse country: Scientometric analysis and ecological interactions in Brazil. **Biological Invasions**, v. 18, n. 12, p. 3713–3725, 2016.

FROESE, R.; PAULY., D. FishBase. Disponível em: <www.fishbase.org>. Acesso em: 1/10/2017.

GALLARDO, B.; CLAVERO, M.; SÁNCHEZ, M. I.; VILÀ, M. Global ecological impacts of invasive species in aquatic ecosystems. **Global Change Biology**, v. 22, n. 1, p. 151–163, 2016.

GARCÍA-BERTHOU, E. The characteristics of invasive fishes: What has been learned so far? **Journal of Fish Biology**, v. 71, p. 33–55, 2007.

- GOMES, L. C.; GOLOMBIESKI, J. I.; GOMES, A. R. C.; BALDISSEROTTO, B. Biologia do jundiá *Rhamdia quelen* (Teleostei, Pimelodidae). **Ciência Rural**, v. 30, n. 1, p. 179–185, 2000.
- GONZÁLEZ-SUÁREZ, M.; MUGABO, M.; DECENCIÈRE, B.; et al. Disentangling the effects of predator body size and prey density on prey consumption in a lizard. **Functional Ecology**, v. 25, p. 158–165, 2011.
- GOSCH, N. J. C.; POPE, K. L.; MICHALETZ, P. H. Stomach capacities of six freshwater fishes. **Journal of Freshwater Ecology**, v. 24, n. 4, p. 645–649, 2009.
- GOZLAN, R. E. Introduction of non-naive freshwater fish: is it all bad? **Fish and Fisheries**, v. 9, p. 106–115, 2008.
- GOZLAN, R. E.; BURNARD, D.; ANDREOU, D.; BRITTON, J. R. Understanding the Threats Posed by Non-Native Species: Public vs. Conservation Managers. **PLoS ONE**, v. 8, n. 1, p. 1–10, 2013.
- GUO, Z.; SHEATH, D.; TRIGO, F. A.; BRITTON, J. R. Comparative functional responses of native and high-impacting invasive fishes: Impact predictions for native prey populations. **Ecology of Freshwater Fish**, v. 26, n. 4, p. 533–540, 2017.
- GUTIERRE, S. M. M.; VITULE, J. R. S.; FREIRE, C. A.; PRODOCIMO, V. Physiological tools to predict invasiveness and spread via estuarine bridges: Tolerance of Brazilian native and worldwide introduced freshwater fishes to increased salinity. **Marine and Freshwater Research**, v. 65, p. 425–436, 2014.
- HAYES, K. R.; BARRY, S. C. Are there any consistent predictors of invasion success? **Biological Invasions**, v. 10, n. 4, p. 483–506, 2008.
- HILL, T. D.; DUFFY, W. G.; THOMPSON, M. R. Food-Habits of Channel Catfish in Lake Oahe, South-Dakota. **Journal of Freshwater Ecology**, v. 10, n. 4, p. 319–323, 1995.
- HOLLING, C. S. The Components of Predation as Revealed by a Study of Small-Mammal Predation of the European Pine Sawfly. **The Canadian Entomologist**, v. 91, n. 5, p. 234–261, 1959a.
- HOLLING, C. S. Some Characteristics of Simple Types of Predation and Parasitism. **The Canadian Entomologist**, v. 91, n. 7, p. 385–398, 1959b.
- HULME, P. E. Beyond control: Wider implications for the management of biological invasions. **Journal of Applied Ecology**, v. 43, p. 835–847, 2006.
- IAP. Programa Estadual para Espécies Exóticas Invasoras do Estado do Paraná. **Instituto Ambiental do Paraná**, 2008.
- IAP. Portaria nº 059, de 15 de abril de 2015. Reconhece a Lista Oficial de Espécies Exóticas Invasoras para o Estado do Paraná, estabelece normas de controle e dá

outras providências. **Instituto Ambiental do Paraná**, Curitiba, PR.

IUCN. **IUCN Red List Categories and Criteria: Version 3.1**. 2nd ed. Gland, Switzerland and Cambridge, UK, 2012.

JACKSON, D. C. Natural History and Fisheries. In: TUCKER, C. S.; HARGREAVES, J.A. (Eds.); **Biology and Culture of Channel Catfish**. Amsterdam: Elsevier, 2004.

JACKSON, M. C.; RUIZ-NAVARRO, A.; BRITTON, J. R. Population density modifies the ecological impacts of invasive species. **Oikos**, v. 124, n. 7, p. 880–887, 2015.

JESCHKE, J.; KOPP, M.; TOLLRIAN, R. Consumer-food systems: why type I functional responses are exclusive to filter feeders. **Biological Reviews**, v. 79, p. 337–349, 2004.

JORDAN, S. M.; NEUMANN, R. M.; SCHULTZ, E. T. Distribution, habitat use, growth, and condition of a native and an introduced catfish species in the Hudson River estuary. **Journal of Freshwater Ecology**, v. 19, n. 1, p. 59–67, 2004.

JULIANO, S. A. Nonlinear curve fitting. In: SCHEINER, S.M.; GUREVITCH, J. (Eds.); **Design and Analysis of Ecological Experiments**. Oxford: Oxford University Press, 2001. p.178–196.

JUNQUEIRA, A. O. R. Guilty or innocent? The need to improve the assessment of impacts caused by non-native species. **Aquatic Conservation: Marine and Freshwater Ecosystems**, v. 23, p. 641–645, 2013.

KOLAR, C. S.; LODGE, D. M. Progress in invasion biology: Predicting invaders. **Trends in Ecology and Evolution**, v. 16, n. 4, p. 199–204, 2001.

KOPF, R. K.; SHAW, C.; HUMPHRIES, P. Trait-based prediction of extinction risk of small-bodied freshwater fishes. **Conservation Biology**, v. 31, n. 3, p. 581–591, 2017.

KOTTELAT, M.; FREYHOF, J. **Handbook of European freshwater fishes**. Berlin: Kottelat, Cornol, Switzerland and Freyhof, 2007.

KULHANEK, S. A.; RICCIARDI, A.; LEUNG, B. Is invasion history a useful tool for predicting the impacts of the world's worst aquatic invasive species? **Ecological Applications**, v. 21, n. 1, p. 189–202, 2011.

KUMSCHICK, S.; GAERTNER, M.; VILÀ, M.; et al. Ecological impacts of alien species: Quantification, Scope, Caveats, and Recommendations. **BioScience**, v. 65, n. 1, p. 55–63, 2015.

LANG, R. P.; ROMAIRE, R. P.; TIERSCH, Y. R. Induction of Early Spawning of Channel Catfish in Heated Earthen Ponds. **North American Journal of Aquaculture**, v. 65, p. 73–81, 2003.

LATINI, A. O.; PETRERE JR., M. Reduction of a native fish fauna by alien species:

an example from Brazilian freshwater tropical lakes. **Fisheries Management and Ecology**, v. 11, p. 71–79, 2004.

LAVERTY, C.; GREEN, K. D.; DICK, J. T. A.; et al. Assessing the ecological impacts of invasive species based on their functional responses and abundances. **Biological Invasions**, v. 19, n. 5, p. 1653–1665, 2017.

LEPRIEUR, F.; BROSSE, S.; GARCÍA-BERTHOU, E.; et al. Scientific uncertainty and the assessment of risks posed by non-native freshwater fishes. **Fish and Fisheries**, v. 10, p. 88–97, 2009.

LÉVÊQUE, C.; OBERDORFF, T.; PAUGY, D.; STIASSNY, M. L. J.; TEDESCO, P. A. Global diversity of fish (Pisces) in freshwater. **Hydrobiologia**, v. 595, p. 545–567, 2008.

LIMA JUNIOR, D. P.; MAGALHÃES, A. L. B.; PELICICE, F. M.; et al. Aquaculture expansion in Brazilian freshwaters against the Aichi Biodiversity Targets. **Ambio**, p. 1–14, 2018. doi: 10.1007/s13280-017-1001-z

LIMA JUNIOR, D. P.; PELICICE, F. M.; VITULE, J. R. S.; AGOSTINHO, A. A. Aquicultura, Política e Meio Ambiente no Brasil: Novas Propostas e Velhos Equívocos. **Natureza & Conservação**, v. 10, n. 1, p. 88–91, 2012.

LINDSEY, C. C. Body Sizes of Poikilotherm Vertebrates at Different Latitudes. **Evolution**, v. 20, p. 456–465, 1966.

LIU, C.; COMTE, L.; OLDEN, J. D. Heads you win, tails you lose: Life-history traits predict invasion and extinction risk of the world's freshwater fishes. **Aquatic Conservation: Marine and Freshwater Ecosystems**, v. 27, p. 773–779, 2017.

LOCKWOOD, J. L.; HOOPES, M.; MARCHETTI, M. **Invasion Ecology**. Oxford: Blackwell Publishing, 2007.

LÖVEI, G. L.; LEWINSOHN, T. M.; THE BIOLOGICAL INVASIONS IN MEGADIVERSE REGIONS NETWORK. Megadiverse developing countries face huge risks from invasives. **Trends in Ecology and Evolution**, v. 27, n. 1, p. 2–3, 2012.

LOWRY, E.; ROLLINSON, E. J.; LAYBOURN, A. J.; et al. Biological invasions: A field synopsis, systematic review, and database of the literature. **Ecology and Evolution**, v. 3, n. 1, p. 182–196, 2013.

LUNDBERG, J. G.; KOTTELAT, M.; SMITH, G. R.; STIASSNY, M. L. J.; GILL, A. C. So Many Fishes, So Little Time: An Overview of Recent Ichthyological Discovery in Continental Waters. **Annals of the Missouri Botanical Garden**, v. 87, n. 1, p. 26, 2000.

MACHADO, A. B. M.; DRUMMOND, G. M.; PAGLIA, A. P. **Livro Vermelho da Fauna Brasileira Ameaçada de Extinção**. 1 ed. Belo Horizonte: Fundação Biodiversitas, 2008.

MACK, R. N.; SIMBERLOFF, D.; LONSDALE, W.; et al. Biotic Invasions: Causes, Epidemiology, Global Consequences, and Control. **Ecological Applications**, v. 10, n. 3, p. 689–710, 2000.

MACRAE, P. S. D.; JACKSON, D. A. The influence of smallmouth bass (*Micropterus dolomieu*) predation and habitat complexity on the structure of littoral zone fish assemblages. **Canadian Journal of Fisheries and Aquatic Sciences**, v. 58, p. 342–351, 2001.

MATSUZAKI, S. S.; TAKAMURA, N.; ARAYAMA, K.; et al. Potential impacts of non-native channel catfish on commercially important species in a Japanese lake, as inferred from long-term monitoring data. **Aquatic Conservation: Marine and Freshwater Ecosystems**, v. 21, p. 348–358, 2011.

MCKINNEY, M. L.; LOCKWOOD, J. L. Biotic homogenization: A few winners replacing many losers in the next mass extinction. **Trends in Ecology and Evolution**, v. 14, n. 11, p. 450–453, 1999.

MÉDOC, V.; SPATARO, T. Predicting the Impact of Invasive Species: A Look Forward on the Comparative Functional Response Approach. **Revue d'Ecologie (Terre et Vie)**, v. 70, n. 12, p. 114–126, 2015.

MÉDOC, V.; THUILLIER, L.; SPATARO, T. Opportunistic omnivory impairs our ability to predict invasive species impacts from functional response comparisons. **Biological Invasions**, 2017. <https://doi.org/10.1007/s10530-017-1628-5>

MEJÍA MOJICA, H.; PAREDES LIRA, M. E.; BELTRÁN LÓPEZ, R. G. Primer registro y establecimiento del bagre de canal *Ictalurus punctatus* (Siluriformes: Ictaluridae) en un tributario del Río Balsas, México. **Hidrobiologica**, v. 23, n. 3, p. 456–459, 2013.

MENEZES, N. A.; WEITZMAN, S. H. Systematics of the Neotropical fish subfamily Glandulocaudinae (Teleostei: Characiformes: Characidae). **Neotropical Ichthyology**, v. 7, n. 3, p. 295–370, 2009.

MICHELAN, T. S.; THOMAZ, S. M.; MORMUL, R. P.; CARVALHO, P. Effects of an exotic invasive macrophyte (tropical signalgrass) on native plant community composition, species richness and functional diversity. **Freshwater Biology**, v. 55, n. 6, p. 1315–1326, 2010.

MILLER, T. J.; CROWDER, L. B.; RICE, J. A.; BINKOWSKI, F. P. Body size and the ontogeny of the functional response in fishes. **Canadian Journal of Fisheries and Aquatic Sciences**, v. 49, p. 805–812, 1992.

MMA. Portaria n^o 445, de 17 de dezembro de 2014. Peixes e Invertebrados Aquáticos Ameaçados, 2014. **Ministério do Meio Ambiente**.

MURDOCH, W. W.; OATEN, A. Predation and Population Stability. **Advances in Ecological Research**, v. 9, p. 1–131, 1975.

MYERS, N.; MITTERMEIER, R. A.; MITTERMEIER, C. G.; DA FONSECA, G. A. B.; KENT, J. Biodiversity hotspots for conservation priorities. **Nature**, v. 403, p. 853–858, 2000.

NAYLOR, R. L.; WILLIAMS, S. L.; STRONG, D. R. A Gateway for Exotic Species. **Science**, v. 294, p. 1655–1656, 2001.

NOVOA, A.; DEHNEN-SCHMUTZ, K.; FRIED, J.; VIMERCATI, G. Does public awareness increase support for invasive species management? Promising evidence across taxa and landscape types. **Biological Invasions**, v. 19, p. 3691–3705, 2017.

NUÑEZ, M. A.; PAUCHARD, A. Biological invasions in developing and developed countries: Does one model fit all? **Biological Invasions**, v. 12, n. 4, p. 707–714, 2010.

OATEN, A.; MURDOCH, W. W. Functional Response and Stability in Predator-Prey Systems. **The American Naturalist**, v. 109, n. 967, p. 289–298, 1975.

OLDEN, J. D.; HOGAN, Z. S.; ZANDEN, M. J. VANDER. Small fish, big fish, red fish, blue fish: Size-biased extinction risk of the world's freshwater and marine fishes. **Global Ecology and Biogeography**, v. 16, n. 6, p. 694–701, 2007.

ORSI, M. L.; AGOSTINHO, A. A. Introdução de espécies de peixes por escapes acidentais de tanques de cultivo em rios da Bacia do Rio Paraná, Brasil. **Revista Brasileira de Zoologia**, v. 16, n. 2, p. 557–560, 1999.

OTA, R. R.; MESSAGE, H. J.; GRAÇA, W. J.; PAVANELLI, C. S. Neotropical Siluriformes as a Model for Insights on Determining Biodiversity of Animal Groups. **PLoS ONE**, v. 10, n. 7, p. 1–13, 2015.

PAOLUCCI, E. M.; MACISAAC, H. J.; RICCIARDI, A. Origin matters: Alien consumers inflict greater damage on prey populations than do native consumers. **Diversity and Distributions**, v. 19, p. 988–995, 2013.

PARKER, I. M.; SIMBERLOFF, D.; LONSDALE, W.; et al. Impact: Toward a framework for understanding the ecological effects of invaders. **Biological Invasions**, v. 1, p. 3–19, 1999.

PATERSON, R. A.; DICK, J. T. A.; PRITCHARD, D. W.; et al. Predicting invasive species impacts: A community module functional response approach reveals context dependencies. **Journal of Animal Ecology**, v. 84, p. 453–463, 2015.

PELICICE, F. M.; AZEVEDO-SANTOS, V. M.; VITULE, J. R. S.; et al. Neotropical freshwater fishes imperilled by unsustainable policies. **Fish and Fisheries**, v. 18, n. 6, p. 1119–1133, 2017.

PELLAN, L.; MÉDOC, V.; RENAULT, D.; SPATARO, T.; PISCART, C. Feeding choice and predation pressure of two invasive gammarids, *Gammarus tigrinus* and *Dikerogammarus villosus*, under increasing temperature. **Hydrobiologia**, v. 781, p. 43–54, 2016.

PENK, M.; SAUL, W.-C.; DICK, J. T. A.; et al. A trophic interaction framework for identifying the invasive capacity of novel organisms. **Methods in Ecology and Evolution**, p. 1–9, 2017.

PIEDRAS, S. R. N.; POUHEY, J. L. O. F.; MORAES, P. R. R. Comportamento Alimentar e Reprodutivo de Peixes Exóticos e Nativos Cultivados na Zona Sul do Rio Grande do Sul. **Revista Brasileira de Agrociência**, v. 12, n. 3, p. 341–344, 2006.

PRITCHARD, D. W.; PATERSON, R. A.; BOVY, H. C.; BARRIOS-O'NEILL, D. frair: An R package for fitting and comparing consumer functional responses. **Methods in Ecology and Evolution**, p. 1–7, 2017.

PYŠEK, P.; RICHARDSON, D. M.; PERGL, J.; et al. Geographical and taxonomic biases in invasion ecology. **Trends in Ecology and Evolution**, v. 23, n. 5, p. 237–244, 2008.

R CORE TEAM (2015). R: A language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. <https://www.r-project.org/>

RAHEL, F. J.; OLDEN, J. D. Assessing the effects of climate change on aquatic invasive species. **Conservation Biology**, v. 22, n. 3, p. 521–533, 2008.

RALL, B. C.; BROSE, U.; HARTVIG, M.; et al. Universal temperature and body-mass scaling of feeding rates. **Philosophical Transactions of the Royal Society B: Biological Sciences**, v. 367, p. 2923–2934, 2012.

RICCIARDI, A. Predicting the impacts of an introduced species from its invasion history: An empirical approach applied to zebra mussel invasions. **Freshwater Biology**, v. 48, n. 6, p. 972–981, 2003.

RICCIARDI, A.; ATKINSON, S. K. Distinctiveness magnifies the impact of biological invaders in aquatic ecosystems. **Ecology Letters**, v. 7, p. 781–784, 2004.

RICCIARDI, A.; HOOPES, M.; MARCHETTI, M. P.; LOCKWOOD, J. L. Progress toward understanding the ecological impacts of non-native species. **Ecological Monographs**, v. 83, n. 3, p. 263–282, 2013.

RODEWALD, A. D. Spreading messages about invasives. **Diversity and Distributions**, v. 18, p. 97–99, 2012.

ROGERS, D. Random Search and Insect Population Models. **Journal of Animal Ecology**, v. 41, n. 2, p. 369–383, 1972.

RUESINK, J. L. Global analysis of factors affecting the outcome of freshwater fish introductions. **Conservation Biology**, v. 19, n. 6, p. 1883–1893, 2005.

SALO, P.; KORPIMÄKI, E.; BANKS, P. B.; NORDSTRÖM, M.; DICKMAN, C. R. Alien predators are more dangerous than native predators to prey populations. **Proceedings of the Royal Society B**, v. 274, p. 1237–1243, 2007.

SANTOS, A. F. G. N.; GARCÍA-BERTHOU, E.; HAYASHI, C.; SANTOS, L. N. When habitat complexity increases predation risk: Experiments with invasive and neotropical native fishes. **Marine and Freshwater Research**, v. 64, n. 8, p. 752–760, 2013.

SAVINI, B. D.; OCCHIPINTI-AMBROGI, A.; MARCHINI, A.; et al. The top 27 animal alien species introduced into Europe for aquaculture and related activities. **Journal of Applied Ichthyology**, v. 26, p. 1–7, 2010.

SHARPE, D. M. T.; DE LEÓN, L. F.; GONZÁLEZ, R.; TORCHIN, M. E. Tropical fish community does not recover 45 years after predator introduction. **Ecology**, v. 98, n. 2, p. 412–424, 2017.

SHEPARD, S.; JACKSON, D. C. Channel Catfish Maturation in Mississippi Streams. **North American Journal of Fisheries Management**, v. 25, p. 1467–1475, 2005.

SIMBERLOFF, D. Confronting introduced species: A form of xenophobia? **Biological Invasions**, v. 5, p. 179–192, 2003a.

SIMBERLOFF, D. How much information on population biology is needed to manage introduced species? **Conservation Biology**, v. 17, n. 1, p. 83–92, 2003b.

SIMBERLOFF, D.; MARTIN, J. L.; GENOVESI, P.; et al. Impacts of biological invasions: What's what and the way forward. **Trends in Ecology and Evolution**, v. 28, n. 1, p. 58–66, 2013.

SIMBERLOFF, D.; VITULE, J. R. S. A call for an end to calls for the end of invasion biology. **Oikos**, v. 123, p. 408–413, 2014.

SINCLAIR, A. R. E.; PECH, R. P.; DICKMAN, C. R.; et al. Predicting effects of predation on conservation of endangered prey. **Conservation Biology**, v. 12, n. 3, p. 564–575, 1998.

SKALSKI, G. T.; GILLIAM, J. F. Functional responses with predator interference: Viable alternatives to the Holling Type II model. **Ecology**, v. 82, n. 11, p. 3083–3092, 2001.

SOUTH, J.; DICK, J. T. A.; MCCARD, M.; BARRIOS-O'NEILL, D.; ANTON, A. Predicting predatory impact of juvenile invasive lionfish (*Pterois volitans*) on a crustacean prey using functional response analysis: Effects of temperature, habitat complexity and light regimes. **Environmental Biology of Fishes**, v. 100, p. 1155–1165, 2017.

SPEZIALE, K. L.; LAMBERTUCCI, S. A.; CARRETE, M.; TELLA, J. L. Dealing with non-native species: What makes the difference in South America? **Biological Invasions**, v. 14, p. 1609–1621, 2012.

STRAYER, D. L.; EVINER, V. T.; JESCHKE, J. M.; PACE, M. L. Understanding the

long-term effects of species invasions. **Trends in Ecology and Evolution**, v. 21, n. 11, p. 645–651, 2006.

TATARENKOV, A.; BARRETO, F.; WINKELMAN, D. L.; AVISE, J. C. Genetic Monogamy in the Channel Catfish, *Ictalurus punctatus*, a Species with Uniparental Nest Guarding. **Copeia**, n. 4, p. 735–741, 2006.

TAYLOR, N. G.; DUNN, A. M. Size matters: Predation of fish eggs and larvae by native and invasive amphipods. **Biological Invasions**, v. 19, n. 1, p. 89–107, 2017. Springer International Publishing.

THIELE, J.; KOLLMANN, J.; MARKUSSEN, B.; OTTE, A. Impact assessment revisited - improving the theoretical basis for management of invasive alien species. **Biological Invasions**, v. 12, n. 7, p. 2025–2035, 2010.

THOMAZ, S. M.; CUNHA, E. R. The role of macrophytes in habitat structuring in aquatic ecosystems: Methods of measurement, causes and consequences on animal assemblages' composition and biodiversity. **Acta Limnologica Brasiliensia**, v. 22, n. 2, p. 218–236, 2010.

TODD, C. R.; KOEHN, J. D.; PEARCE, L.; et al. Forgotten fishes: What is the future for small threatened freshwater fish? Population risk assessment for southern pygmy perch, *Nannoperca australis*. **Aquatic Conservation: Marine and Freshwater Ecosystems**, v. 27, p. 1290–1300, 2017.

TOWNSEND, C. R.; WINTERBOURN, M. J. Assessment of the environmental risk posed by an exotic fish: The proposed introduction of Channel Catfish (*Ictalurus punctatus*) to New Zealand. **Conservation Biology**, v. 6, n. 2, p. 273–282, 1992.

TROCA, D. F. A.; VIEIRA, J. P. Potencial Invasor dos Peixes Não Nativos Cultivados na Região Costeira do Rio Grande do Sul, Brasil. **Boletim do Instituto de Pesca**, v. 38, n. 2, p. 109–120, 2012.

TRUMPICKAS, J.; MANDRAK, N. E.; RICCIARDI, A. Nearshore fish assemblages associated with introduced predatory fishes in lakes. **Aquatic Conservation: Marine and Freshwater Ecosystems**, v. 21, p. 338–347, 2011.

TYUS, H. M.; NIKIRK, N. J. Abundance, Growth, and Diet of Channel Catfish, *Ictalurus punctatus*, in the Green and Yampa Rivers, Colorado and Utah. **The Southwestern Naturalist**, v. 35, n. 2, p. 188–198, 1990.

VILÀ, M.; BASNOU, C.; PYŠEK, P.; et al. How well do we understand the impacts of alien species on ecosystem services? A pan-European, cross-taxa assessment. **Frontiers in Ecology and the Environment**, v. 8, n. 3, p. 135–144, 2010.

VITOUSEK, P. M.; D'ANTONIO, C.; LOOPE, L. L.; WESTBROOKS, R. Biological invasions as global environmental change. **American Scientist**, v. 84, p. 468–478, 1996.

VITULE, J. R. S. **Distribuição, Abundância e Estrutura Populacional de Peixes**

Introduzidos no Rio Guaraguaçu, Paranaguá, Paraná, Brasil. 143f. Tese (Doutorado em Zoologia) – Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, 2008.

VITULE, J. R. S. Introdução de peixes em ecossistemas continentais brasileiros: revisão, comentários e sugestões de ações contra o inimigo quase invisível. **Neotropical Biology and Conservation**, v. 4, n. 2, p. 111–122, 2009.

VITULE, J. R. S.; AGOSTINHO, A. A.; AZEVEDO-SANTOS, V. M.; et al. We need better understanding about functional diversity and vulnerability of tropical freshwater fishes. **Biodiversity and Conservation**, v. 26, p. 757–762, 2017.

VITULE, J. R. S.; BRAGA, M. R.; ARANHA, J. M. R. Ontogenetic, spatial and temporal variations in the feeding ecology of *Deuterodon langei* Travassos, 1957 (Teleostei: Characidae) in a Neotropical stream from the Atlantic rainforest, southern Brazil. **Neotropical Ichthyology**, v. 6, n. 2, p. 211–222, 2008.

VITULE, J. R. S.; FREIRE, C. A.; SIMBERLOFF, D. Introduction of non-native freshwater fish can certainly be bad. **Fish and Fisheries**, v. 10, p. 98–108, 2009.

VITULE, J. R. S.; FREIRE, C. A.; VAZQUEZ, D. P.; NUÑEZ, M. A.; SIMBERLOFF, D. Revisiting the Potential Conservation Value of Non-Native Species. **Conservation Biology**, v. 26, n. 6, p. 1153–1155, 2012.

VITULE, J. R. S.; SKÓRA, F.; ABILHOA, V. Homogenization of freshwater fish faunas after the elimination of a natural barrier by a dam in Neotropics. **Diversity and Distributions**, v. 18, p. 111–120, 2012.

VITULE, J. R. S.; UMBRIA, S. C.; ARANHA, J. M. R. Registro de Ocorrência do Bagre do Canal *Ictalurus punctatus* (Rafinesque, 1818) em uma Importante Bacia Hidrográfica da Floresta Atlântica Paranaense. I Simpósio Brasileiro sobre Espécies Exóticas Invasoras. Brasília: Ministério do Meio Ambiente, 2005.

VITULE, J. R. S.; UMBRIA, S. C.; ARANHA, J. M. R. Introduction of the African catfish *Clarias gariepinus* (BURCHELL, 1822) into Southern Brazil. **Biological Invasions**, v. 8, p. 677–681, 2006.

VUCIC-PESTIC, O.; RALL, B. C.; KALINKAT, G.; BROSE, U. Allometric functional response model: Body masses constrain interaction strengths. **Journal of Animal Ecology**, v. 79, p. 249–256, 2010.

WALTHER, G. R.; ROQUES, A.; HULME, P. E.; et al. Alien species in a warmer world: Risks and opportunities. **Trends in Ecology and Evolution**, v. 24, n. 12, p. 686–693, 2009.

WELLBORN, T. L. Channel Catfish Life History and Biology. Southern Regional Aquaculture Center, **SRAC Publication**, n. 180, p. 1–4, 1988.

WELCOMME, R. L. **International introductions of inland aquatic species.** FAO Fisheries Technical Paper, 1988.

ZANATTA, A. S.; RAMOS, I. P.; DA SILVA, R. J.; LANGEANI, F.; CARVALHO, E. D. Pisces, Siluriformes, Ictaluridae, *Ictalurus punctatus* (Rafinesque, 1818): First record in middle Paranapanema river reservoir, aquaculture and exotic species dispersion. **Check List**, v. 6, n. 4, p. 589–591, 2010.

ANEXO



Ministério da Educação
UNIVERSIDADE FEDERAL DO PARANÁ
Setor de Ciências Biológicas
Comissão de Ética no Uso de Animais
(CEUA)



Nº 1027

CERTIFICADO

A Comissão de Ética no Uso de Animais do Setor de Ciências Biológicas da Universidade Federal do Paraná (CEUA/BIO – UFPR), instituída pela Resolução Nº 86/11 do Conselho de Ensino Pesquisa e Extensão (CEPE), de 22 de dezembro de 2011, **CERTIFICA** que os procedimentos utilizando animais no projeto de pesquisa abaixo especificado estão de acordo com a Diretriz Brasileira para o Cuidado e a Utilização de Animais para fins Científicos e Didáticos (DBCA) estabelecidas pelo Conselho Nacional de Controle de Experimentação Animal (CONCEA) e com as normas internacionais para a experimentação animal.

STATEMENT

The Ethics Committee for Animal Use from the Biological Sciences Section of the Federal University of Paraná (CEUA/BIO – UFPR), established by the Resolution Nº 86/11 of the Teaching Research and Extension Council (CEPE) on December 22nd 2011, **CERTIFIES** that the procedures using animals in the research project specified below are in agreement with the Brazilian Guidelines for Care and Use of Animals for Scientific and Teaching purposes established by the National Council for Control of Animal Experimentation (CONCEA) and with the international guidelines for animal experimentation.

PROCESSO/PROCESS: 23075.160195/2016-43

APROVADO/APPROVAL: 18/10/2016 – R.O. 09/2016

TÍTULO: Resposta funcional de peixes nativos do rio Guaraguaçu – PR considerando-se a estruturação de habitat causada por uma gramínea invasora

TITLE: Functional response of native and nonnative fishes of Guaraguaçu river – PR considering the habitat structuration promoted by an invasive grass

AUTORES/AUTHORS: Jean Ricardo Simões Vitule, Larissa Faria

DEPARTAMENTO/DEPARTMENT: Engenharia Ambiental


Prof. Dra. Ana Vitória Fischer da Silva
Coordenadora da CEUA