

UNIVERSIDADE FEDERAL DO PARANÁ

HUGO DE ANDRADE GONÇALVES DOS SANTOS

AVALIAÇÃO DA QUALIDADE AMBIENTAL DO RIO GUARAGUAÇU, PARANÁ
(BRASIL), UTILIZANDO BIOMARCADORES EM PEIXE NEOTROPICAL, *HOPLIAS
MALABARICUS* (BLOCH, 1794)

CURITIBA

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MALABARICUS (BLOCH, 1794)

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Orientadora: Profa. Dra. Helena Cristina da Silva de Assis

Coorientador: Prof. Dr. Andre Andrian Padial

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“AQUELE QUE AJUDA OS OUTROS SIMPLEMENTE PORQUE ISSO DEVE OU
PRECISA SER FEITO, E PORQUE É A COISA CERTA A FAZER, É SEM DÚVIDA, UM
VERDADEIRO SUPER-HERÓI.” (STAN LEE)

RESUMO

A água é um recurso limitado e essencial para a vida que sofre contaminação por diversos xenobióticos oriundos de atividades antrópicas. Dentre os principais contaminantes encontrados em corpos hídricos, pode-se destacar os agrotóxicos e os elementos traço, que são caracterizados por possuírem alta persistência ambiental e capacidade de bioacumulação em organismos aquáticos, o que podem acarretar diversos problemas a saúde. O rio Guaraguaçu encontrado no bioma da Mata Atlântica, é o principal rio do litoral paranaense por sua biodiversidade única, importância econômica e abastecimento de água para os municípios de Matinhos, Pontal do Paraná e Paranaguá. No entanto, esse rio vem sofrendo ao longo dos anos com a contaminação de suas águas devido ao crescimento urbano e o lançamento de esgoto não tratado. Além disso, sofre influência de um aterro sanitário localizado próximo a sua região intermediária em um de seus afluentes, o rio Pery. Deste modo, o objetivo deste estudo foi avaliar a influência da ação antrópica na qualidade da água do Rio Guaraguaçu a partir das análises químicas da água e de biomarcadores em peixes da espécie *Hoplias malabaricus*. Os peixes foram coletados em três setores com gradientes ecológicos distintos ao longo do rio, sendo: S1- prístino; S2- impactado; S3- menos impactado. Amostras de água foram coletadas para análise da presença de elementos traço e agrotóxicos. Nos peixes, foram analisados biomarcadores bioquímicos, histopatológicos e de genotoxicidade. Músculo de peixe foi coletado para análise de bioacumulação de elementos traço. A partir da avaliação da qualidade ambiental do rio Guaraguaçu, os resultados encontrados mostraram que a atividade da acetilcolinesterase cerebral, diminuiu no setor 2 comparado aos setores 1 e 3, indicando a presença de compostos anticolinesterásicos na água. Brânquia e fígado foram os tecidos que mais apresentaram alterações, principalmente no setor 2, com aumento da atividade do sistema antioxidante. A lipoperoxidação aumentou tanto no setor 2 como no 3, caracterizando um dano celular. Biomarcadores histopatológicos mostraram diferentes lesões no fígado e brânquias dos peixes do setor 2. Para os biomarcadores de genotoxicidade, a presença de micronúcleo ocorreu em pelo menos um organismo de cada setor, indicando presença de substâncias mutagênicas na água. O presente trabalho buscou evidenciar a problemática atual da qualidade da água do Rio Guaraguaçu para criar um banco de dados de suma importância para o conhecimento tanto dos residentes com grau de vulnerabilidade social que necessitam do rio para sobrevivência e subsistência quanto para futuras tomadas de decisões por parte dos governantes.

Palavras-chaves: Ecotoxicologia; Sistema antioxidante; Modelo biológico; Genotoxicidade.

ABSTRACT

Water is a limited and essential resource for life that suffers contamination from various xenobiotics originating from anthropogenic activities. Among the main contaminants found in water bodies, pesticides and trace elements can be highlighted, which are characterized by their high environmental persistence and bioaccumulation capacity in aquatic organisms, leading to various health problems. The Guaraguaçu River, located in the Atlantic Forest biome, is the main river in the coastal region of Paraná, Brazil, due to its unique biodiversity, economic importance, and water supply to the municipalities of Matinhos, Pontal do Paraná, and Paranaguá. However, this river has been experiencing water contamination over the years due to urban growth and the discharge of untreated sewage. Additionally, it is influenced by a landfill located near its intermediate region in one of its tributaries, the Pery River. Therefore, the objective of this study was to evaluate the influence of anthropogenic activities on the water quality of the Guaraguaçu River through chemical analysis of water and biomarkers in *Hoplias malabaricus* fish. The fish were collected in three sectors with distinct ecological gradients along the river: Sector 1 - pristine; Sector 2 - impacted; Sector 3 - less impacted. Water samples were collected for analysis of trace elements and pesticides. Biochemical, histopathological, and genotoxicity biomarkers were analyzed in the fish. Fish muscle samples were collected for analysis of trace element bioaccumulation. The results of the Guaraguaçu River environmental quality assessment showed that the activity of cerebral acetylcholinesterase decreased in Sector 2 compared to Sectors 1 and 3, indicating the presence of anticholinesterase compounds in the water. Gill and liver tissues showed the most alterations, especially in Sector 2, with an increase in the activity of antioxidant system. Lipoperoxidation increased in both Sector 2 and 3, indicating cellular damage. Histopathological biomarkers revealed different lesions in the liver and gills of fish from Sector 2. For genotoxicity biomarkers, the presence of micronuclei occurred in at least one organism from each sector, indicating the presence of mutagenic substances in the water. The present study aimed to highlight the current issues regarding the water quality of the Guaraguaçu River, to create a highly important database for the knowledge of both residents with social vulnerability who rely on the river for survival and sustenance, as well as for future decision-making by policymakers.

Keywords: Ecotoxicology; Antioxidant system; Biological model; Genotoxicity

LISTA DE ABREVIATURAS E SIGLAS

ACh - Acetilcolina

AChE - Acetilcolinesterase

Al – Alumínio

ANOVA – Análise de Variância

APA – Área de Proteção Ambiental

As – Arsênio

ATP – Adenosina Trifosfato

BL – Blebbed

BN – Binucleus

CAT - Catalase

Cd - Cádmio

CDNB – 1-Cloro-2,4-Dinitrobenzeno

Cfa - Clima subtropical, com verão quente

CONAMA - Conselho Nacional do Meio Ambiente

Cr - Cromo

CTAF – Centro de Tecnologias Avançadas em Fluorescência UFPR

Cu - Cobre

CYP450 - Citocromo P450

DNA - Ácido Desoxirribonucleico

EEG – Estação Ecológica do Guaraguaçu

EC – Commission Regulation of the European Community (Regulamento da Comissão da Comunidade Europeia)

EROS - Espécies Reativas de Oxigênio

FAO – Food and Agriculture Organization (Organização das Nações Unidas para Alimentação e Agricultura)

Fe – Ferro

FOX – Ferrous Oxidation in Xylenol orange (Oxidação Ferrosa do laranja de Xilenol)

GPx – Glutathione Peroxidase

GR - Glutathione Redutase

GSH – Glutathione Reduzida

GSSG - Glutathione oxidada ou Glutathione dissulfeto

GST - Glutathione-S-Transferase

HCl – Ácido Clorídrico

H₂O - Água

H₂O₂ - Peróxido de Hidrogênio

Hg – Mercúrio

HNO₃ – Ácido Nítrico

IAP – Instituto Ambiental do Paraná

ICMBio – Instituto Chico Mendes de Conservação da Biodiversidade

ICP-OES – Espectrometria de Emissão Óptica com Plasma Indutivamente Acoplado

KNO₃ – Nitrato de Potássio

LAA – Laboratório de Análises Ambientais

LAMAQ – Laboratório Multiusuário de Análises Químicas

LASB – Laboratório de Análise e Síntese em Biodiversidade

LB – Lobed

LEC – Laboratório de Ecologia e Conservação

LPO - Lipoperoxidação

LQ – Limite de Quantificação

LTA – Laboratório de Toxicologia Ambiental

MC – Células Mucosas

MET – Metalotioneína

MG – Minas Gerais

Mn - Manganês

N – Focos Necróticos

NaBH₄ – Borohidreto de Sódio

NADPH - Nicotinamida Adenina Dinucleotídeo Fosfato

NaOH – Hidróxido de Sódio

ND – Não Detectado

Ni – Níquel

NT – Notched

O₂ – Oxigênio

O²⁻ - Radical Superóxido

OH⁻ - Radical Hidroxila

PAHs - Polycyclic Aromatic Hydrocarbons (Hidrocarbonetos Policíclicos Aromáticos -HPAs)

Pb - Chumbo

PCA1 – Eixo x

PCA2 – Eixo y

PCoA – Análises de Coordenadas Principais

PCoA1 – Eixo 1 (x)

PCoA2 – Eixo 2 (y)

PERMANOVA – Análise Multivariada de Permutação de Variância

pH – Potencial Hidrogeniônico

PL – Lamela Primária

PNRH - Política Nacional de Recursos Hídricos

POPs – Poluentes Orgânicos Persistentes

RDC – Resolução da Diretoria Colegiada

S1 – Setor ou Seção 1

S2 – Setor ou Seção 2

S3 – Setor ou Seção 3

SEM – Scanning Electron Microscope (Microscópio Eletrônico de Varredura -MEV)

SINGREH - Sistema Nacional de Gerenciamento de Recursos Hídricos

SisBio – Sistema de Autorização e Informação em Biodiversidade

SL – Lamela Secundária

SOD - Superóxido Dismutase

TECLAB – Tecnologia em Análises Laboratoriais

UFPR – Universidade Federal do Paraná

U.S. EPA – United States Environmental Protection Agency (Agência de Proteção Ambiental dos Estados Unidos)

U.S. Geological Survey - United States Geological Survey (Serviço Geológico dos Estados Unidos)

UTFPR – Universidade Tecnológica Federal do Paraná

V – Vaso Sanguíneo

VC – Vacuolated

Zn - Zinco

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1 APRESENTAÇÃO DA DISSERTAÇÃO

O presente trabalho contempla o trabalho de pesquisa do discente Hugo de Andrade Gonçalves dos Santos do Programa de Pós-Graduação em Ecologia e Conservação da Universidade Federal do Paraná (UFPR). Inicialmente, apresenta-se uma introdução geral afim de ingressar no assunto do trabalho e construir o cenário em que se encontra a presente pesquisa, as hipóteses a serem testadas, seguindo pelos objetivos geral e específicos que o nortearam. O presente trabalho apresentou resultados significativos que irá gerar uma publicação científica no formato de artigo, que será enviado para a revista Science of The Total Environment (STOTEN).

Esta dissertação está apresentada no formato de artigo, seguindo o modelo em língua inglesa, apresentando resumo, introdução, metodologia, resultados e discussão, além da conclusão. O trabalho intitulado como: “Biomonitoring Top Fish Predators using Biomarkers Unveil Human Impacts in a Coastal River from a World Heritage Site”, contempla a análise de biomarcadores bioquímicos, histopatológicos e de genotoxicidade em traíras (*H. malabaricus*), além da avaliação dos parâmetros físico-químicos da água, e análises químicas de agrotóxicos e elementos traço em água e em músculo de peixe, para se avaliar a qualidade da água do principal rio da costa paranaense, o Rio Guaraguaçu.

Por fim, apresenta-se as considerações finais abordando os resultados do trabalho de forma simplificada e unificada, com algumas recomendações para futuros projetos e trabalhos.

2. INTRODUÇÃO GERAL

Do total da superfície terrestre mais de 70% é recoberta por água, sendo aproximadamente 97% de água salgada, encontrada em oceanos e mares (Esteves, 1998). Dentre as reservas de água doce, mais de 2% são encontrados em geleiras, estabelecendo uma porcentagem menor que 1% de água disponível para consumo em rios, lagos e aquíferos (Esteves, 2011; Häder et al, 2020), demonstrando a necessidade de se cuidar de um recurso tão limitado. Do total de água doce disponível no mundo, aproximadamente 12% se encontram em território brasileiro (Silva; Pompêo; Paiva, 2015; Elste et al., 2019). A importância da água, pode ser ainda evidenciada quando 60% do corpo humano adulto é composto por esse recurso e em alguns organismos essa composição pode alcançar 90%, sendo essencial para todos os organismos vivos (U.S. Geological Survey, 2019).

Logo, o desenvolvimento da população humana junto do crescimento exacerbado da população mundial, da expansão urbana e do modelo consumista/capitalista de viver, estão ligados diretamente ao uso da água de forma direta ou indireta como recurso essencial para diversos processos, entre as quais se destaca o abastecimento, geração de energia, irrigação na agricultura, produção, navegação e aquicultura (Moraes; Jordão, 2002; Tundisi; Tundisi, 2011). Contudo, desde o início do século XX, esse recurso tem enfrentado grande pressão e competição devido a conflitos de interesses entre os seres humanos, resultando em uma redução em sua qualidade para consumo e ameaçando a existência de organismos nos ecossistemas aquáticos. Isso é principalmente causado pelo lançamento de esgoto, contaminação por agrotóxicos e elementos traço, superexploração, invasão de espécies exóticas, mudanças climáticas, regulação de fluxo (como a construção de reservatórios) e mudanças no uso da terra, que levam à erosão do solo e ao assoreamento dos corpos hídricos (Silva; Pompêo; Paiva, 2015; Dudgeon, 2019).

Dentre os principais contaminantes dos corpos hídricos, pode-se destacar os agrotóxicos e elementos traço. A degradação da água por agrotóxicos é um dos principais causadores da contaminação dos recursos hídricos, junto com a contaminação por esgotos domésticos, já que são encontrados em águas superficiais e subterrâneas do mundo todo, em função do amplo uso em áreas agrícolas e urbanas (Armas et al., 2007; Klaassen; Watkins III, 2012; Sharma et al., 2019; Rathi; Kumar; Vo, 2021). Com seus altos riscos ambientais e à saúde pública, a legislação brasileira possui a lei nº 7.802/1989 voltada sobretudo para o ciclo de vida dos agrotóxicos, desde a produção até o descarte ecologicamente correto dos resíduos, além do registro, classificação, controle, inspeção e fiscalização desses produtos (Brasil,

1989). A preocupação com os agrotóxicos se deve a variedade de classes e composições que podem possuir. Os agrotóxicos podem variar de acordo com a espécie alvo a ser exterminada, desde fungicidas, rodencidas, inseticidas até herbicidas. No entanto, muitas vezes não só as espécies alvo são afetadas assim como todo o meio ambiente circundante. A composição dessas substâncias também pode variar bastante desde organoclorados, organofosforados, carbamatos e piretróides, tornando-se uma das principais causas de contaminação ambiental (Sharma et al., 2019; Souza et al., 2020).

A concentração exacerbada dos elementos traço no meio ambiente, está relacionado principalmente a atividades do ser humano, como no uso em tintas anti-incrustantes utilizadas em embarcações, despejo de resíduos pelas indústrias, refinarias de petróleo, processos de clareamento de metais, fabricação de plásticos e carvão ativado, encontrados em agrotóxicos, na queima de combustíveis, mineração, descarte incorreto de resíduos sólidos, utilizados em baterias e até em aditivos alimentares como é o caso do manganês (Pereira, 2004; Esteves, 2011). Considerados na sua maioria elementos com grande poder carcinogênico, mutagênico, teratogênico, nefrotóxico, imunotóxico, neurotóxico, com capacidade de desestruturar proteínas enzimáticas (Al-Sabti et al., 1994; Moraes; Jordão, 2002; Klaassen; Liu; Diwan, 2009; Häder et al., 2020; Nordberg; Nordberg, 2022).

Por isso, o Brasil possui em sua legislação normas, resoluções, regulamentações e leis voltadas para avaliação da qualidade da água, controle dos efluentes liberados nos corpos hídricos, controle das atividades potencialmente poluidoras e criação de instituições voltadas para segurança hídrica, que mesmo sendo deficientes e necessitem de aperfeiçoamentos, são capazes de determinar os níveis de poluição aceitáveis de determinados sistemas, onde o uso de suas águas são indispensáveis para atividades humanas (Pereira, 2004; Häder et al., 2020). A resolução CONAMA nº 357/2005 tem por finalidade classificar os corpos de água e instaurar diretrizes ambientais para o seu enquadramento, como também estabelecer condições e padrões de lançamento de efluentes. O artigo 2º desta resolução, nos parágrafos XXI e XXII, estabelece que ensaios ecotoxicológicos são determinantes para análise do efeito deletério de agentes físicos ou químicos a diversos organismos aquáticos com intuito de avaliar o potencial de risco à saúde humana (CONAMA, 2005).

Para complementar e alterar a resolução 357/05 foi criada a resolução CONAMA 430/2011, que dispõe sobre as condições, parâmetros, padrões e diretrizes para controle do lançamento de efluentes em corpos hídricos (CONAMA, 2011). Além das resoluções CONAMA, em 1997 foi criada a lei nº 9433 que institui a Política Nacional de Recursos Hídricos (PNRH), junto da criação do Sistema Nacional de Gerenciamento de Recursos

Hídricos (SINGREH). No artigo 1º dessa lei nos parágrafos I e II, a importância e a preocupação com a água foram evidenciadas, ao caracterizar esse recurso como um bem de domínio público, limitado e dotado de valor econômico (Brasil, 1997).

No entanto, no Brasil a legislação voltada para a ecotoxicidade de lançamento de fontes poluidoras é pouco exigente (Magalhães; Ferrão Filho, 2008), mesmo tendo estudos na área de ecotoxicologia que já avaliam os efeitos que determinados contaminantes podem causar em organismos vivos em diferentes níveis de organização (Castro et al., 2014; Cavalcanti et al., 2016; Dalzochio et al., 2016; Pereira et al., 2020). Com a falta de legislações mais abrangentes, diversos rios brasileiros importantes para abastecimento, lazer e subsistência correm o risco de degradação, como o rio Guaraguaçu, importante rio da planície litorânea do Paraná devido a sua rica biodiversidade e seus serviços ambientais (Contente; Stefanoni; Spach, 2011; Elste et al. 2019; Cavallini; Reis; Tiepolo, 2020).

2.1. RIO GUARAGUAÇU

O litoral do Paraná (Brasil) que ocupa uma área de cerca de 6.058 km², é caracterizado pela presença do bioma da Mata Atlântica considerado *hotspot* da biodiversidade. As características morfológicas e de relevo da região é determinada pela presença da Serra do Mar e a planície litorânea (Torres, 2019), logo a presença de chuvas orográficas na região litorânea é comum (IAT, 2006). O clima da região é subtropical úmido com uma precipitação média anual de 2.500 mm com um padrão claro de chuva sazonal (Lana et al. 2001; Abreu-Mota et al., 2014), invernos secos com precipitações de até 60 mm e verão chuvoso podendo ultrapassar 1000 mm de precipitação (Silva, 2008; Vitule, 2008).

No litoral paranaense existem duas bacias hidrográficas, a bacia da baía de Guaratuba e da baía de Paranaguá, que são subdivididas em várias sub-bacias e fazem parte da bacia do Atlântico Sul (Staszczak; Rocha, 2018). Um dos principais rios tributários da bacia hidrográfica da baía de Paranaguá encontrado em sua região sul, é o rio Guaraguaçu (25°45'W e 48°35'S") que possui padrão meandrante característico de rios de baixa energia, percorrendo a planície litorânea e envolto por floresta ombrófila densa de terras baixas, abrangendo assim os municípios do Pontal de Paraná, Paranaguá e Matinhos (IAT, 2006; Cavallini, 2018; Elste et al., 2019; Torres, 2019; Cavallini; Reis; Tiepolo, 2020). Caracteriza-se por ser um rio com grande influência também de marés, tendo seu vazamento em direção ao mar durante a maré baixa, com um certo período de estagnação e sofrendo posteriormente refluxo durante a enchente da maré (IAT, 2006).

O Rio Guaraguaçu possui importância ambiental, além de possuir relevância para os municípios do litoral do Paraná, provendo o abastecimento de água para os municípios de Pontal do Paraná, Matinhos e Paranaguá (Silva, 2008; Elste et al, 2019), assim como uma área utilizada para prática de atividades e lazer, como a pesca que pode ser esportiva ou de subsistência para os moradores da região (Reis et al., 2015). Entretanto, o aumento das contaminações e degradações ao longo dos trechos do rio Guaraguaçu ameaçam os usos múltiplos desse rio (Fig. 1).



Fig. 1. Possíveis fontes de contaminação dos corpos hídricos, como por exemplo o Rio Guaraguaçu.

Nessa região, há pouca proteção das margens e há presença de atividades de agricultura. Mesmo sendo considerada em sua maioria de origem familiar/subsistência, possuindo como principais cultivos a banana, mandioca, o feijão, o milho e o arroz (ZEE, 2016), existe a possibilidade da utilização de agrotóxicos causando riscos de contaminantes nos afluentes do rio Guaraguaçu, e do maior escoamento de contaminantes por águas pluviais. Além disso, a presença da mineradora Nova Prata na rodovia Alexandra Matinhos (PR-508) voltada para materiais relacionados a construção civil, como areia, e da presença constante de embarcações no rio e tráfego de carros pela rodovia PR-407, pode aumentar a contaminação por combustíveis, óleos, graxas e outros fluidos (IAT, 2006; Reis et al., 2015; ZEE, 2016; Cavallini, 2018; Araújo; Vitule; Padial, 2021).

No entanto, a contaminação ocorre principalmente devido ao lançamento de efluentes doméstico e o escoamento de contaminantes para um de seus principais afluentes

(cujo canal foi retificado para escoamento), o rio Pery, a partir de um aterro sanitário encontrado no município do Pontal do Paraná (Singo; Araújo-Ramos; Rocha, 2020). Tal fenômeno é principalmente observado após o verão, quando a região recebe milhares de turistas, sobrecarregando os sistemas de drenagem e tratamento de esgoto, caracterizando a contaminação ao longo do rio como heterogênea (Elste et al, 2019). O gradiente de impacto antrópico do rio Guaraguaçu, desde sua nascente mais preservada, passando por uma área a jusante de maior ação antrópica onde se encontra o rio Pery, até sua foz caracterizada pela transição para um ambiente estuarino, assim como sua variação sazonal, já foi descrito anteriormente e tem relação com a diversidade de plantas aquáticas, ocorrência de espécies invasoras e impactos ecológicos como homogeneização biótica (Araújo; Vitule; Padial, 2021; Sato; Costa; Padial, 2021; Galvanese et al, 2022). Logo, a utilização do biomonitoramento se torna uma ferramenta essencial ao avaliar a qualidade ambiental através da análise dos possíveis efeitos de contaminantes na saúde dos organismos, especificamente peixes ao se tratar do presente trabalho.

2.2. AVALIAÇÃO DE QUALIDADE AMBIENTAL/MODELO BIOLÓGICO

A escolha do modelo biológico na avaliação da qualidade ambiental depende de inúmeras características inerentes aos organismos. Dentre os principais modelos biológicos utilizados, destacam-se: as microalgas, crustáceos, peixes, invertebrados bentônicos, bivalves, poliquetas (López-Doval; Barata; Díez, 2015) e até girinos de determinadas espécies de sapo (Fernandes et al., 2021). Os organismos devem seguir pelo menos alguns dos requisitos seguintes para serem utilizados no biomonitoramento: abundantes e com ampla distribuição; possuírem relevante representação ecológica; conhecimento prévio de sua biologia, fisiologia e hábitos alimentares; possuir importância comercial; fácil cultivo e serem nativos (Magalhães; Ferrão Filho, 2008). Apesar de todas as possibilidades possíveis para uso no monitoramento ambiental, a escolha do organismo será dependente da área de estudo e dos objetivos do trabalho a ser realizado (Resh, 2008).

Os peixes são organismos essenciais para os ambientes aquáticos circundantes para avaliar a condição e funcionamento desses ecossistemas, entretanto, a presença de xenobióticos na água pode causar efeitos adversos no metabolismo desses organismos (Burkina; Zlabek; Zamaratskaia, 2015; Ballesteros et al., 2017). No Brasil, algumas espécies de peixes utilizadas em trabalhos para biomonitoramento, são o *Geophagus brasilienses*

(Oliveira et al., 2019), *Hoplias malabaricus* (Pantaleão et al., 2006) e *Rhamdia quelen* (Souza-Bastos et al., 2017).

O *H. malabaricus* (Bloch, 1794) (Fig. 2), popularmente conhecido como traíra, é caracterizado como predador de topo de cadeia, sendo comumente utilizado em trabalhos para análise de biomagnificação de certos contaminantes de rios e reservatórios por sua capacidade de indicar respostas de efeitos crônicos e de bioacumulação (Castro et al., 2014; Mela et al., 2014). A espécie é carnívora com uma estratégia de predação por emboscada (“senta e espera”) não sendo considerada uma boa nadadora (Chu-Koo; Pérez, 2007), com a sua fase adulta principalmente relacionada na maioria das vezes a ambientes limnéticos (lênticos), em águas rasas e perto de vegetações marginais ou submersas (Carvalho; Fernandes; Moreira, 2002; Reis et al., 2017; Paula; Risso; Martinez, 2021; Leite et al., 2021).



Fig. 2. *Hoplias malabaricus*.

Apresenta boca com dentes adaptados para segurar e engolir presas inteiras, possuindo hábitos alimentares generalistas que são dependentes de sua fase de vida, considerada estritamente insetívora durante sua fase juvenil e na fase adulta preferencialmente piscívora (Moraes; Barbola, 1995; Silva, 2008; Pessoa et al., 2013; Montenegro et al., 2013). Conserva grande amplitude ecológica ao mostrar resistência a diferentes perturbações, possuindo uma das maiores tolerâncias à privação alimentar registradas ao conseguir manter suas taxas metabólicas mesmo após um longo período de jejum, aguentam altas temperaturas da água, sobrevivem em ambientes com baixos níveis de oxigênio dissolvido (Barbieri, 1989; Costa et al., 2007; Cruz-Esquivel; Marrugo-Negrete, 2022), além de conseguirem viver em ambientes aquáticos com pH ácido, como encontrado em riachos no Peru (Chu-Koo; Pérez, 2007).

O ciclo reprodutivo da traíra ocorre tanto em ambientes de água limpa quanto em ambientes degradados e acontece quase que exclusivamente nos períodos de primavera (setembro-outubro) e verão, apesar de possuir um desenvolvimento gonadal assíncrono caracterizado pelo parcelamento da desova, considerado como uma adaptação da espécie para evitar a competição pela busca de alimento e local de desova (Barbieri, 1989; Gomes et al., 2015; Melo et al., 2017). O tamanho médio para maturação sexual se encontra por volta dos 16 cm de comprimento (Moraes; Barbola, 1995), entretanto sabe-se que a partir dos 23 cm 100 % das fêmeas já se tornam aptas à reprodução (Barbieri, 1989), conhecido por ser um peixe capaz de ultrapassar os 40 cm de comprimento (Balboni; Colautti; Baigún, 2011) chegando até no máximo 65 cm (Chu-Koo; Pérez, 2007) e pesar mais de 1 kg (Carvalho et al., 2022). Destaca-se que apesar de ser uma espécie com hábito alimentar voraz (Monteiro; Rantin; Kalinin, 2013), possui comportamento de cuidado parental com suas crias (Gomes et al., 2015; Melo et al., 2017).

Os estudos toxicológicos com traíra no geral, possuem como objetivo principal analisar os impactos de contaminantes aos ambientes aquáticos in situ (Lozano et al., 2013; Carvalho; Fernandes; Moreira, 2002), ou são controladas em laboratório para análise das respostas a determinadas concentrações de contaminantes já que a mesma possui habilidade de se adaptar a condições experimentais (Costa et al., 2007; Silva de Assis et al., 2013; Paula; Risso; Martinez, 2021). A biologia reprodutiva, comportamental, morfológica e fisiológica de *H. malabaricus* também já foi foco de diversos estudos a fim de detalhar tal espécie (Barbieri, 1989; Domanico; Delfino; Freyre, 1993; Marques; Gurgel; Lucena, 2001; Carvalho; Fernandes; Moreira, 2002; Chu-Koo; Pérez, 2007; Balboni; Colautti; Baigún, 2011; Lima et al., 2017), assim como já foi evidenciado a relevância da espécie para a alimentação humana (Castro et al., 2014).

Logo, possuindo a maioria das características para ser utilizado como modelo biológico a traíra se tornou uma ótima espécie para avaliação da qualidade ambiental de corpos hídricos. No Rio Guaraguaçu uma das espécies nativas de peixe com ampla distribuição durante toda sua extensão é o *H. malabaricus* (Silva, 2008; Occhi, 2020; Carvalho et al., 2022), mostrando a possibilidade e relevância de analisar biomarcadores nessa espécie.

2.3. BIOMARCADORES DE CONTAMINAÇÃO AMBIENTAL

O uso de biomarcadores pode ser instrumento eficaz em várias etapas do biomonitoramento da qualidade ambiental de ecossistemas aquáticos (Van der Oost; Beyer; Vermeulen, 2003). Existem diversos biomarcadores que se destacam em bioensaios e refletem a saúde dos organismos e podem indicar a presença, o efeito e em alguns casos até o grau de contaminação do ambiente, como: os biomarcadores bioquímicos, de genotoxicidade, hematológicos, fisiológicos e histopatológicos. Dentre os biomarcadores mais utilizados no monitoramento ambiental, estão os biomarcadores bioquímicos. O uso de biomarcadores bioquímicos podem corroborar para detecção de alterações ambientais de forma precoce. Esses biomarcadores permitem a identificação de efeitos no organismo em níveis mais basais de organização (celular, bioquímico, molecular), reforçando a compreensão de repostas mais rápidas, antes que ocorra a morte do indivíduo (Van der Oost; Beyer; Vermeulen, 2003; Friberg et al., 2011; López-Doval; Barata; Díez, 2015).

Tratando-se de biomarcadores bioquímicos para avaliação de neurotoxicidade, destaca-se a análise da atividade enzimática da acetilcolinesterase (AChE). Os organofosforados e carbamatos são relatados como inibidores eficazes da AChE (Cavalcanti et al., 2016), se ligando ao sítio ativo da enzima e inibindo sua ação. Logo, a função de hidrolisar o neurotransmissor acetilcolina (ACh) em colina e ácido acético é interrompida, fazendo que ocorra acumulação da acetilcolina na fenda sináptica, atuando assim sobre o sistema nervoso parassimpático, ocasionando a hiperexcitação colinérgica, danos irreversíveis e morte do indivíduo (Van der Oost; Beyer; Vermeulen, 2003; Cavalcanti et al., 2016). A diminuição da atividade da acetilcolinesterase pode afetar a coordenação motora dos organismos (Burkina; Zlabek; Zamaratskaia, 2015), podendo interferir em comportamentos, locomoção e na fisiologia dos organismos.

Enzimas dos sistemas de biotransformação podem corroborar para a desintoxicação/transformação de xenobióticos e seus metabólitos, e o sistema antioxidante pode atuar no combate de radicais livres para evitar que ocorram danos oxidativos nos diferentes tecidos analisados. Como a metabolização de xenobióticos no organismo ocorre principalmente no fígado, é um dos órgãos mais utilizados para análises destes biomarcadores de biotransformação.

O sistema de biotransformação nos organismos, como nos peixes, ocorre em duas fases. As reações da fase I estão relacionadas com a adição de um grupo funcional polar (-OH, -NH₂, -SH ou -COOH) em compostos lipofílicos, incluindo reações de oxidação,

redução e hidrólises, caracterizando a fase inicial de desintoxicação e excreção (Kroon; Streten; Harries, 2017; Klaassen; Watkins III, 2012). Os principais grupos de enzimas envolvidas nas reações da fase de biotransformação são as pertencentes ao grupo Citocromo P450 (CYP450). Na fase II da biotransformação ocorre a conjugação do xenobiótico ou seus metabólitos com um ligante endógeno. A glutathione-S-transferase (GST), enzima da fase II, catalisa a conjugação de metabólitos da fase I com a forma reduzida de glutathione (GSH), facilitando assim a excreção de produtos químicos ao adicionar mais grupos polares tornando-os mais hidrofílicos (Van der Oost; Beyer; Vermeulen, 2003; Burkina; Zlabek; Zamaratskaia, 2015; Kroon; Streten; Harries, 2017) intensificando a excreção de xenobióticos.

Assim como o sistema de biotransformação, o sistema antioxidante também é muito importante, pois ajuda para a redução de estresses oxidativos ocasionados por contaminantes. O estresse oxidativo caracteriza-se por ser um desequilíbrio entre a geração de compostos oxidantes, como o radical superóxido (O_2^-), peróxido de hidrogênio (H_2O_2) e o radical hidroxila (OH^-) que são espécies reativas de oxigênio (EROS) e a atuação do sistema antioxidante, o que favorece a ocorrência de lesões oxidativas em macromoléculas e estruturas celulares, podendo resultar em morte celular (Winston; Di Giulio, 1991; Gutteridge, 1993; Barbosa et al., 2010). Com a presença de contaminantes no ambiente, os organismos aumentam a produção de espécies reativas de oxigênio, sendo a mitocôndria uma das principais fontes endógenas geradoras de EROS (Rover Júnior et al., 2001).

Desta forma, enzimas como a superóxido dismutase (SOD), catalase (CAT), encontrada em todas as células aeróbicas e em altos níveis no fígado e rim dos organismos, e glutathione peroxidase (GPx) são ativadas e ajudam na defesa antioxidante (Fig. 3) no organismo (Barbosa et al., 2010; Fernandes et al., 2021). A SOD, uma metaloenzima que pode ser encontrada no citosol, sendo dependente de cobre (Cu^{2+}) e zinco (Zn^{2+}) (SOD-Cu/Zn), e na mitocôndria necessitando do manganês como cofator (SOD-Mn) (Meister; Anderson, 1983; Yadav; Trivedi, 2009; Barbosa et al., 2010), catalisa o radical superóxido em peróxido de hidrogênio (McCords; Fridovich, 1969) que posteriormente será degradado pela CAT em água (H_2O) e oxigênio (Pastorino et al., 2021). A GPx pode atuar na catalização de peróxido de hidrogênio, corroborando com a atividade da CAT, além de contribuir na redução de outros peróxidos em seus álcoois correspondentes (Mills, 1957; Mannervick, 1985). A GPx durante o processo de catalização de peróxidos, emprega a glutathione reduzida (GSH) como cofator, gerando glutathione oxidada ou glutathione dissulfeto (GSSG) como produto, sendo necessário a ação da glutathione redutase (GR) para a regeneração de GSH, a partir de GSSG, na presença de nicotinamida adenina dinucleotídeo fosfato (NADPH) (Meister;

Anderson, 1983; Huber; Almeida, 2008). Logo, tais enzimas, são importantes para evitar que danos oxidativos ocorram, sendo especialmente importante na proteção da membrana celular contra a lipoperoxidação (LPO) (Dalzochio et al., 2016; Kroon; Streten; Harries, 2017).

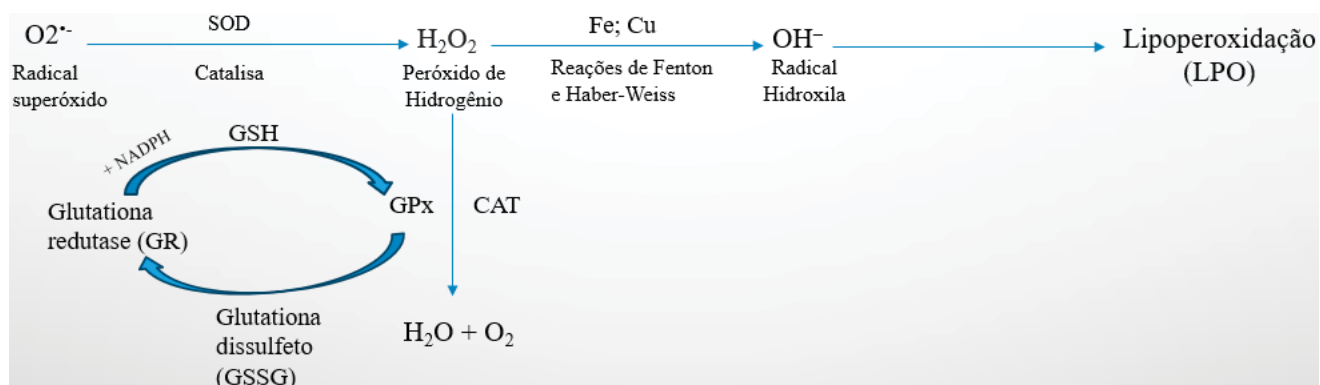


Fig. 3. Sistema antioxidante. Caso a GPx e CAT não catalisem o peróxido de hidrogênio, a partir da presença de ferro e cobre, pode ocorrer Reações de Fenton e Haber-Weiss, produzindo radical hidroxila, o qual, não possui enzima conhecida capaz de catalisá-lo, logo ocorre lipoperoxidação devido ao acúmulo de radical hidroxila.

Além do uso de atividades enzimáticas como biomarcadores bioquímicos, também é comumente utilizada a metalotioneína (MET) como biomarcador, principalmente para controle da concentração de elementos traço no organismo da maioria dos grupos de animais (El-Khayat et al., 2020). A metalotioneína é uma proteína não enzimática que possui baixo peso molecular, alto teor de cisteína e ausente de aminoácidos aromáticos, sintetizada primariamente no fígado e rins, sua produção é dependente da disponibilidade de minerais como zinco e selênio no organismo (Thirumoorthy et al., 2011). Por apresentar grupos tiol ($-SH$) dos resíduos de cisteína permitindo que a MET se ligue a metais pesados específicos, essa proteína possui propriedades de desintoxicação de elementos traço não essenciais como mercúrio e cádmio, além de trabalhar na homeostase de elementos essenciais como cobre e manganês. Ademais, a metalotioneína atua como antioxidante contra espécies reativas de oxigênio, protegendo contra danos no DNA e estresse oxidativo (Amiard et al., 2006; Atli; Canli, 2008; Fabrin et al., 2018; Nordberg; Nordberg, 2022).

Outros biomarcadores são comumente utilizados para complementar as respostas adquiridas com a avaliação dos biomarcadores bioquímicos e assim ilustrar os inúmeros danos causados pelos xenobióticos em diferentes níveis de organização no organismo, como os biomarcadores de genotoxicidade, através da análise de Micronúcleo Píscero por exemplo, e os biomarcadores histopatológicos.

Dentre os órgãos mais utilizados em análises histopatológicas, assim como na análise bioquímica, pode-se destacar o fígado, principal órgão para biotransformação de

xenobióticos, e a brânquia. A análise de brânquia em peixes é amplamente utilizada por se tratar de um órgão com várias funções, incluindo troca gasosa, regulação iônica e excreção de metabólitos (Hedayati, 2018).

Biomarcadores de genotoxicidade são essenciais para verificação de danos/modificações no DNA, do potencial carcinogênico de contaminantes e assim acrescentar informações para alcançar um melhor direcionamento dos estressores existentes em determinados ambientes aquáticos (Palhares; Grisolia, 2002; Yadav; Trivedi, 2009). Dentre os testes e métodos para avaliação genotóxica em organismos, pode-se destacar o ensaio de Micronúcleo Písceo, bastante utilizado com peixes para avaliação na ocorrência de mutagênese e conseqüentemente o surgimento de micronúcleos nas células desses organismos (Al-Sabti; Metcalfe, 1995; Obiakor; Okonkwo; Ezeonyejiaku, 2012; Oliveira; Valdes, 2019).

Os micronúcleos presentes nos eritrócitos dos peixes, são corpos contendo cromatina citoplasmática formados quando fragmentos cromossômicos sem centrômero ou cromossomos se retardam durante a anáfase e não se incorporam aos núcleos das células filhas durante a divisão celular (Al-Sabti; Metcalfe, 1995; Pantaleão et al., 2006). Os micronúcleos são identificados facilmente como resultado de atividade de quebra cromossômica (clastogênica) pelos contaminantes (Palhares; Grisolia, 2002; Ali; El-Shehawi; Seehy, 2008; Martins; Paz; Brentano, 2010).

Logo, utilizar inúmeros biomarcadores com diferentes objetivos para análise, gera uma ampla perspectiva sobre a qualidade ambiental ao fornecer dados baseados nos efeitos integrados de variados estressores ambientais na saúde dos organismos, populações, comunidades e ecossistema como um todo, tornando essa estratégia amplamente utilizada em pesquisas avançadas e planos de gestão ambiental eficazes (Georgieva et al., 2021). Essa prática de uso de multibiomarcadores, é considerada vantajosa por permitir ações corretivas em áreas impactadas, por permitir ações preventivas para a conservação ambiental e criar informações capazes de alertar toda a população humana que pode estar sendo afetada por diversos contaminantes naquele ambiente (Burkina; Zlabek; Zamaratskaia, 2015; Kroon; Streten; Harries, 2017; Salgado et al., 2021).

Portanto, essa dissertação busca contribuir com o melhor entendimento da qualidade da água do rio Guaraguaçu e assim criar uma base de dados capaz de fomentar a criação de novas leis e tomadas de decisões por parte dos governantes, além de evidenciar a problemática do rio para a população ribeirinha que usa o rio para subsistência.

2.4. HIPÓTESES

As hipóteses preditivas seriam que o setor que sofre com uma ação antrópica maior, sofrerá mais alterações dos biomarcadores e que o peixe *Hoplias malabaricus* seria um modelo biológico ideal para análise de forma complementar de todos os biomarcadores, conseguindo diferenciar a qualidade ambiental de cada setor.

2.5. OBJETIVOS

2.5.1. GERAL

Avaliar a influência da ação antrópica e ambiental na qualidade da água utilizando análises químicas e biomarcadores de contaminação ambiental em peixe predador de topo de cadeia trófica *H. malabaricus*.

2.5.2. ESPECÍFICOS

Avaliar os parâmetros físico-químicos da água para melhor representar e interpretar as possíveis respostas dos biomarcadores;

Quantificar a concentração de diferentes tipos de agrotóxicos na água e determinados elementos traço na água e biota, como alumínio (Al), arsênio (As), cádmio (Cd), cobre (Cu), cromo (Cr), chumbo (Pb), ferro (Fe), manganês (Mn), níquel (Ni), mercúrio (Hg) e zinco (Zn) para compreender as interferências antrópicas existentes no rio Guaraguaçu e como isso pode interferir na vida dos peixes e conseqüentemente na vida da população que depende do rio para sobrevivência e subsistência;

Evidenciar a influência do rio Pery, único afluente da margem direita, a partir da análise dos biomarcadores e dos parâmetros da água, sobre o rio Guaraguaçu para avaliar a possibilidade da formação de uma divisão de setores com diferentes gradientes ecológicos.

1 **3. Biomonitoring Top Fish Predators using Biomarkers Unveil Human Impacts in a**
2 **Coastal River from a World Heritage Site'**

3

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33 3.1. HIGHLIGHTS:

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35 ➤ Fe, Al and Mn were detected in water.

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37 ➤ Presence of blood genotoxicity in impacted and non-impacted sectors.

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39 ➤ Biochemical and histopathological biomarkers alterations in impacted sectors.

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41 ➤ Essential trace elements were detected in fish tissue.

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66 3.2. ABSTRACT

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68 The use of biomarkers in fish for biomonitoring is a valuable approach to reveal
69 effects of human impacts on biota health. Top predator fish are effective models for
70 monitoring human activities' impacts on aquatic ecosystems. The Guaraguaçu River, the
71 largest river-system on coastal region of South Brazil and a World Heritage site, is
72 ecologically and economically important for nearby municipalities with tourism potential. The
73 river receives contaminants from disorderly urban growth, including discharges of domestic
74 sewage and small fishery boats mainly, particularly during the tourist season. Our study
75 aimed to assess impact of anthropogenic activities on water quality in the Guaraguaçu River
76 by analyzing environmental contamination biomarkers in the top fish predator *Hoplias*
77 *malabaricus*. Fish were collected using a fyke net trap across sectors representing a gradient
78 of anthropic impact: sector 1 - pristine; sector 2 – impacted; and sector 3 - less impacted.
79 Water samples were collected to analyze the presence of trace elements and pesticide.
80 Biomarkers of the antioxidant system, histopathology, genotoxicity, neurotoxicity, and
81 concentration of trace elements were analyzed in fish tissues. In water samples Al, Fe and Mn
82 were detected, but no pesticides were found. In fish muscle, zinc and iron were detected.
83 Brain acetylcholinesterase activity decreased in impacted sectors, indicating neurotoxic
84 effects. The antioxidant system increased activity in gills and liver, and damage from
85 lipoperoxidation was observed, particularly in sector 2 when compared to sector 1, suggesting
86 oxidative stress. Histopathological biomarkers revealed lesions in the liver and gills of fish in
87 impacted sectors. Micronuclei, a genotoxicity biomarker, were observed in organisms from all
88 sectors. Our results demonstrate the detrimental effects of poor water quality on biota health,
89 even when contaminants are not detected in water. This information is important for the
90 social-vulnerable residents who rely on the river for survival, as well as for decisions-makers.

91

92 Keywords: Ecotoxicology; Environmental health; Biomonitor; Water contaminants;
93 Guaraguaçu River; social-vulnerable residents.

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98 3.3. INTRODUCTION

99 Human activities significantly influence water quality, disrupting the natural, and
100 causing imbalances in aquatic ecosystems (Chaudhry; Malik, 2017; Malik et al., 2020).
101 Pollution from sewage, pesticides, mining, intense navigation, and improper waste disposal
102 are primary environmental problems resulting from human actions (Häder et al., 2020; Malik
103 et al., 2020). These issues characterize the Anthropocene era, a geological period defined by
104 the profound influence of human activities on Earth. Unfortunately, these actions have
105 consequences for humans, leading to controversies and illustrating the concept of the "tragedy
106 of the commons" (Dudgeon, 2019). In this sense, ecotoxicological studies have becoming
107 increasingly relevant, particularly in freshwater ecosystems. Indeed, pesticides and trace
108 elements are among the main contaminants found in water bodies (Khoshnood, 2017). These
109 substances are prevalent in surface water and groundwater globally, primarily due to their
110 extensive use in agricultural and urban settings. Pesticides exhibit varying compositions and
111 properties, leading to differences in environmental persistence and toxicity levels. They can
112 have detrimental effects on non-target organisms, including humans, such as carcinogenic,
113 mutagenic, teratogenic, or endocrine-disrupting effects (Hashimi; R. Hashimi; Ryan, 2020;
114 Souza et al., 2020).

115 Trace elements or trace metals (Jarapala; Kandlakunta; Thingnganing, 2014),
116 previously referred to as heavy metals (Duffus, 2002), and are naturally occurring chemical
117 elements found in small concentrations, typically below 0.1% by volume. Some trace
118 elements, such as iron (Fe), zinc (Zn), manganese (Mn), and copper (Cu), are essential for the
119 proper functioning of living organisms, playing critical roles in various physiological
120 processes (Sfakianakis et al., 2015). However, other trace elements such as mercury (Hg),
121 lead (Pb), cadmium (Cd), chromium (Cr), and nickel (Ni) lack recognized biological functions
122 and are generally toxic to a wide range of organisms. It is worth noting that even essential
123 elements can become highly toxic to plants and animals at high concentrations, such as copper
124 (Mela et al., 2013; Tesser; Rocha and Castro, 2021).

125 Trace elements and pesticides pollution present a significant problem due to their
126 bioaccumulation tendency in living organisms, particularly in the liver, muscle, and kidney,
127 even in the absence of detectable concentrations in water (Mela et al., 2014; Ali et al., 2020).
128 In addition to their bioaccumulative nature, trace elements can generate free radicals and
129 reactive oxygen species through oxidative mechanisms, causing DNA damage and impairing

130 crucial enzymes essential for bodily functions (Häder et al., 2020; Nordberg; Nordberg,
131 2022).

132 Biomonitoring has emerged as a method to assess changes in environmental quality
133 using living organisms, allowing for the monitoring of stressors' effects on biological system
134 across various regions worldwide, including Africa (Barhoumi et al., 2012; Saad Abdelkarim,
135 2020), Asia (Saleh; Marie, 2016; Kumar et al., 2021), Europe (Pastorino et al., 2021) and
136 South America (Santana et al., 2018; Montes et al., 2020). The use of biomonitors and
137 biomarkers are crucial in biomonitoring for evaluation of water quality management and
138 conservation (Santana et al., 2018). Among the biomonitors, fish are an excellent biological
139 model for studying aquatic ecosystems due to their constant exposure to environmental
140 conditions. They inhabit diverse aquatic environments, occupy various trophic levels (Calado
141 et al., 2019) and play a significant ecological role in influencing trophic structure, nutrient
142 cycling, and energy flow within the food chain (Kroon; Streten; Harries, 2017). This study
143 used the freshwater fish species *H. malabaricus* (Bloch, 1794, order Characiformes, family
144 Erythrinidae), commonly known as trahira, that is widely distributed in the Neotropical region
145 extends across South America and Central America, including Mexico, in rivers, reservoirs,
146 and lakes, exhibiting ecological plasticity (Chu-Koo; Pérez, 2007; Grassi et al., 2017; Leite et
147 al., 2021; Cruz-Esquivel; Marrugo-Negrete, 2022), and normally consumed by humans
148 (Lozano et al., 2013). As a top predator in the food chain, it is commonly used in
149 toxicological studies as a biomonitoring species (Mela et al., 2014; Paulino et al., 2020;
150 Escalante-Rojas et al., 2021; Leite et al., 2021; Paula; Risso and Martinez, 2021; Cruz-
151 Esquivel; Marrugo-Negrete, 2022). In the Guaraguaçu River of Paraná (Fig. 1), Brazil, this
152 native species holds ecological significance as a potent top predator (Gazola-Silva et al.,
153 2007).

154 Biomarkers provide insights into the effects on organisms at cellular, biochemical, and
155 molecular levels, allowing for the understanding of early responses prior to individual
156 mortality (Van der Oost; Beyer and Vermeulen, 2003). Analysis of biotransformation system,
157 neurotoxicity, genotoxicity and antioxidant system are commonly used biomarkers in
158 biomonitoring studies. Histopathological biomarkers are also employed to assess integrated
159 tissue and organ injuries as they reveal morphological changes, based on the duration and
160 intensity of exposure to the xenobiotic, as well as the adaptive capacity of organisms in cases
161 of chronic exposure where the toxic agent causes cellular injury without resulting in death.
162 (Georgieva et al., 2021; Leão-Buchir et al., 2023).

163 Tropical and sub-tropical freshwater ecosystems from the global south are still
164 understudied compared to those in the Northern hemisphere. One of the still understudied
165 ecosystems is the Guaraguaçu River, a Coastal River in South Brazil, located within a set of
166 estuaries, called ‘Lagamar mosaic’, with high ecological and economical importance
167 considered by UNESCO as a World Heritage Site (<https://whc.unesco.org/en/list/>). Although
168 Lagamar mosaic is one of the most well-preserved remnants of one of the world’s biodiversity
169 hotspots (i.e., the Atlantic Forest, Myers et al. 2000), the Guaraguaçu River have been
170 suffering from intense degradation due to disorderly urban growth, including discharges of
171 domestic sewage and small fishery boats mainly, particularly during the tourist season (Elste
172 et al. 2019). It was expected that more pronounced biomarker responses would be observed in
173 the river sector considered to be more affected by human activities. The aim of this study was
174 to evaluate the influence of the environment and anthropogenic activities on water quality by
175 analyzing chemical analysis and biomarkers of environmental contamination in the top
176 predator fish species *H. malabaricus*. It is important to emphasize that this is the first study
177 using biomarkers in fish from Guaraguaçu River.

178

179 3.4. MATERIALS AND METHODS

180 3.4.1. Study Area

181 The Guaraguaçu River (25°45'W and 48°35'S) is part of the fourth largest sub-basin of
182 the Paraná coastal plain, known as the Guaraguaçu River Basin, with a drainage area of 395.5
183 km². It originates in the Serra da Prata within the Saint-Hilaire/Lange National Park, a well-
184 preserved area in the state of Paraná (Contente; Stefanoni and Spach, 2011), and flows into
185 the Paranaguá Bay through the Cotinga Channel (Cavallini; Reis and Tiepolo, 2020).
186 Spanning 60 km across the coastal plain, the river experiences a subtropical climate
187 characterized by hot, rainy, and humid summers (Cfa). It is renowned for its biodiversity and
188 environmental services, making it the largest and most significant river on the Paraná coast.
189 The municipalities of Pontal do Paraná, Paranaguá, and Matinhos rely on it for their water
190 supply (Vitule; Umbria and Aranha, 2006; Elste et al., 2019).

191 Despite the presence of conservation units such as the Guaraguaçu Ecological Station,
192 Palmito and Rio da Onça State Parks, and the Guaratuba Environmental Protection Area
193 (APA), as well as two indigenous lands inhabited by the Guarani ethnic group (Elste et al.,
194 2019), the Guaraguaçu river area faces constant transformation and degradation. This can be
195 attributed to several environmental impacts, including irregular agricultural areas along the
196 Paraná coast, port activities at the Port of Paranaguá (Contente; Stefanoni and Spach, 2011),

197 mining operations, urban expansion, aquaculture tanks, and the discharge of domestic
198 effluents (Elste et al., 2019). Furthermore, a landfill located in the municipality of Pontal do
199 Paraná is releasing contaminants into one of the river's main tributaries, the Pery River, which
200 has been modified for drainage purposes. These factors contribute to the ongoing
201 environmental deterioration of the area (Singo; Araújo-Ramos and Rocha, 2020).

202

203 3.4.2. Fish and water sampling

204 The fish species *H. malabaricus* was sampled in collaboration with the Laboratory of
205 Analysis and Synthesis in Biodiversity (LASB) and Laboratory of Ecology and Conservation
206 (LEC-DEA) from the Federal University of Paraná (UFPR). The LASB and LEC are
207 responsible for the "Guaraguaçu Project," a long-term project for monitoring the biodiversity
208 of the Guaraguaçu River (see <https://lasbufprbio.wixsite.com/home>).

209 The fish sampling was conducted at 16 specific locations that were characterized and
210 georeferenced by the Guaraguaçu Project. These sampling points were divided into three
211 sectors along the Guaraguaçu River, each representing distinct ecological gradients (Fig. 4).
212 *H. malabaricus* is characterized as an opportunistic sedentary predator, being normally
213 present in the vegetation of the margins of lentic environments, having territorial
214 characteristics due to its reproductive and parental care (Chu-Koo; Pérez, 2007; Montenegro
215 et al., 2013; Gomes et al., 2015). The classification of the river into different sectors was
216 primarily based on the influence of tides, which is one of the main characteristics considered
217 in this division, together with urbanization and human occupation:

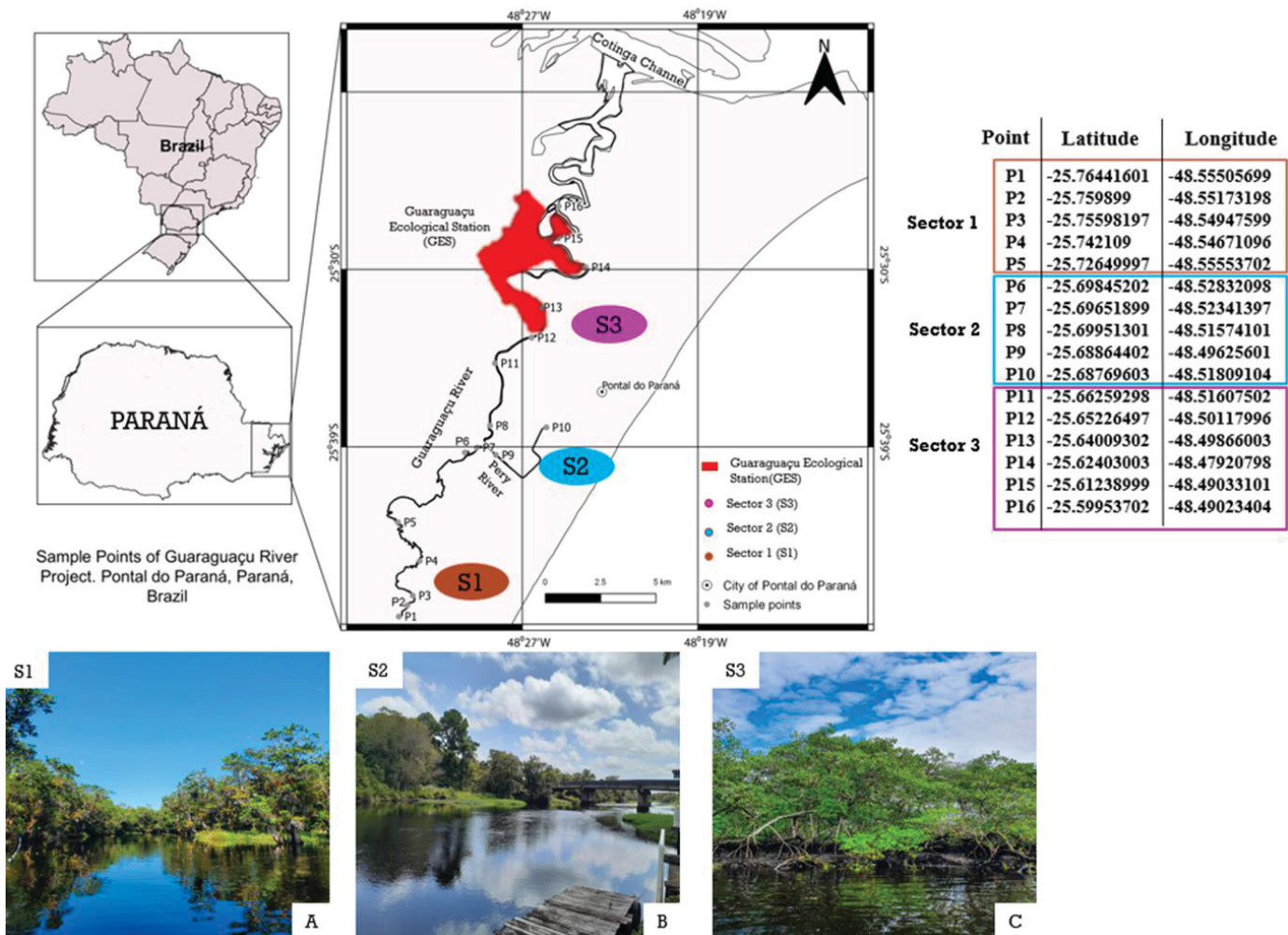
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219 Sector 1 (S1, preserved) (Fig. 4A): More pristine; located upstream of the river near
220 the springs at the base of the mountains; characterized by a well-preserved area with little or
221 no anthropogenic interference. This sector exhibits higher fish abundance in all trophic levels
222 and is not directly influenced by tides. It is notable the presence of a large and dense
223 "caixetal" (pioneer vegetal formation influenced by river).

224 Sector 2 (S2, contaminated) (Fig. 4B): Located downstream; characterized by higher
225 levels of anthropogenic activity. In this area, there are fishermen, ranches, landfill, marinas
226 and irregular housing settlements. The presence of the mining company also contributes to the
227 anthropogenic impact. This sector is more susceptible to pollution due to sewage discharge
228 and presence of landfill, and it may be considered visually the most polluted compared to
229 other sectors of the river. However, the tidal influence is relatively low.

230 Sector 3 (S3, less impacted) (Fig. 4C): The mouth of Paranaguá Bay marks the
 231 estuarine transition (brackish water), mangrove region (pioneer vegetal formation influenced
 232 by river and sea). The presence of Palmito State Park (Guaraguaçu Ecological Station- EEG),
 233 contributed to the conservation efforts. The tidal effects are more pronounced in this sector,
 234 resulting in higher salinity levels compared to upstream areas. Fishing activities are more
 235 prevalent in this sector, reflecting the importance of the estuarine environment for fishery
 236 resources. Additionally, this sector of the river tends to have a wider channel due to the
 237 proximity to the sea (IAT, 2006; Cavallini et al., 2018; Araújo; Vitule and Padial, 2021).

238



239 Fig. 4. Guaraguaçu River and river sectors. A- Sector 1; B- Sector 2; C- Sector 3.

240 Sampling fish of sector 1 was carried out between points 1 and 3 (approximately 600
 241 meters between the points). In sector 2, between points 6 and 8 (approximately 985 meters
 242 between the points), while in sector 3, organisms were collected between points 15 and 16
 243 (approximately 1,400 meters between the points).

244 The fish were collected using fyke net traps with a mesh size of 5 mm and a net of 10
245 x 1 m, as well as wicker fish traps. Twenty-seven specimens were collected, with eleven
246 individuals in sectors 1 and 2, and five in sector 3. There were slight variations among
247 individuals in terms of size, life stage, and male/female ratio (Supplementary Material 1). The
248 sampling was conducted under the authorization of the Brazilian Institute of Biodiversity
249 Conservation (ICMBio), in accordance with the Permanent License for Collection of
250 Zoological Material (SisBio) No. 24779-1 (Authentication Code: 26744745). This license
251 ensures compliance with animal welfare and ethical standards according to international
252 guidelines. Additionally, the species *H. malabaricus* (Bloch, 1794) from the Guaraguaçu
253 River was registered and assigned the number MHNCI 6190 at the Capão da Imbuia Natural
254 History Museum - Curitiba, Paraná, Brazil.

255 The sampling was conducted during March/April, which correspond to the end of
256 summer, and a period with a higher likelihood of high tide influence. This period of year is
257 also characterized by increase rainfall, which can contribute to surface runoff and percolation
258 of waste into the river (Contente; Stefanoni and Spach, 2011). Anthropogenic activities, such
259 as tourism and recreation, are more prevalent during this period, potentially leading to water
260 contamination.

261 During the sampling, water physicochemical parameters including salinity, pH,
262 temperature, conductivity, and transparency were measured. These parameters provide
263 insights into the overall water quality and can help identify potential sources of
264 contamination. The water pH was measured using a portable pH meter PG1400 (GEHAKA).
265 Conductivity was quantified using a CG1400 conductivity meter (GEHAKA). Temperature
266 was determined by calculating the average of temperatures measured by the pH meter and
267 conductivity meter. Transparency was measured using a Secchi Disk. Although salinity was
268 not directly measured with a salinometer due to the logistical limitations, it is present only in
269 sector 3 (transition with the mangrove) (Galvanese et al., 2022) and it was calculated from
270 conductivity by a mathematical conversion.

271 The composite samples of surface water were collected using amber bottles (1L per
272 sector) for pesticide analysis and plastic bottles with the addition of 1.5 ml of ultrapure nitric
273 acid (1L per sector) for trace element analysis. The samples were maintained in ice during the
274 transportation and subsequent laboratory analysis.

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277

278 3.4.3. Samples for biomarkers and chemical analysis

279 The fish were anesthetized using benzocaine and their size and weight were measured.
280 Blood samples were taken from the caudal vein using a heparinized syringe for the
281 micronucleus test. Subsequently, euthanasia was performed by spinal cord sectioning. A
282 fragment of the liver and gills from each specimen was collected for histopathological
283 analysis and fixed in ALFAC solution (80% alcohol, formaldehyde, and glacial acetic acid)
284 for 16 hours.

285 A fragment of muscle, liver, brain, posterior kidney, and gills was sampled for
286 biochemical biomarkers and stored in liquid nitrogen for transport. In the Environmental
287 Toxicology Laboratory (LTA) at the Federal University of Paraná, the samples were stored at
288 -80°C until analysis.

289 Muscle tissue samples from *H. malabaricus* were collected in the field, with five
290 separates muscle samples being obtained per sector of the Guaraguaçu River. These samples
291 were then stored in a freezer for subsequent analysis of trace elements to assess
292 bioaccumulation levels.

293

294 3.4.4. Analysis of pesticides in water

295 After storing the samples in amber bottles in the refrigerator, they were sent to the
296 Technology in Laboratory Analysis (TECLAB). The "United States Environmental Protection
297 Agency" (U.S. EPA, 1996a) Continuous Liquid-Liquid Extraction (EPA 3520 C) method was
298 used for sample preparation and extraction. This method describes a procedure for isolating
299 organic compounds from samples, including appropriate concentration techniques to prepare
300 the extract for the specific analysis. After extraction, analysis was performed using a GCMS
301 240 adv functionality gas chromatograph (Agilent GC) based on the U.S. EPA (2014) Gas
302 Chromatography/Mass Spectrometry (GC-MS) method for Semivolatile Organic Compounds
303 (EPA 8270E).

304

305 3.4.5. Analysis of trace elements in water

306 Cadmium (Cd), lead (Pb), copper (Cu), chromium (Cr), manganese (Mn), and nickel
307 (Ni) were quantitatively analyzed using an Atomic Absorption Spectrophotometer, GBC -
308 Avanta model, at the Multiuser Laboratory of Chemical Analysis (LAMAQ) at the Federal
309 Technological University of Paraná (UTFPR). The analysis method involved a digestion
310 procedure utilizing concentrated nitric acid. Specifically, 100 mL of sample, along with
311 standards, and blank solutions, were heated in Erlenmeyer flasks on a heating plate. Triplicate

312 samples were prepared, with each sample subjected to 2 mL of nitric acid. Subsequently, 100
313 mL of the digested sample were evaporated at a time until reaching 500 mL of evaporated
314 sample for each sector. Once cooled, the residue was transferred to a 100 mL volumetric
315 flask, and 1 mL of a 13% potassium nitrate (KNO_3) solution was added. The final volume of
316 100 mL was achieved by dilution with distilled water.

317 For the preparation of the calibration curve, standards of 0.5 mg/L, 1 mg/L, 1.5 mg/L,
318 and 2 mg/L were used. For the preparation of the blank, 100 mL of distilled water was
319 subjected to the same analytical process as the samples. The standards and the blank were also
320 transferred to 100 mL volumetric flasks and supplemented with a 13% KNO_3 solution, but in
321 a volume of 2 mL. Both the samples and the standards, as well as the blank, were analyzed
322 using the atomic absorption spectrophotometer.

323 The analysis of aluminum (Al), arsenic (As), iron (Fe), and zinc (Zn) followed
324 practically the same method used for the other trace elements, with only some alterations
325 regarding the sample quantity and the equipment used. At the Plant Nutrition Laboratory of
326 UFPR, triplicates of the samples were prepared with a volume of 80 mL for each analyzed
327 sector in Erlenmeyer flasks, under digestion with 1.6 mL of concentrated nitric acid and a
328 constant temperature of 80 °C on a heating plate. Triplicate blanks were prepared with 80 mL
329 of MilliQ water plus 1.6 mL of concentrated nitric acid in each sample. After evaporating the
330 samples to 25 mL, an Inductively Coupled Plasma Optical Emission Spectrophotometer (ICP-
331 OES) instrument, specifically a Varian 720-ES, was used to analyze the trace elements. The
332 results were expressed in mg/L.

333 For mercury (Hg) analysis, an Inductively Coupled Plasma Optical Emission
334 Spectrophotometer (ICP-OES) by Thermo Scientific, model iCAP 6500, with axial view was
335 used. This instrument is located at the Environmental Analysis Laboratory (LAA) of the
336 Department of Chemistry at the UFPR. The analytical curve was constructed using a 100
337 mg/L standard solution of mercury (Hg) from AccuStard (New Haven, USA), which is a
338 mono-element standard solution. The determination of Hg was performed through the
339 chemical vapor generation technique within a concentration range of 0.2 to 5.0 $\mu\text{g/L}$ in a 1%
340 (v/v) nitric acid (HNO_3) medium. The analytical curve exhibited a correlation coefficient
341 above 0.999. For the preparation of solutions for chemical vapor generation, 1 % m/v solid
342 sodium borohydride (NaBH_4) of analytical grade from Reatec (Brazil), 0.4% (m/V) sodium
343 hydroxide (NaOH) of analytical grade from Synth (Brazil), 6 M/L concentrated hydrochloric
344 acid (HCl) of analytical grade from Reatec (Brazil) previously distilled, and 65% HNO_3 from
345 Merck (Germany) were used.

3.4.6. Analysis of trace elements in muscle of *H. malabaricus*

The methodology was based on and adapted from the United States Environmental Protection Agency (U.S. EPA, 1996b) method 3052, which involves microwave-assisted acid digestion of siliceous and organic-based matrices, for the analysis of Cd, Cr, Pb, Ni, Mn, Cu, As, Zn, Fe and Al in the samples.

In the Plant Nutrition Laboratory at UFPR, the samples were placed in an oven at a constant temperature of 40°C for drying. After 24 hours, the dried samples were weighed and reached a constant weight of approximately 0.1 g. The samples were then transferred to Teflon tubes for microwave digestion. To initiate the digestion, 3 mL of HNO₃ was added to each sample and allowed to react for another 24 hours. Subsequently, an additional 1 mL of nitric acid and 1 mL of hydrogen peroxide (H₂O₂) 35% were added, and the sample was left to rest for another day for the reaction of HNO₃ and H₂O₂. Before placing the samples in the microwave, 3 mL of Milli-Q water (ultrapure) were added to each Teflon tube. During the digestion process, triplicate blank solutions were also prepared in a similar manner to the samples. The digestion process was carried out using a MARS 6 microwave digestion system.

The heating program in the microwave consisted of a pre-digestion stage with two steps. The first step involved a 5-minute heating ramp up to 80°C, followed by another 5-minute ramp up to 130°C. Finally, there was a 15-minute cooling period. For the digestion process, two stages were used with a 5-minute heating ramp up, varying power from 1030-1800 W, temperature ranging from 125-180°C, each stage lasting 10 minutes. After digestion, Milli-Q water was added to the samples to reach a volume of 20 mL. An Inductively Coupled Plasma-Optical Emission Spectrometry (ICP-OES) instrument, specifically a Varian 720-ES, was used to analyze the trace elements. The results were expressed in mg/kg (= µg/g).

3.4.7. Biochemical biomarkers

The muscle and brain were homogenized in potassium phosphate buffer (0.1 M, pH 7.5) at a ratio of 1:10 (w/v) using a micro homogenizer, and centrifuged for 20 minutes at 12,000 xg, 4 °C. For the muscle and brain, the supernatant was used for acetylcholinesterase (AChE) activity. The analysis of AChE activity was based on the method of Ellmann et al. (1961), modified for microplate by Silva de Assis (1998). The absorbance was read at a wavelength of 405 nm.

The liver was homogenized in potassium phosphate buffer (0.1 M, pH 7.0) at a ratio of 1:10 (w/v), while the kidney at a ratio of 1:5 (w/v), and then centrifuged for 30 minutes at 15,000 xg, 4 °C. Aliquots of the supernatant were taken for measurement of glutathione S-

380 transferase (GST) activity, superoxide dismutase (SOD) activity, catalase (CAT) activity,
381 glutathione peroxidase (GPx) activity, concentration of non-protein thiols (GSH), and
382 lipoperoxidation (LPO). The gills were homogenized in potassium phosphate buffer (0.1 M,
383 pH 7.0) at a ratio of 1:5 (w/v), followed by centrifugation for 30 minutes at 15,000 xg, 4 °C.
384 Aliquots of the supernatant were taken for measurement of GST activity, SOD activity, CAT
385 activity, GPx activity, GSH concentration, and LPO.

386 The analysis of Glutathione S-transferase (GST) activity followed the method
387 proposed by Keen et al. (1976) based on the catalysis of the reaction between the substrate 1-
388 chloro-2,4-dinitrobenzene (CDNB) and reduced glutathione (GSH) by GST. The
389 measurement was performed at a wavelength of 340 nm.

390 The activity of SOD was evaluated using the method proposed by Gao et al. (1998),
391 which is based on the ability of superoxide dismutase to inhibit the auto-reduction of
392 pyrogallol. The absorbance was measured at a wavelength of 440 nm.

393 The method for CAT analysis was based on Aebi (1984). The principle of the method
394 is to evaluate the decrease in absorbance at 240 nm due to the consumption of H₂O₂ by
395 catalase, resulting in the production of oxygen (O₂) and water (H₂O). The absorbance was
396 measured at 240 nm.

397 The analysis of GPx activity was performed using the method described by Hafeman
398 et al. (1974). Based on the measurement of the decrease in absorbance at 340 nm, caused by
399 the reduction of oxidized glutathione (GSSG), which is catalyzed by glutathione reductase
400 (GR) in the presence of nicotinamide adenine dinucleotide phosphate (NADPH).

401 The concentration of GSH was analyzed using the method developed by Sedlak and
402 Lindsay (1968). The absorbance was measured at 405 nm. The concentration of GSH was
403 determined by comparing it to a standard curve of GSH.

404 The analysis of LPO was performed using the FOX assay (Ferrous Oxidation in
405 Xylenol Orange), proposed by Jiang et al. (1992). This method is based on the rapid oxidation
406 of iron (Fe⁺²) mediated by peroxides under acidic conditions, followed by the formation of the
407 Fe⁺³-xylenol orange complex in the presence of the stabilizer butylated hydroxytoluene. The
408 absorbance was measured at 570 nm.

409 A fragment of liver was used for metallothionein (MET) analysis. The liver was
410 homogenized in a solution consisting of Tris HCl buffer (20 mM, pH 8.6), 1.71 g of sucrose,
411 50 µL of phenylmethylsulfonyl fluoride (PMSF), and 1 µL of β-mercaptoethanol, at a ratio of
412 1:5 (w/v). The homogenate was then centrifuged for 30 minutes at 15,000 xg, 4 °C. From the
413 supernatant, two aliquots were taken: 300 µL for metallothionein activity analysis and 10 µL

414 for protein analysis. The method was proposed by Viarengo et al. (1997) and the absorbance
415 was measured at a wavelength of 405 nm. A negative control was performed, as well as a
416 standard curve using GSH.

417 The Bradford method (1976) was used for protein quantification in the samples, with a
418 standard curve prepared using bovine serum albumin.

419

420 3.4.8. Histopathological biomarkers

421 In the laboratory, the ALFAC solution was replaced with 70% alcohol and kept for 24
422 hours, followed by another change to 70% alcohol and kept until the samples were embedded
423 in Paraplast®. The liver and gill samples were cut using a microtome, with a thickness set at 5
424 µm, and stained with hematoxylin/eosin. Images were captured using an Olympus BX51
425 microscope at the Center for Advanced Fluorescence Technologies (CTAF-UFPR). The
426 Lesion Index was calculated based on histopathological findings, according to Bernet et al.
427 (1999) and modified by Mela et al. (2013). All identified lesions and tissue alterations were
428 classified into categories based on their biological significance: 1 - minimal lesion, easily
429 reversible; 2 - moderate lesion, reversible in most cases; and 3 - severe lesion, generally
430 irreversible. Scores ranging from 0 to 6 were assigned to establish the severity of the lesions.
431 The lesion index for each group of liver or gill lesions was calculated using the following
432 formula:

$$433 \quad I_{org} = \sum_{rp} \sum_{alt} (a \times w)$$

434 , where: org represents the organ (constant), rp represents the reaction pattern, alt
435 represents the alteration, a represents the score value and w represents the importance factor
436 of the lesion.

437 For scanning electron microscopy analysis of gills, the samples were fixed and stored
438 in 3% glutaraldehyde for a minimum period of 24 hours. They then underwent a gradual
439 dehydration process in ethanol (from 50% to 100% in 10-minute intervals), followed by
440 critical point drying using a Bal-tec CPD 030 instrument. Once the samples were completely
441 dehydrated, they were gold-coated using a Balzers SCD 050 device. Finally, readings and
442 images of the tissue lesions were obtained using a JEOL JSM 6360-LV scanning electron
443 microscope (SEM) at the Electron Microscopy Center of UFPR.

444

445

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447 3.4.9. Biomarkers of genotoxicity

448 To assess the frequency of micronuclei, the technique described by Hooftman and
449 Raat (1982) was used. In the field, a drop of blood was placed on a microscope slide, and
450 using a 45-degree angle, a coverslip was slid to perform a smear, resulting in one slide per
451 fish. The slides were left to air dry and then fixed in absolute ethanol for 15 minutes. Once
452 dry, the slides were stained with 10% Giemsa diluted in 90 mL of phosphate/sorensen buffer
453 (pH 6.8) and left for 12 minutes. For each slide, 2000 cells/erythrocytes were analyzed under
454 a 100x magnification using an optical microscope. The frequency of the following nuclear
455 morphological alterations, proposed by Carrasco et al. (1990), was also determined in the
456 blood samples: Blebbed (BL), Lobed (LB), Vacuolated (VC), Notched (NT), and Binucleus
457 (BN).

458

459 3.4.10. Statistical analysis

460 The analysis of the data was performed by examining the variation in biomarker
461 responses (response variable) among the three sampling sectors (predictor variable).

462 The Levene and Shapiro-Wilk tests were used to assess the assumptions of
463 homogeneity of variances and normality, respectively. The assumptions were met in most of
464 the analyses, allowing the use of one-way ANOVA followed by Tukey's test. In cases where
465 the assumptions were not met, a permutation ANOVA was performed or a data log
466 transformation was made. The results were expressed as mean \pm standard error with a
467 significance level of $p < 0.05$. For an integrated analysis of the biochemical biomarker
468 responses in the gills, liver, and kidney, multivariate statistics using Principal Coordinates
469 Analysis (PCoA) were employed, considering only the first two axes to describe differences
470 between sectors and similarities among biomarkers. The analysis was conducted using
471 RStudio software (R CORE TEAM, 2017), and the PCoA graphs were generated using
472 STATISTICA software.

473 For the analysis of trace elements in the muscle of *H. malabaricus*, a PERMANOVA
474 (Permutation Multivariate Analysis of Variance) with Euclidean distance matrix was
475 performed using RStudio. This analysis was conducted to compare the influence of trace
476 elements among the sampled sectors. The PERMANOVA allows for the assessment of
477 differences among groups while considering multivariate data.

478 The physicochemical parameters did not undergo statistical analyses. This is mainly
479 due to the nature of quantifying and obtaining data related to a "snapshot" period of the
480 parameters, as the Guaraguaçu River is influenced by tides that occur twice a day, during the

481 flood tide and ebb tide. In addition, water features were estimated in only one sampling point
 482 per sector, so there were no real replicates to compare sites. As a result, it becomes
 483 impractical to conduct statistical analysis for comparing the sectors in terms of
 484 physicochemical parameters, as they do not represent concrete daily patterns or even have
 485 independent replicates. However, the findings were able to describe trends in the river, as well
 486 supported by previous studies (IAT, 2006; Vitule; Umbria and Aranha, 2006; Elste et al.,
 487 2019).

488

489 3.5. RESULTS

490 3.5.1. Physicochemical parameters

491 Although with only one snapshot sampling, analysis of the water's physicochemical
 492 parameters (Table 1) suggests an apparent difference into the three sectors, particularly
 493 regarding conductivity and transparency. The conductivity in sector 2 was higher than in
 494 sector 3, which is directly influenced by tides. When analyzing transparency, there was a scale
 495 from greater to lesser transparency.

496 Table 1. Mean values and standard errors (Mean \pm SE) of the physical-chemical parameters of the sixteen
 497 collection points of the "Guaraguaçu Project" divided to characterize the three sectors of the Guaraguaçu River.

Sector	Conductivity (uS/cm ²)	pH	Transparency (cm)	Temperature (°C)	Salinity (ppm)
1	26.3 \pm 1.18	4.8 \pm 0.18	92.4 \pm 10.40	23.3 \pm 0.16	0.0089
2	74.9 \pm 25.20	5.8 \pm 0.18	81.6 \pm 18.60	23.6 \pm 0.19	0.0217
3	49.5 \pm 1.94	5.4 \pm 0.29	76.3 \pm 3.57	23.4 \pm 0.06	0.0177

498 Legend: Points 1-5 = Sector 1; Points 6-10 = Sector 2; Points 11-16 = Sector 3. Points 8 and 9 (Pery rectified
 499 channel) of the guaraguaçu project belonging to sector 2 had extremely high conductivity: 154.4 and 110 uS/cm
 500 respectively. Salinity is expressed only as a mean value.

501

502 3.5.2. Chemical Analysis

503 3.5.2.1. Pesticides in water

504 The intermediate region of the Guaraguaçu River can be considered the most impacted
 505 by anthropogenic stressors. However, the results of the analysis of pesticides in water
 506 (Supplementary Material 2) showed that these contaminants are not present in significant
 507 quantities in any of the three sectors. All sectors had concentrations below the limit of
 508 quantification (LQ) and, therefore, when compared to CONAMA 357/05, almost all are below
 509 the maximum concentration allowed by the legislation, except the pesticides Aldrin +
 510 Dieldrin, Pentachlorophenol, Chlordane, Endrin and Lindane. Because as some are below the

511 quantification limit, there is no way to know the exact value, besides that there are some
512 pesticides that do not have a limit by legislation (see all data in Supplementary Material 2).

513

514 3.5.2.2. Trace elements in water

515 The Guaraguaçu River and its tributaries are classified for the most part, according to
516 CONAMA 357/05 as class 2, water that can be used for human consumption after
517 conventional treatment, protection of aquatic communities, recreation, irrigation of
518 vegetables, aquaculture, and fishing activities (ZEE, 2016).

519 The results showed that the Guaraguaçu River region does not exhibit significant
520 contamination by no essential trace elements, at least when analyzed in water (Supplementary
521 Materials 3, 4 and 5). Manganese (Mn), an essential element, was quantified, but its
522 concentration in all three sectors were below the maximum limit allowed by legislation (total
523 manganese - maximum of 0.1 mg/L) according to CONAMA 357/05. Fe and Al were
524 quantified and are above the legal limits (Dissolved Aluminum - maximum of 0.1 mg/L;
525 Dissolved Iron - maximum of 0.3 mg/L) according to CONAMA 357.

526

527 3.5.2.3. Trace elements in the muscle of *H. malabaricus*

528 In the water samples, manganese, iron and aluminum were quantified. However, in the
529 analysis of trace elements in fish muscle tissues, several elements were quantified, including
530 chromium (Cr), copper (Cu), manganese (Mn), iron (Fe), zinc (Zn), aluminum (Al), and
531 arsenic (As), which were included due to their biological importance in the metabolism and
532 physiological processes of aquatic organisms (Supplementary Material 5). Brazilian
533 legislation attempts to establish limits for trace elements in food by Resolution of the
534 Collegiate Board - RDC (2013). It establishes the MERCOSUR Technical Regulation on
535 maximum limits for inorganic contaminants in food (Brazil, 2013) and Decree No. 55.871
536 (1965), which, although repealed in 2019, remains the only document that establishes
537 maximum limits for nickel, zinc, copper, and chromium in food (BRAZIL, 1965). Comparing
538 Brazilian legislation is largely in line with international regulations as Food and Agriculture
539 Organization (FAO, 1983) and Commission Regulation of the European Community (EC,
540 2006).

541 Among the trace elements analyzed, lead (Pb), cadmium (Cd), and nickel (Ni) were
542 not quantified in the muscle tissue of *H. malabaricus* in any of the three sectors, similar to

543 water samples. Sector 3 was the only sector where at least one organism had concentrations of
544 As and Cu above the limit of quantification of the equipment (Arsenic = 0.02 mg/L; Copper =
545 0.01 mg/L). However, the copper concentration remained below the limit determined by
546 legislation. Al, Fe and Mn do not have a maximum limit of concentration in food according to
547 the legislation. Aluminum was quantified in all organisms from sector 3 of the Rio
548 Guaraguaçu. The arsenic value found in a single organism from sector 3 (5.569 mg/kg)
549 exceeded the concentration limit stipulated by the legislation (As = 1 mg/kg). Chromium was
550 found in two organisms from sector 2 (3.469 and 2.281 mg/kg) and one organism from sector
551 3 (5.551 mg/kg), exceeding the limit (Cr = 0.1 mg/kg). When analyzing the concentration of
552 zinc, only five organisms out of fifteen had concentrations below the limit stipulated by
553 Decree No. 55,871 of 1965 (50 mg/kg). Iron and zinc were the only elements quantified in all
554 organisms analyzed in all sectors.

555 The correlation of each original variable (trace elements) with the x and y axes (PCA1
556 and PCA2, respectively), revealed that two elements stood out in explaining nearly 99% of
557 the influence on the sectors of the Guaraguaçu River (Supplementary Material 7). The x-axis
558 (PCA1) explained approximately 82% of the variation and, according to the loadings, was
559 positively associated with the concentration of Zn. On the other hand, the y-axis (PCA2)
560 explained approximately 17% of the variation and was negatively associated with Fe. By
561 analyzing the ordination graph (Fig. 5), it can be observed that Fe is more correlated with
562 sector 3, while Zn is more correlated with S1. However, from a statistical point of view, there
563 is no significant difference among the three sectors regarding trace element concentrations (p-
564 value = 0.19 > 0.05) (Supplementary Material 6).

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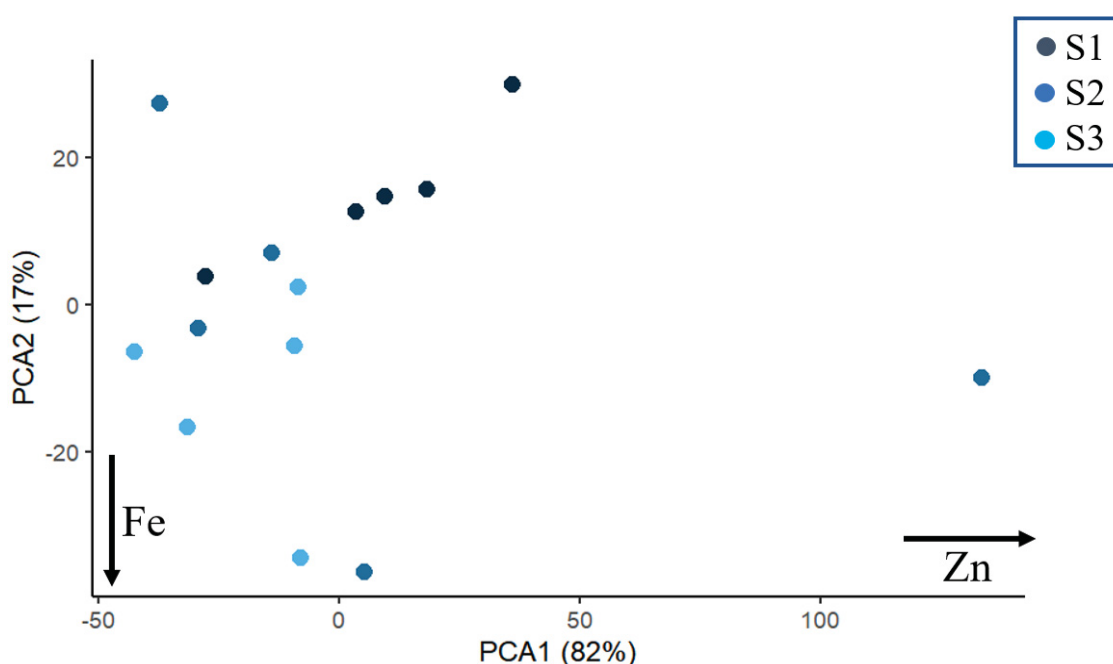
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571 Fig. 5. Principal Coordinate Analysis ordination, showing the first two axes (PCA1 and PCA2) that represent
 572 most of the data variation (percentages in the graph). The Fe and Zn elements are the two that are mostly related
 573 to the axes, with high Fe values mainly in sector 3(S3), and high Zn values in sector 1 (S1).
 574

575 3.5.3. Biochemical biomarkers

576 The Guaraguaçu River showed significant differences among the three sectors
 577 analyzed based on biochemical biomarker analysis in the brain, muscle, gills, liver, and
 578 kidney (Supplementary Material 8). In the brain, there was a significant difference in AChE
 579 activity ($F=10.17$; $p=0.0008$) (Fig. 6A), with a low activity observed in S2, which is
 580 considered more impacted by human activities.

581 In muscle tissue, a significant difference was observed ($F= 7.95$; $p = 0.002$) among the
 582 sectors, with a low acetylcholinesterase activity in S3, which is less impacted, compared to
 583 S2, which is more influenced by human activities (Fig. 6B). However, there was no
 584 significant difference between S1 (control sector and considered pristine) and the other two
 585 sectors.

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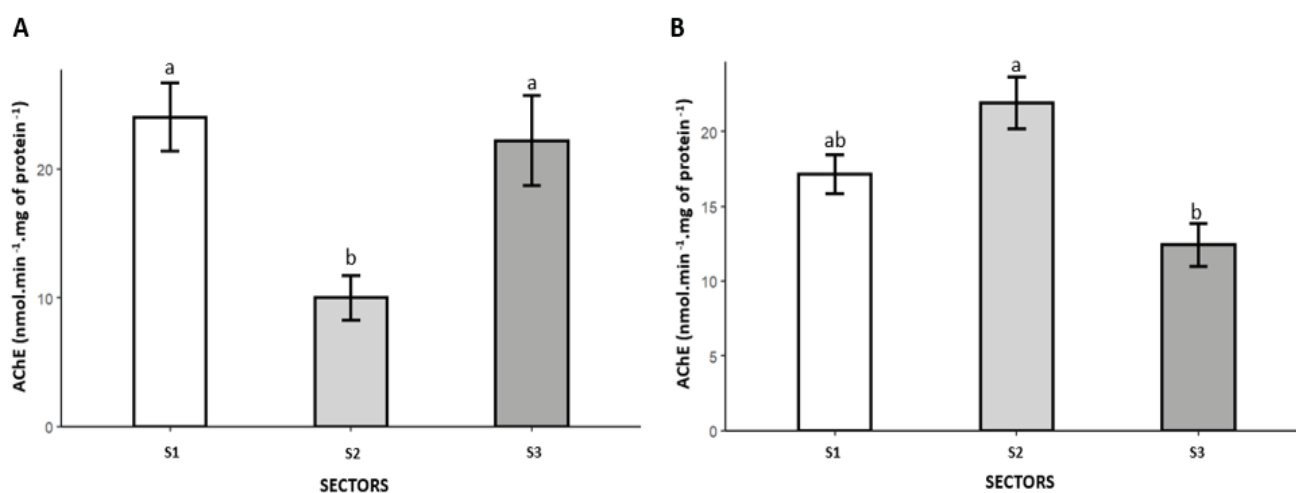
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595 Fig. 6. Acetylcholinesterase (AChE) activity (Mean \pm Standard Error) in the brain (A) and muscle (B) of *Hoplias*
 596 *malabaricus* in three sectors characterized by different ecological gradients. Different letters indicate significant
 597 difference ($p < 0.05$) by Tukey's test.

598 In the comparison among the three sectors regarding the effects of biomarker activities
 599 in the fish gills, a higher activity of GST was observed in S3 ($F = 7.96$; $p = 0.003$). For GSH,
 600 there was no significant difference among the three sectors ($F = 1.55$; $p = 0.244$), and the
 601 activity of CAT decreased in S2 and S3 ($F = 8.46$; $p = 0.002$). However, there was an increase
 602 in GPx activity ($F = 8.48$; $p = 0.003$), LPO ($F = 17.33$; $p < 0.01$), and SOD activity ($F = 11.24$; p
 603 $= 0.0006$) in S2 and S3. Analyzing the Principal Coordinate Analysis (PCoA) (Fig. 7A),
 604 created to interpret the percentage of explanation that the biomarkers exert on the sectors of
 605 the Guaraguaçu River, it is considered that, except for the higher CAT activity in S1, all other
 606 biomarkers showed a higher correlation and activity in S2 and S3, in addition to an increase in
 607 LPO. In fact, there is a visual separation among the sectors considering the set of biomarkers,
 608 where S1 is the one that stands apart from the others.

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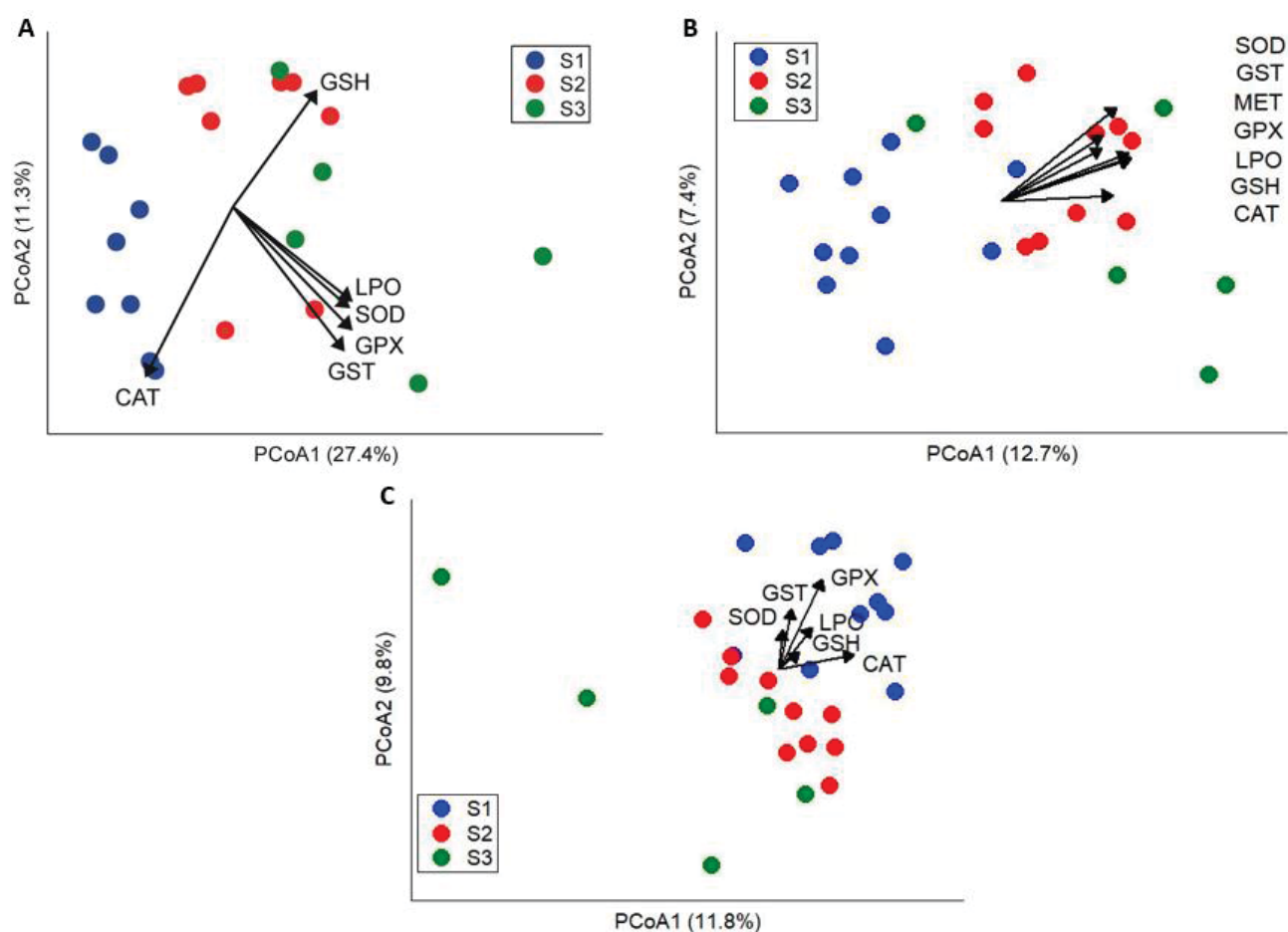
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616 Fig. 7. Principal Coordinate Analysis ordination of biochemical biomarkers in gills (A): Axis 1 (PCoA1)
 617 explains 27.4% of the variation, while Axis 2 (PCoA2) explains 11.3% of the variation; PCoA of biochemical
 618 biomarkers in liver (B): Axis 1 (PCoA1) explains 12.7% of the variation, while Axis 2 (PCoA2) explains 7.4%
 619 of the variation. The order of the arrows is directly related to the order of the biomarkers in the upper right
 620 corner of the graph; and PCoA of biochemical biomarkers in kidney (C): Axis 1 (PCoA1) explains 11.8% of the
 621 variation, while Axis 2 (PCoA2) explains 9.8% of the variation.

622 In terms of the effects of biomarker activities found in the liver, it can be noted that
 623 there was an increase in the GST activity ($F= 3.68$; $p= 0.04$) and GPx activity ($F= 5.97$; $p=$
 624 0.01) in S2 compared to S1, but there was no significant difference between S3 and the other
 625 two sectors in the activity of both enzymes. However, for GSH ($F= 25.79$; $p< 0.01$), SOD
 626 activity ($F= 16.29$; $p<0.01$), and LPO ($F= 17.39$; $p<0.01$), there was an increase in S2 and S3
 627 when compared to S1. CAT activity ($F= 1.33$; $p= 0.28$) and MET ($F= 0.71$; $p= 0.05079$) did
 628 not show significant differences among the three sectors. The PCoA used to illustrate the
 629 collective responses of the liver biomarkers (Fig. 7B) shows that the biomarkers, for the most
 630 part, exhibited higher responses in Sectors 2 and 3, with S2 showing higher activity than S1 in
 631 almost all biomarkers. As in the gill biomarkers, there is a clear separation among sectors for
 632 the combination of liver biomarkers, highlighting the regionalization of health impacts on the
 633 fish in the Guaraguaçu River.

634 The posterior kidney is known to have the primary function of excreting contaminants
635 and their metabolites from organisms, making it extremely important for biomarker analysis.
636 In the case of the posterior kidney, it was found that the only biomarkers that showed
637 significant differences among the three sectors were GST ($F= 18.31$; $p<0.01$) and GPx ($F=$
638 14.82 ; $p = 0.0001$), with increased activities in sectors 2 and 3 (Fig. 7C). Similar to the
639 previous results, there is a distinct difference among the sectors when considering the set of
640 biomarkers in the kidney.

641

642 3.5.4. Histopathological biomarkers

643 The histopathological biomarkers demonstrated a spatial difference in the
644 Guaraguaçu River among the three sampling sectors, when analyzed using optical
645 microscopy in gills (Fig.8 A-F) and liver (Fig. 9 A-D) and scanning electron microscopy in
646 gills (Fig. 8 G-L), indicating the likely presence of xenobiotics such as trace elements. More
647 pronounced responses of the biomarkers, consequently leading to greater tissue lesions and
648 alterations, were observed in the fish collected in sector 2, considered to be more affected
649 by anthropogenic activities. When analyzing the mean Bernet index for each sector in each
650 tissue relative to the number of individuals collected in each sector, a significant difference
651 was found among the three sectors both for gills ($p<0.05$; $F\text{-value} = 98.9$) (Fig. 8M) and for
652 liver ($p<0.05$; $F\text{-value} = 25.6$) (Fig. 9E), with sector 2 showing the highest indexes (25.27
653 and 24.18, respectively for gills and liver), followed by sector 3 (with 3.2 and 7.6), and
654 lastly sector 1 (with 0.73 and 2.91).

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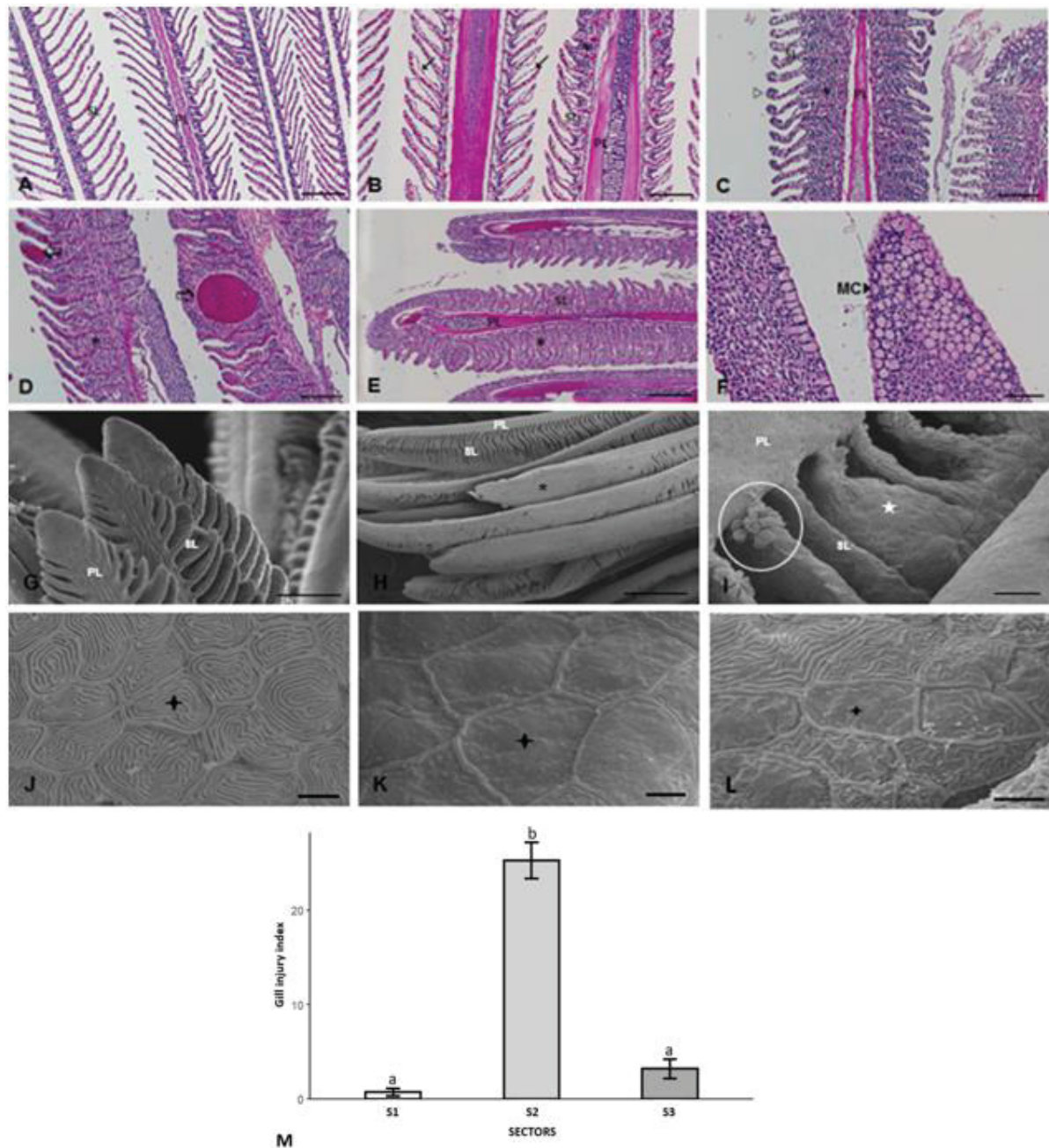
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665 Fig. 8. Histological gill sectors of *H. malabaricus* counterstained with hematoxylin/eosin using optical
 666 microscopy (A-F), and images using scanning electron microscopy (G-L). (A) Gill of sector 1: Primary
 667 lamella (PL) and secondary lamella (SL); (B) Gill sector 3: Primary lamella (PL), secondary lamella (SL),
 668 epithelial lifting (✓) and partial hyperplasia (*); (C) Gill sector 2: Primary lamella (PL), partial hyperplasia (*)
 669 in secondary lamella (SL) and rolling of secondary lamella (▷); (D) Gill sector 2: Aneurysm (⇔) and partial
 670 hyperplasia (*) in secondary lamella (SL); (E) Gill sector 2: Primary lamella (PL) and total hyperplasia of
 671 secondary lamella (SL); (F) Gill sector 2: Total hyperplasia of secondary lamella (SL) and increase of mucus
 672 cells (MC). Scale bar: (A-E) 100 μm, (F) 50 μm. (G) Gill of sector 1: General view of gill filaments and
 673 lamellae showing normal morphological features. Primary lamellae (PL) and secondary lamellae (SL). Scale
 674 bar: 50 μm; (H) Gill of sector 2: Primary lamellae (PL) and secondary lamellae (SL). It is possible to observe
 675 Hyperplasia in secondary lamella (SL) with lamellae fusion (*). Scale bar: 100 μm; (I) Gill of sector 2:
 676 Primary lamellae (PL) and secondary lamellae (SL), partial hyperplasia (in circle) and aneurysm (★). Scale
 677 bar: 20 μm; (J) Gill of sector 1: Note well-organized pavement cells and organized microridges (✦). Scale

678 bar: 5 μ m; **(K)** Gill of sector 2: We can see alteration in the branchial epithelium with significant reduction of
679 microridges (*) in pavement cells. Scale bar: 5 μ m; **(L)** Gill of sector 3: Branchial epithelium with reduction
680 of microridges (*) in pavement cells. Scale bar: 5 μ m.; **(M)** Graph of the histopathological lesion index in the
681 gill tissue. Results are expressed as mean \pm standard error for normal data. Analysis of variances ANOVA (p
682 <0.05 ; F-value = 98.9). Different letters mean statistically significant differences between sectors.

683 The changes found in the gills, particularly in S2, included aneurysm and hyperplasia
684 with partial fusion of the secondary lamellae (Fig. 8D), folding of the tips of the secondary
685 lamellae (Fig. 8C), hyperplasia with total fusion of the secondary lamellae and an increase in
686 mucous cells (Fig. 8F), with a higher percentage of occurrence in sector 2 related to increase
687 of mucous cells (27.17%) and hyperplasia with total fusion of the secondary lamellae
688 (23.20%), classified as level 1 (minimal lesion) and level 2 (moderate lesion), respectively, in
689 the biological importance grading of the Bernet index (Bernet et al., 1999). In S3, the
690 observed alterations were epithelial lifting and hyperplasia with partial fusion (Fig. 8B).
691 When analyzing the images captured by SEM, alterations in the gill epithelium with reduced
692 microridges were observed in gills of fish from S2 and S3 (Fig. 8K and 8L), with major
693 alterations found in S2. In S1, intact gill structures without tissue alterations were observed
694 (Fig. 8A and Fig. 8G).

695 In the liver, most of the alterations also occurred in fish tissues from sector 2 of the
696 Guaraguaçu River. Among the alterations found in fish from sector 2, the presence of
697 melanomacrophage centers (Fig. 9B), blood congestion in sinusoids, hepatocyte
698 vacuolization (Fig. 9D), and necrosis can be mentioned. The alterations related to
699 hepatocyte vacuolization (30%) and necrosis (26%) had a higher percentage of occurrence
700 in sector 2, indicating two irreversible changes. In fish from sector 3, which is considered to
701 have less anthropogenic influence, the presence of cell death or necrosis (Fig.9C) was still
702 observed.

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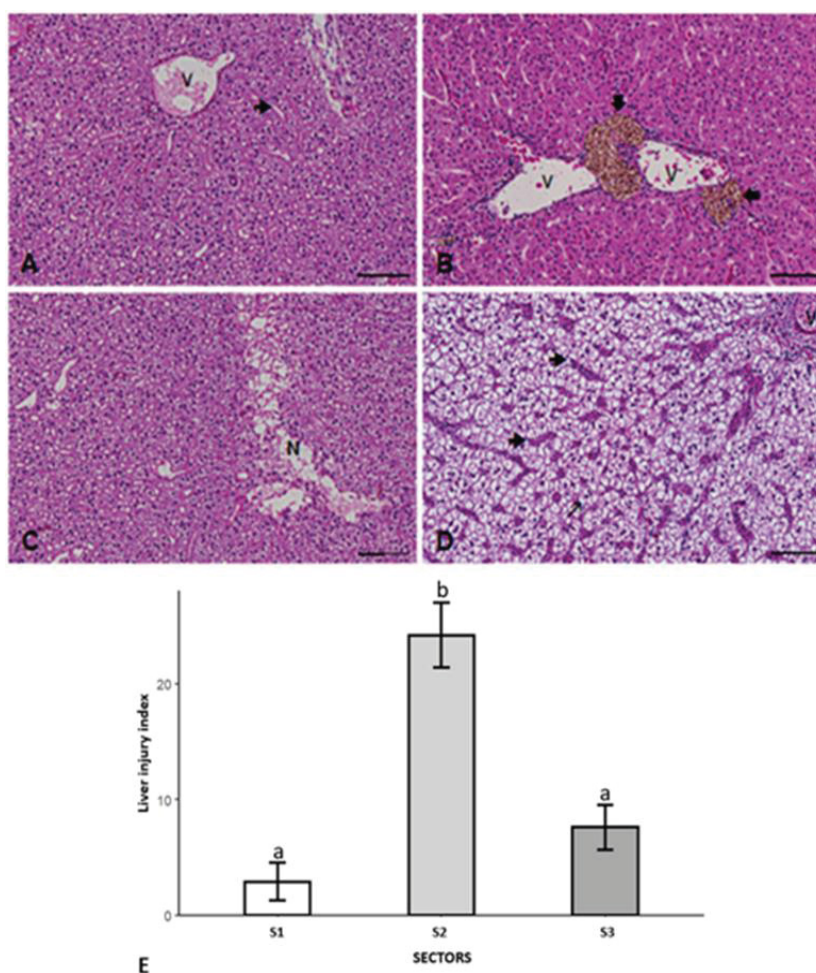
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714 Fig. 9. Histopathological liver of *H. malabaricus* counterstained with hematoxylin/eosin. (A) Sector 1: Blood
 715 vessel (V) and sinusoids (➔); (B) Sector 2: Melanomacrophages centers (➡) surrounding the blood vessels;
 716 (C) Sector 3: Necrotic foci (N); (D) Liver sector 2: Blood vessel (V), blood congestion in sinusoids (➔) and
 717 vacuolation of hepatocytes (↗). Scale bars = 100 μ m; (E) Graph of the histopathological lesion index in the
 718 liver tissue. Results are expressed as mean \pm standard error. Analysis of variances ANOVA ($p < 0.05$; F-value
 719 = 25.6). Different letters mean statistically significant differences.

720

721 3.5.5. Biomarker of genotoxicity

722 The results showed that there was no statistically significant difference in the presence
 723 of micronuclei or other nuclear morphological alterations (blebbed, notched, lobed, binucleus,
 724 and vacuolated) among the three analyzed sectors of the Guaraguaçu River (Table 2). From
 725 the analysis of 2000 erythrocytes, the presence of micronuclei occurred in all three sectors,
 726 with three micronuclei found in S1, two in S2, and two in S3, totaling seven micronuclei for
 727 all the blood samples analyzed.

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731

732 Table 2. Frequency of Micronucleus and nuclear morphological alterations in erythrocytes of *H. malabaricus* in
 733 the three sectors of Guaraguaçu River. (Median; 1st quartile; 3rd quartile).

Section	Statistical	Nuclear morphological alterations					
		Micronucleus	Blebbled	Notched	Lobed	Binucleus	Vacuolated
	F-value	0.285	0.386	3.381	2.303	0.277	1.635
	P-value	0.754	0.683	0.050	0.121	0.760	0.215
S1		3 (0; 0; 0.5) a	17 (1; 0; 2) a	50 (4; 2.5; 6.5) a	17 (1; 0; 2) a	5 (0; 0; 1) a	98 (10; 5.5; 12) a
S2		2 (0; 0; 0) a	14 (1; 0; 3) a	25 (2; 1; 3) a	26 (1; 1; 4) a	3 (0; 0; 0.5) a	70 (3; 1; 8) a
S3		2 (0; 0; 0) a	4 (1; 0; 1) a	27 (5; 3; 6) a	2 (0; 0; 1) a	2 (0; 0; 1) a	64 (11; 11; 12) a

734 Statistics were performed by ANOVA test followed by Tukey's post hoc test. When normality assumptions were
 735 not found, an ANOVA with permutations was performed. Same letter means no significant difference ($p > 0.05$ or
 736 $p = 0.05$).

737

738 3.6. DISCUSSION

739 The Guaraguaçu River have a great social and economic importance in providing
 740 water supply and subsistence fishing (Reis et al., 2015) mostly to social-vulnerable residents
 741 of Pontal do Paraná, Paranaguá and Matinhos municipalities (Elste et al., 2019). In addition,
 742 this river plays an important seasonal role for sport fishing tourism; and is central to aquatic
 743 biodiversity conservation (Vitule; Umbria and Aranha, 2006). In spite of that, our results
 744 clearly demonstrated that human impacts are reflecting in environmental health of the river.

745 This is in line with the recent ecological studies carried out in Guaraguaçu River.
 746 Eutrophication process and mass-development of invasive aquatic macrophytes, particularly
 747 the African tanner-grass *Urochloa arrecta* (Hack. ex T. Durand & Schinz) Morrone &
 748 Zuloaga occurs mainly in the intermediate region of the river (Elste et al., 2019; Araújo;
 749 Vitule; Padial, 2021). Recent studies have reported the biotic homogenization impacts related
 750 to the gradient of anthropogenic impact found in the Guaraguaçu River (Sato; Costa; Padial,
 751 2021; Galvanese et al., 2022). Here, we add to this knowledge by investigating how the health
 752 of a top predator fish is affected by the human impact gradient of the Guaraguaçu River.

753

754

755

756 3.6.1. Physicochemical parameters

757 The results are in line with the well-known increase in contamination and degradation
758 along the intermediate stretches of the Guaraguaçu River (Elste et al. 2019), posing a threat to
759 its multiple uses. The lower pH in Sector 1, which is considered pristine is influenced almost
760 exclusively by the Serra do Mar (Abreu-Mota et al., 2014). The biogenic material imported
761 from upstream rivers settles in this region due to its almost stagnant water character, resulting
762 from low water flow. Because of organic matter decomposition, the environment becomes
763 more acidic, giving the water a brownish coloration. Additionally, the absence of a constant
764 flow of vessels leads to minimal particulate matter, which explains the higher water
765 transparency in this sector.

766 It can be observed that the conductivity in sector 2 is higher than in sector 3, which is
767 influenced by tidal effects due to its proximity to the mouth of the Guaraguaçu River.
768 Consequently, sector 3 would be expected to have the highest conductivity and salinity in the
769 river (Galvanese et al., 2022). This can be mainly explained by highlighting that Points 8 and
770 9 of the "Guaraguaçu Project," with conductivities of 154.4 and 110 $\mu\text{S}/\text{cm}$, respectively, are
771 located within the straightened channel of the Pery River, near the landfill in the municipality
772 of Pontal do Paraná, and in the presence of domestic sewage discharge. These points possible
773 be influenced by increased conductivity due to excessive concentrations of elements such as
774 phosphorus and nitrogen, and their ions, which are naturally limiting factors for biotic growth
775 (Souza et al., 2019; Singo; Araújo-Ramos and Rocha, 2020). This finding confirms and
776 strengthens the results of other studies conducted in different water bodies worldwide, where
777 anthropogenic nutrient inputs significantly alter physicochemical parameters such as
778 conductivity (Arif; Kumar and Parveen, 2020; Wu et al., 2020). In fact, high salinity has been
779 previously observed in sector 3 (Bora; Thomaz and Padial, 2020), however, a profile of the
780 salt wedge with measurements considering tides and seasonal variations is still necessary to
781 establish the salinity value in this sector.

782

783 3.6.2 Chemical Analysis

784 3.6.2.1 Pesticides and trace elements in water

785 The results may indicate that agriculture around of the Guaraguaçu River, as shown
786 and stated by the Ecological Economic Zoning of Paraná (2016), have poor impact in the
787 water quality. Indeed, it is historically mostly of family/subsistence origin, with the main
788 crops being banana, cassava, beans, corn, and rice. However, it is worth noting that even

789 though no pesticides were detected using the method employed, this does not necessarily
790 indicate the absence of pesticides in the region.

791 Analyzing the presence of trace elements on water, manganese, iron and aluminum
792 were found. Two of them, Fe and Al, exceeding the legal limits and the intrinsic
793 characteristics of the geomorphology and pedology of the studied region should take into
794 account. The influence of the rainy collection period can contribute to the increase of these
795 essential trace elements due to weathering of rocks and leaching of the surrounding soils into
796 the river. However, it is worth noting that previous works have shown that even in the
797 sediment matrix there is no significant presence of trace elements in the region of the
798 Guaraguaçu River. In Angeli et al. (2020), when analyzing 135 sediment sample surfaces in a
799 wide region of the Paranaguá Bay estuarine complex, where the Guaraguaçu River flows, it
800 was also shown that there is no strong influence of trace elements. Samples collected at the
801 mouth of the Guaraguaçu River did not demonstrate significant contamination by elements
802 such as Cr, Cu, Ni, and Pb.

803 Despite that, in Cavallini et al. (2018), when analyzing fecal samples of *Lontra*
804 *longicaudis* (OLFERS, 1818), a top predator in the Guaraguaçu River, lead concentrations
805 around 1 mg.L⁻¹ were found, indicating abnormalities. This reinforces the idea of the
806 possibility of trophic biomagnification of trace elements in the biota. Concentrations of
807 cadmium, manganese, and lead were also determined in Neotropical otter feces (Cavallini;
808 Reis and Tiepolo, 2020), which are considered potentially toxic and bioaccumulative metal
809 ions. This contamination may be related primarily to the proximity of the Guaraguaçu River
810 mouth to the port of Paranaguá, as well as the historical and rapid development of the Paraná
811 coast.

812 Therefore, even if trace elements have not been quantified or have low concentration
813 values in the water matrix, it does not necessarily indicate that the region is not influenced by
814 these elements. Species such as *H. malabaricus*, as well as the Neotropical otter used in some
815 studies, are top predator species that can undergo biomagnification processes, leading to
816 harmful effects on their health. This is particularly concerning as these animals are commonly
817 consumed by humans, as demonstrated in the study by Leite et al. (2021), which investigated
818 the bioconcentration of trace elements in the muscle tissue in two Neotropical rivers in Brazil,
819 providing evidence of biomagnification along the food chain.

820

821

822

823 3.6.2.2. Trace elements in the muscle of *H. malabaricus*

824 Trace elements, especially non-essential ones, are mostly considered elements with
825 significant carcinogenic, mutagenic, teratogenic, nephrotoxic, immunotoxic, and neurotoxic
826 potential, capable of disrupting enzymatic proteins in the bodies of living organisms, such as
827 fish (Häder et al., 2020; Nordberg; Nordberg, 2022). Therefore, the results suggested that the
828 Guaraguaçu River is not experiencing concerning levels of pollution when considering
829 contamination by non-essential elements such as Pb, Cd, and Ni, as they were not quantified
830 in the fish muscle tissue. However, the non-essential element Cr was found in three
831 individuals (two in sector 2 and one in sector 3) with concentrations above the legal limits.
832 Chromium can be naturally found in rocks, soil, and living organisms, but its concentration
833 tends to increase in aquatic environments due to certain human activities such as industrial
834 processes and wastewater discharge, causing serious health problems in organisms (Jarapala;
835 Kandlakunta; Thingnganing, 2014), including oxidative stress, DNA damage, cell apoptosis,
836 and alterations in gene expression (Leite et al., 2021).

837 In addition to non-essential trace elements, there are those considered essential for life,
838 which are major components of enzymes, hormones, and animal body cells (Qu et al., 2014).
839 However, if there is an excessive concentration of these essential elements such as Zn, Fe, Cu,
840 and Mn in the water body, sediment, or other matrices, they can cause serious health problems
841 in organisms.

842 Manganese plays an important role as a cofactor of various enzymes, being an
843 essential element found in various sources such as food, soil, and water (Li; Yang, 2018).
844 Zinc is associated with the maintenance of the immune system, body growth, cell division,
845 DNA synthesis, cellular metabolism, reproduction, and participation in protein and cell
846 membrane structure (Garai et al., 2021). It is also considered to have a protective effect
847 against the toxicity of cadmium and lead (Kumar et al., 2021). Similarly, iron (Fe) plays an
848 important role in the growth and development of organisms, being essential for cellular
849 metabolism. However, excessive iron can cause tissue damage (Ayhan; Yaman, 2022), while
850 extremely high concentrations of zinc can decrease immune function (Javed; Usmani, 2016).
851 Zinc toxicity is species-specific and varies with different stages of fish development,
852 environmental factors, and the concentration of the element in the environment. It can result
853 in gill tissue destruction, alterations in swimming behavior, respiratory problems leading to
854 cardiac failure, and fish mortality (Skidmore, 1964).

855 In Kumar et al. (2021), a study conducted in three fishing locations in Mumbai, India,
856 which are influenced by boat traffic and domestic and industrial effluents, bioaccumulation of
857 metals was observed in thirty fish species. Similar to the current study, zinc (Zn) was the most
858 found element in fish. However, it is noted that the concentration of Zn in the present study is
859 much higher than the concentrations reported in Kumar et al. (2021), which consequently
860 found chromium (Cr) concentrations above the legal limits set by Brazilian regulations in
861 almost all analyzed species.

862 Therefore, it can be observed that the concentration of a specific trace element in
863 organisms depends on the species being studied, the nature and feeding habits of the fish, the
864 studied environment, external influences on that environment, such as potential sources of
865 contamination, and the sampling period (Tesser; Rocha and Castro, 2021; Leite et al., 2021).
866 Among the sources of trace elements, natural sources can be highlighted, mainly related to
867 geological leaching of rocks and soil erosion caused by water flow, as well as anthropogenic
868 sources, which can include the discharge of domestic and industrial effluents into water
869 bodies, mining activities, and agriculture (Voigt et al., 2015).

870 The intrinsic characteristics of each fish species are strong factors in the accumulation
871 of trace elements. Voigt et al. (2015) observed that high concentrations of Al, Zn, Fe, and Mn
872 in all analyzed tissues can possibly be related to sediment contamination and the life history
873 and interaction of the *Geophagus brasiliensis* species. Similarly, due to *H. malabaricus* being
874 a carnivorous species with an ambush predation strategy, primarily inhabiting lentic
875 environments such as shallow waters near marginal or submerged vegetation during its adult
876 phase (Reis et al., 2017; Paula; Risso and Martinez, 2021; Leite et al., 2021), the influence of
877 sediment can be a factor in the possibility of bioaccumulation of trace elements.

878 The higher concentration of iron observed in the fish from S3, as demonstrated by the
879 ordination (Fig. 2), can be related to the geology and geomorphology of the area. This sector
880 is characterized by the presence of marine terraces, which represent ancient marine levels that
881 have varied over the past six thousand years. These terraces have an erosive surface,
882 characterized by dark brown coloration caused by the enrichment of organic matter and iron
883 hydroxides. This, in turn, influences the coloration of the Guaraguaçu River waters, which are
884 predominantly transparent but have a black-reddish hue due to both soil erosion and material
885 originating from the forest (IAT, 2006).

886 Among the main soils found in the coastal plain of Paraná, Espodosols and
887 Organossols can be highlighted. Organossols are hydromorphic soils, poorly evolved, and
888 mainly derived from organic matter in different stages of decomposition under permanent

889 water saturation conditions (IAT, 2006). On the other hand, Espodosols are characterized by
890 their sandy texture with the accumulation of organic matter and/or iron oxides. They are also
891 moderately to extremely acidic soils and can have high levels of extractable aluminum
892 (Zaroni; Santos, 2021).

893 Espodosols mainly occur in flat terrain and are therefore found only in the coastal
894 plain of Paraná. Due to the large amount of sand and high permeability, these soils are highly
895 unsuitable for agricultural use (Silva et al., 2013), which also explains the low agricultural
896 influence in the region, with a focus on family farming. Thus, the concentration of organic
897 matter, iron oxides, and the acidic pH found in the waters of sector 1 of the Guaraguaçu
898 River, possibly related to the zinc concentration, may be associated with the intrinsic
899 characteristics of the soils in the region.

900 Another factor that could explain the concentration of Mn in almost all analyzed
901 individuals, as well as Fe and Zn in all organisms is the fact that these trace elements are
902 essential. The presence of these elements in a larger number of organisms and in considerable
903 concentrations could be another explanation for the possibility of bioaccumulation, as shown
904 in various studies with different fish species in environments experiencing similar
905 anthropogenic interference to the Guaraguaçu River (Kamaruzzaman et al., 2011; Jesus et al.,
906 2014; Jarapala; Kandlakunta; Thingnganing, 2014; Javed; Usmani, 2016). The sampling
907 period can also affect the bioavailability of these elements in the environment. For example,
908 there may be a higher concentration of trace elements, such as aluminum, during rainy periods
909 (Leite et al., 2021), due to the increased input of sediments and effluents carried by rainfall.

910 The presence of aluminum (Al) occurring almost exclusively in the muscle tissues of
911 fish from S1 and S3, characterized as pristine and minimally impacted, respectively, suggests
912 that the concentration be derived from natural sources. Aluminum in natural waters originates
913 from weathering of rocks and minerals, and there are no specific regulatory limits for its
914 concentration in fish. However, it is important to investigate the influence of aluminum on the
915 health of fish in the Guaraguaçu River, as this element can be neurotoxic and can cause
916 respiratory and reproductive diseases to animals (Gemsemer et al., 2018).

917

918 3.6.3. Biochemical biomarkers

919 The decrease in acetylcholinesterase activity in S2 may be related to the presence of a
920 neurotoxic substance with anticholinesterase effects, potentially impairing motor coordination
921 and locomotion of organisms (Oliveira et al., 2019). The inhibition of AChE activity is widely

922 known as an exposure biomarker for organophosphate and carbamate pesticides (Fajardo and
923 Ocampo, 2018). However, diverse contaminants have also been classified as inhibitors of
924 AChE activity, including metals and other classes of organic environmental pollutants (Fu et
925 al., 2018).

926 The gills are the main and often the first route of contact with xenobiotics in addition
927 to being a crucial organ for respiration and osmoregulation in fish (Kumar et al., 2017). GST
928 is one of the enzymes involved in the biotransformation of xenobiotics into more hydrophilic
929 metabolites for subsequent excretion (Kroon; Streten; Harries, 2017), which also plays a role
930 in biological stress control due to abiotic factors. The increase in GST activity in S3 can be
931 directly related to the presence of salinity, as this sector is known to be influenced by tides.
932 This change in water salinity can cause osmotic stress in aquatic animals such as fish (Evans;
933 Kultz, 2020). The presence of salinity may also explain the decrease in catalase activity in the
934 gills of S3. Mozanzadeh et al. (2021) demonstrated in their study that an increase in salinity
935 could reduce catalase activity in certain fish tissues, such as the liver. This suggests that the
936 oxidative stress response of fish to changes in water salinity are related to the specificity of
937 each species, as well as the developmental stage of the organism and the concentration of salt
938 in the water.

939 Both xenobiotics and environmental factors such as salinity, temperature, and pH can
940 cause oxidative stress by producing reactive oxygen species (ROS), which can be neutralized
941 by antioxidant system (SOD, CAT, GPx, and GST) as well as GSH (Chowdhury; Saikia,
942 2020; García-Caparrós et al., 2021). The increase in SOD and GPx activity may be related to
943 the presence of elevated concentrations of ROS, especially in S2, which is heavily influenced
944 by anthropogenic activities. Despite the increased activity of antioxidant enzymes, the
945 lipoperoxidation also increased, which can indicate the presence of concerning concentrations
946 of certain xenobiotics.

947 Liver is commonly used for biomarker analysis because it is one of the main organs
948 responsible for the metabolism and detoxification of xenobiotics. The cofactor reduced
949 glutathione plays a role in binding to GST and GPx enzymes to detoxify the body from
950 xenobiotics and combat reactive oxygen species, respectively (Burkina; Zlabek and
951 Zamaratskaia, 2015; Kroon; Streten; Harries, 2017). The increased of GST, GPx and SOD
952 activities, and concentration of GSH and LPO demonstrate that Sector 2 indeed be the most
953 impacted by anthropogenic actions, leading to changes in enzymatic activities and increased

954 cases of lipoperoxidation, which are damages to the lipid membrane, indicating tissue damage
955 in the fish.

956 Metallothionein plays a crucial role in immune response by participating in the
957 detoxification and transport of metals within the bodies of various animal groups, including
958 vertebrates and invertebrates. El-Khayat et al. (2020) observed a strong positive correlation
959 between metallothionein activity and the presence of metals such as cadmium, lead, and
960 copper in various fish tissues, with the liver showing the highest correlation, highlighting the
961 potential of metallothionein as a biomarker in toxicological studies and for assessing
962 environmental stress. However, in the case of the Guaraguaçu River, no significant difference
963 in metallothionein concentration was observed among the three sectors of the river.

964

965 3.6.4. Histopathological biomarkers

966 The use of histopathological analysis provides additional information when
967 combined with various other biomarkers, allowing for a more comprehensive and integrated
968 study in biomonitoring. Among the lesions found in the gills of sector 2, only hyperplasia
969 with partial or total fusion indicating moderate alterations with potential reversibility and
970 are related to an increase in the number of cells, consequently reducing the space between
971 the gill filaments. The presence of hyperplasia, as well as an increase in mucous cells,
972 represents tissue protection mechanisms against pathogens and contaminants from the
973 external environment, however it hinders gas exchange by the organisms, decreasing blood
974 oxygenation (Mallatt, 1985; Marinović et al., 2021).

975 In previous studies focused on aquatic environments contaminated by sewage
976 discharge (Liebel; Tomotake and Oliveira-Ribeiro, 2013; Pereira et al., 2020), the presence
977 of trace elements (Salgado et al., 2021; Oliveira et al., 2022), and pesticides (Oliveira et al.,
978 2019), organisms exhibited similar responses and lesions to those found in the present
979 study. These include epithelial lifting of the secondary lamella found in sector 2, that can
980 happen due to infiltration of fluid between epithelium and basement membrane that
981 ultimately increase diffusion distance for gas exchange; aneurysms, found principally in
982 gills of sector 2, are related to blood accumulation and dilation of the branchial artery
983 (Flores-Lopes; Thomaz, 2011); and loss of microridges in pavement cells related to the gills
984 of sector 2 and 3, where this structure which primarily serve to increase the surface area of
985 cells in contact with the external environment and also act as structures capable of retaining
986 mucus to protect the entire gill structure (Mela et al., 2013). Blood congestion, can leads to

987 the disruption of pillar cells due to the intense blood flow to the secondary lamellae, can
988 cause aneurysms (Hassaninezhad et al., 2014).

989 Freitas et al. (2022) studied histopathological biomarkers in *H. malabaricus* in the
990 Mearim River, located in the Brazilian Amazon, which is affected by contamination from
991 anthropogenic activities. The fish in this region also exhibited biological responses to
992 contaminants, including congestion, aneurysms, hyperplasia with partial or total fusion of
993 lamellae, mucus proliferation, and epithelial lifting, as well as the fish captured in sector 2
994 of the present work. All these alterations indicate a strong influence of anthropogenic
995 actions and water degradation due to the presence of contaminants in the study area,
996 suggesting that gill alterations do not have a specific relationship with a particular
997 xenobiotic, as similar responses can be observed for different contaminants. According to
998 Oliveira et al., 2022, the gill lesions reflect a generalized stress response.

999 Another important aspect to highlight is the influence of seasonality on the
1000 responses of biomarkers to xenobiotics (Salgado et al., 2019; Marinović et al., 2021). The
1001 concentration of water contaminants can vary with the water flow and the influence of a
1002 wetter or drier period. Salgado et al. (2021) found higher responses of histopathological
1003 biomarkers in fish collected during the cold-dry period, which may be related to lower
1004 precipitation and, consequently, lower water flow, facilitating higher concentrations of
1005 contaminants in the water or even in sediments. However, Pereira et al. (2020), when
1006 evaluating environmental contamination of a river in northeastern Brazil through the
1007 analysis of histopathological biomarkers in the gills of *Psectrogaster amazônica*, found that
1008 the indexes of histological alterations were higher during the rainy season, which can be
1009 explained by the greater input of contaminants through surface runoff. Therefore, it is
1010 crucial to conduct a comprehensive study on the influence of seasonality on organism
1011 responses to contaminants, as the results can vary from region to region and, primarily, due
1012 to the biology of the individuals.

1013 Among the liver alterations and severe lesions, necrosis or cell death, and hepatocyte
1014 vacuolization can be highlighted in this study. Vacuolation, found in liver fish of sector 2,
1015 indicate decrease of stored energy in the form of glycogen or represent a degenerative
1016 change in which there is fluid distension of organelles such as endoplasmic reticulum and
1017 Golgi apparatus (Mela et al., 2007). Increased hepatocyte vacuolation, is often cited as a
1018 toxicological response in fish, although the exact composition of the vacuolation and
1019 mechanism of formation are frequently not elucidated. According to Gonzales et al. (1993),
1020 necrosis found in the fish liver are usually related to contaminants found in water or

1021 sediment. The principle of hepatic necrosis results from the presence of chemicals within
1022 cells causing disturbs on biochemical process as enzyme inhibition, failure on protein
1023 synthesis, carbohydrate metabolism, reactive oxidative species production, damages in cell
1024 membrane and failure of ATP synthesis (Mela et al., 2013; Kumar et al., 2017). These two
1025 irreversible lesions, found in fish's liver of sectors 2 and 3, demonstrate and indicate the
1026 presence of contaminants capable of altering the health of organisms.

1027 Melanomacrophage centers is a collection of macrophages that contain hemosiderin,
1028 lipofuscin, and seroids as well as the melanin pigment caused by inflammation of most
1029 teleost (Rabitto et al., 2005). These structures increase in size or frequency in conditions of
1030 environmental stress and have been suggested as reliable biomarkers for water quality in
1031 terms of both deoxygenation and anthropogenic chemical pollution (Mela et al., 2007,
1032 2013). The presence of melanomacrophage centers, can be associated with regions with
1033 history of contamination (Viana et al., 2021), influenced for example by trace elements as
1034 copper (Mela et al., 2013). These trace elements can affect the normal functioning of the
1035 liver and be responsible for hepatic insufficiency (Savassi et al., 2020). Blood congestion is
1036 a liver dysfunction due to venous congestion, usually as a result of dysfunction of the heart,
1037 which is also known as congestive heart failure (Cotran et al., 2005). The fish liver is
1038 especially liable to chemical products due to the slow blood flow in relation to the cardiac
1039 output. Liver and gills alterations, such as those observed in this study, could result in
1040 severe physiological problems and provide reliable information on stress to a broad range of
1041 environmental pollutants.

1042

1043 3.6.5. Biomarker of genotoxicity

1044 The frequencies of micronuclei generally vary according to the season of the year, the
1045 type of contaminant involved, and the fish species under evaluation (Ali; El-Shehawi; Seehy,
1046 2008; Obiakor; Okonkwo; Ezeonyejiaku, 2012). It is important to highlight the presence of
1047 micronuclei in at least one fish from each sector, which emphasizes the possibility of water
1048 contamination in this river, particularly related to anthropogenic activities (Canedo et al.,
1049 2021) such as domestic sewage discharge and the presence of trace elements (Hussain et al.,
1050 2018; Ali et al., 2020). The presence of these impacts increasingly plausible since previous
1051 studies (Francisco et al., 2019; Lehun et al., 2021) have found micronuclei in organisms
1052 living in regions impacted by practically the same probably causes found in the Guaraguaçu
1053 River. These alterations would rarely occur naturally, as they are considered mutagenic

1054 characteristics, as the increase in frequency is usually influenced by exposure to clastogenic
1055 or aneugenic substances that are closely linked to environmental disturbances (Hayashi, 2016;
1056 Ali et al., 2020).

1057 Salgado et al., 2019 reported the lack of micronuclei or vacuolated alterations, but
1058 blebbed, notched, and lobed nuclei were found. However, Oliveira et al., 2019 observed all
1059 the alterations described in the present study, indicating that the variation in the presence or
1060 absence of nuclear morphological alterations also depends on the contamination history of the
1061 study site and the biomonitor species used, as each organism has different biology. This was
1062 demonstrated in Rodriguez-Cea et al. (2003) using different fish species, where only brown
1063 trout showed potential as a biomonitor species by demonstrating higher sensitivity to
1064 genotoxic compounds such as the trace element cadmium when comparing contaminated and
1065 non-contaminated regions. Indeed, in addition to the characteristics of the study area and the
1066 biological model, one of the main reasons that makes it challenging to use nuclear alterations
1067 in erythrocytes as biomarkers to interpret genotoxic damage is the distinct origins of
1068 micronucleus formation and other nuclear abnormalities (Krupina; Goginashvili; Cleveland,
1069 2021).

1070

1071 3.7. CONCLUSION

1072 In the water samples, pesticides were not detected, just manganese, iron and aluminum
1073 were quantified, indicating a low concentration of trace elements. However, in the muscles of
1074 the fish some trace elements were detected. This can evidence that the study with a top
1075 predator is interesting, because it can analyze the accumulation of xenobiotics over time, since
1076 these animals can consume other living organisms. The use of biomarkers may be more
1077 sensitive than chemical techniques, because, although the presence of pesticides and some
1078 metals in water was not evidenced, prominent responses of biomarkers were presented.
1079 Severe damage was analyzed mainly in sector 2 with a certain level of human impact, such as
1080 neurotoxicity, lipoperoxidation, histopathological damage in the liver and gills and blood
1081 mutagenicity. These results support the hypothesis proposed in this study and create a
1082 valuable dataset that can inform the problems for the social-vulnerable residents who use the
1083 river for survival, as well as for decision-making by policymakers.

1084

1085

1086

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1097

1098 3.9. REFERENCES

1099

1100 ABREU-MOTA, M. A. et al. Sedimentary biomarkers along a contamination gradient in a
1101 human-impacted sub-estuary in Southern Brazil: A multi-parameter approach based on
1102 spatial and seasonal variability. **Chemosphere**, v.103, p.156–163, 2014.

1103

1104 AEBI, H. Catalase in vitro. **Methods in Enzymology**, v.105, p.121-126, 1984.

1105

1106 ALI, D. et al. Fish as bio indicators to determine the effects of pollution in river by using
1107 the micronucleus and alkaline single cell gel electrophoresis assay. **Journal of King Saud
1108 University - Science**, v. 32, p. 2880–2885, 2020.

1109

1110 ALI, F. K.; EL-SHEHAWI, A. M.; SEEHY, M. A. Micronucleus test in fish genome: A
1111 sensitive monitor for aquatic pollution. **African Journal of Biotechnology**, v. 7, n. 5, p.
1112 606–612, 2008.

1113

1114 ANGELI, J.L.F. et al. Statistical assessment of background levels for metal contamination
1115 from a subtropical estuarine system in the SW Atlantic (Paranaguá Estuarine System,
1116 Brazil). **Journal of Sedimentary Environments**, v.5, p.137–150, 2020

- 1117 ARAÚJO, E. S.; VITULE, J. R. S.; PADIAL, A. A. A. Checklist of aquatic macrophytes
1118 of the Guaraguaçu river basin reveals a target for conservation in the Atlantic rainforest.
1119 **Acta Scientiarum. Biological Sciences**, v. 43, n.1, e50542, 2021.
1120
- 1121 ARIF, M.; KUMAR, R.; PARVEEN, S. Reduction in Water Pollution in Yamuna River
1122 Due to Lockdown Under COVID-19 Pandemic. ChemRxiv. Cambridge: Cambridge Open
1123 Engage; This content is a preprint and has not been peer-reviewed, 2020.
1124
- 1125 AYHAN, N.; YAMAN, M. Evaluation of Iron and Zinc Contents of Some Fish Species.
1126 **Biological Trace Element Research**, v. 200, p.1376–1382, 2022.
1127
- 1128 BARHOUMI, S. et al. Spatial and seasonal variability of some biomarkers in *Salaria*
1129 *basilisca* (Pisces: Blennidae): Implication for biomonitoring in Tunisian coasts.
1130 **Ecological Indicators**, v.14, p.222–228, 2012.
1131
- 1132 BERNET, D. et al. Histopathology in fish: proposal for a protocol to assess aquatic
1133 pollution. **Journal of Fish Diseases**, v. 22, n. 1, p. 25–34, 1999.
1134
- 1135 BORA, L.S., THOMAZ, S.M., PADIAL, A.A. Evidence of rapid evolution of an invasive
1136 *poaceae* in response to salinity. **Aquatic Sciences**, v. 82, n.76, 2020.
1137
- 1138 BRADFORD, M. A rapid and sensitive method for the quantitation of microgram
1139 quantities of protein utilizing the principle of protein-dye binding. **Analytical**
1140 **Biochemistry**, v. 72, n. 1, p. 248- 254, 1976.
1141
- 1142 BRASIL. Decreto nº. 55871 de 26 de março de 1965. Modifica o Decreto nº 50.040, de 24
1143 de janeiro de 1961, referente a normas reguladoras do emprego de aditivos para alimentos,
1144 alterado pelo Decreto nº 691, de 13 de março de 1962. Diário Oficial da União, Poder
1145 Executivo, Brasília, 1965.
1146
- 1147 BRASIL. Ministério da Saúde. Agência Nacional de Vigilância Sanitária. Resolução -
1148 RDC n. 42 de 29 de agosto de 2013. Dispõe sobre o Regulamento Técnico MERCOSUL
1149 sobre Limites Máximos de Contaminantes Inorgânicos em Alimentos. Diário Oficial da
1150 República Federativa do Brasil, Brasília, 168, 33, Seção 1, 2013.

- 1151 BURKINA, V., ZLABEK, V., ZAMARATSKAIA, G. Effects of pharmaceuticals present
1152 in aquatic environment on Phase I metabolism in fish. **Environmental Toxicology and**
1153 **Pharmacology**, v. 40, p. 430–444, 2015
1154
- 1155 CALADO, S.L.M. et al. Biochemical and genotoxicity assessment of a polluted urban
1156 river using the native fish *Astyanax altiparanae* Garutti & Britski (Teleostei, Characidae).
1157 **Ecotoxicology and Environmental Contamination**, v. 14, n.1, p. 73-77, 2019.
1158
- 1159 CANEDO, A. et al. Micronucleus test and nuclear abnormality assay in zebrafish (*Danio*
1160 *rerio*): Past, present, and future trends. **Environmental Pollution**, v.290, 2021.
1161
- 1162 CARRASCO, K. R., TILBURY, K. L.; MYERS, M. S. Assessment of the piscine
1163 micronucleus test as an in situ biological indicator of chemical contaminant effects.
1164 **Canadian Journal of Fisheries and Aquatic Sciences**, v. 47, p. 2123-2136, 1990.
1165
- 1166 CAVALLINI, N.G. et al. Determination of Lead (Pb) in stools of *Lontra longicaudis*
1167 (Olfers, 1818) by flame atomic absorption spectrometry (FAAS). **Eclética Química**
1168 **Journal**, v.43, p.70-78, 2018.
1169
- 1170 CAVALLINI, N. G.; REIS, R. A.; TIEPOLO, L. M. O silencioso grito químico: Riscos e
1171 ameaças no rio guaraguaçu sob a perspectiva ecossistêmica / The quiet chemical scream:
1172 Risks and imminent threats in the guaraguaçu river under the ecosystem
1173 system. **Brazilian Journal of Development**, v.6, n.9, p.66540–66553, 2020.
1174
- 1175 CHAUDHRY, F.N.; MALIK, M.F. Factors Affecting Water Pollution: A Review.
1176 **Journal of Ecosystem and Ecography**, v.07, 2017.
1177
- 1178 CHOWDHURY, S.; SAIKIA, S. K. Oxidative Stress in Fish: A Review. **Journal of**
1179 **Scientific Research**, v.12, n.1, p.145–160, 2020.
1180
- 1181 CHU-KOO, F. W.; PÉREZ, A. M. D. BIOLOGÍA Y CULTIVO DEL FASACO *Hoplias*
1182 *malabaricus* Bloch 1794 (CHARACIFORMES: ERYTHRINIDAE). **Folia Amazónica**, v.
1183 16, n. 1–2, p. 11–21, 2007.
1184

- 1185 CONAMA. Resolução CONAMA 357/2005, de 17 de março de 2005. Dispõem sobre a
1186 classificação dos corpos de água e diretrizes ambientais para seu enquadramento, bem
1187 como estabelece as condições e padrões de lançamento de efluentes, e dá outras
1188 providências. Brasília, Ministério do Meio ambiente, 2005.
- 1189
- 1190 CONTENTE, R.F.; STEFANONI, M.F.; SPACH, H.L. Fish assemblage structure in an
1191 estuary of the Atlantic Forest biodiversity hotspot (southern Brazil). **Ichthyological**
1192 **Research**, v. 58, p.38-50, 2011.
- 1193
- 1194 COTRAN, R. S. et al. Pathologic basis of disease. Philadelphia, PA: **Elsevier Saunders**,
1195 2005.
- 1196
- 1197 CRUZ-ESQUIVEL, Á.; MARRUGO-NEGRETE, J. Methylmercury concentrations in
1198 *Prochilodus magdalenae* (Teleostei: Curimatidae) and *Hoplias malabaricus* (Teleostei:
1199 Erythrinidae) in the lower Cauca-Magdalena river system, northern Colombia. **Acta**
1200 **Biologica Colombiana**, v. 27, n. 1, p. 28–35, 1 jan. 2022.
- 1201
- 1202 DUDGEON, D. Multiple threats imperil freshwater biodiversity in the Anthropocene.
1203 **Current Biology**, v. 29, p. 942–995, 2019.
- 1204
- 1205 DUFFUS, J.H. “Heavy Metals”-A Meaningless Term? (IUPAC Technical Report). **Pure**
1206 **and Applied Chemistry**. v.74, p.793–807, 2002.
- 1207
- 1208 EL-KHAYAT, H. M. M. et al. Assessment of metallothionein expression in *Biomphalaria*
1209 *alexandrina* snails and *Oreochromis niloticus* Fish as a biomarker for water pollution with
1210 heavy metals. **Egyptian Journal of Aquatic Biology & Fisheries**, v. 24, n. 2, p. 209–
1211 223, 2020.
- 1212
- 1213 ELLMAN, G.L. et al. A new and rapid colorimetric determination of acetylcholinesterase
1214 activity. **Biochemical Pharmacology**, v.7, p.88– 95, 1961.
- 1215
- 1216 ELSTE, G.A.S. et al. A contaminação do rio Guaraguaçu (Litoral do Paraná):limites e
1217 riscos ao desenvolvimento territorial regional. **Guaju, Matinhos**, v.5, n.2, p. 54-70,
1218 jul./dez. 2019.

- 1219 ESCALANTE-ROJAS, M.C. et al. Integrated use of biomarkers to evaluate the
1220 reproductive physiology of *Astyanax fasciatus* and *Hoplias malabaricus* males (Teleostei:
1221 Characiformes) in polluted reservoirs. **Ecotoxicology and Environmental Safety**, v.208,
1222 2021.
- 1223
- 1224 EUROPEAN COMMUNITY (EC). Commission Regulation of the European Community
1225 (EC) No. 1881/2006, dated December 19, 2006, setting maximum levels for certain
1226 contaminants in foodstuffs. EUR-Lex (L364), p. 5-24, 2006.
- 1227
- 1228 EVANS, T.G.; KÜLTZ, D. The cellular stress response in fish exposed to salinity
1229 fluctuations. **Journal of Experimental Zoology Part A: Ecological and Integrative**
1230 **Physiology**, v.333, n.6, p. 421–435, 2020.
- 1231
- 1232 FAJARDO, L.J., OCAMPO, P.P. Inhibition of acetylcholinesterase activities in
1233 whitegoby, *Glossogobius giuris* from the East Bay of Laguna Lake, Philippines.
1234 **International Journal of Agricultural Technology**, v.14, n.7, p.1181- 1192, 2018.
- 1235
- 1236 FLORES-LOPES, F.; THOMAZ, A.T. Histopathologic alterations observed in fish gills
1237 as a tool in environmental monitoring. **Brazilian Journal of Biology**, 2011.
- 1238
- 1239 FOOD AND AGRICULTURE ORGANIZATION (FAO). Compilation of legal limits for
1240 hazardous substances in fish and fishery products. **FAO Fisheries Circular**, v.464, p.5–
1241 100, 1983.
- 1242
- 1243 FRANCISCO, C.D.M. et al. Genotoxicity assessment of polluted urban streams using a
1244 native fish *Astyanax altiparanae*. **Journal of Toxicology and Environmental Health,**
1245 **Part A**, v. 82, p. 514–523, 2019.
- 1246
- 1247 FREITAS, L.C. et al. Histological biomarkers and biometric data on trahira *Hoplias*
1248 *malabaricus* (Pisces, Characiformes, Erythrinidae): a bioindicator species in the Mearim
1249 river, Brazilian Amazon. **Brazilian Journal of Biology**, v. 82, e263047, 2022.
- 1250

- 1251 FU, H. et al. Acetylcholinesterase Is a Potential Biomarker for a Broad Spectrum of
1252 Organic Environmental Pollutants. **Environmental Science & Technology**, v.52, n.15,
1253 p.8065–8074, 2018.
1254
- 1255 GALVANESE, E. F. et al. Community stability and seasonal biotic homogenisation
1256 emphasize the effect of the invasive tropical tanner grass on macrophytes from a highly
1257 dynamic neotropical tidal river. **Aquatic Sciences**, v. 84, n. 2, 1 abr. 2022.
1258
- 1259 GAO, R. et al. Mechanism of pyrogallol autoxidation and determination of superoxide
1260 dismutase enzyme activity. **Bioelectrochemistry and Bioenergetics**, v.45, n.1, p.41-45,
1261 1998.
1262
- 1263 GARAI, P. et al. Effect of Heavy Metals on Fishes: Toxicity and Bioaccumulation. **The**
1264 **Journal of Clinical Toxicology** , v.11, 2021.
1265
- 1266 GARCÍA-CAPARRÓS, P. et al. Oxidative Stress and Antioxidant Metabolism under
1267 Adverse Environmental Conditions: a Review. **Botanical Review**, v. 87, p. 421–466,
1268 2021.
1269
- 1270 GAZOLA-SILVA, F. F. et al. Chthonerpeton viviparum Parker & Wettstein,
1271 1929(Amphibia, Gymnophiona, Typhlonectinae) in Paraná state, Brazil and the first record
1272 of predation of this species by *Hoplias malabaricus* (Bloch, 1794) (Actinopterygii,
1273 Erythrinidae). **Pan-American Journal of Aquatic Sciences** , v.2, n.3, p. 261-262, 2007.
1274
- 1275 GEMSEMER, R.W. et al. Evaluating the effects of pH, hardness, and dissolved organic
1276 carbon on the toxicity of aluminum to freshwater aquatic organisms under circumneutral
1277 conditions. **Environmental Toxicology and Chemistry**, v. 37, p. 49-60, 2018.
1278
- 1279 GEORGIEVA, E. et al. A Review on Multi-Biomarkers in Fish for the Assessment of
1280 Aquatic Ecosystem Contamination with Organic Pollutants. **Ecologia Balkanica**, v.13,
1281 p.321–330, 2021.
1282
1283

- 1284 GOMES, A. D. et al. The role of ovarian steroids in reproductive plasticity in *Hoplias*
1285 *malabaricus* (Teleostei: Characiformes: Erythrinidae) in tropical reservoirs with different
1286 degrees of pollution. *General and Comparative Endocrinology*, v. 222, p. 1–10, 1 out.
1287 2015.
- 1288
- 1289 GONZALES, G.; CRESPO, S.; BRUSLE, J. Histocytological study of the liver of the
1290 cabrilla sea bass, *Serranus cabrilla* (Teleostei, Serranidae), an available model for
1291 marine fish experimental studies. ***Journal of Fish Biology***, v.43, p.363-73, 1993.
- 1292
- 1293 GRASSI, D.J. et al. Cytogenetic characterization of *Hoplias malabaricus* (Bloch, 1794)
1294 from the Ctalamochita River (Córdoba, Argentina): First evidence for southernmost
1295 populations of this species complex and comments on its biogeography. ***Comparative***
1296 ***Cytogenetics***, v. 11, p.15–28, 2017.
- 1297
- 1298 HÄDER, D.P. et al. Anthropogenic pollution of aquatic ecosystems: emerging problems
1299 with global implications. ***Science of the Total Environment***. v. 713, 2020.
- 1300
- 1301 HAFEMAN, D. G.; SUNDE, R.A.; HOEKSTRA, W.C. Effect of dietary selenium on
1302 erythrocyte and liver glutathione peroxidase in the rat. ***Journal of Nutrition***, v.104, n.4,
1303 p.580-587, 1974.
- 1304
- 1305 HASHIMI, M.H.; HASHIMI, R.; RYAN, Q. Toxic Effects of Pesticides on Humans,
1306 Plants, Animals, Pollinators and Beneficial Organisms. ***Asian Plant Research Journal***,
1307 v. 5, p. 37–47, 2020.
- 1308
- 1309 HASSANINEZHAD, L. et al. Assessment of gill pathological responses in the tropical
1310 fish yellowfin seabream of Persian Gulf under mercury exposure. ***Toxicology Reports***,
1311 v.1, p.621-628, 2014.
- 1312
- 1313 HAYASHI, M. The micronucleus test-most widely used in vivo genotoxicity test. ***Genes***
1314 ***and Environment***, 2016.
- 1315

- 1316 HOOFTMAN, R.N.; RAAT, W.K. Induction of nuclear anomalies (micronuclei) in
1317 peripheral blood erythrocytes of Eastern mudminnow *Umbra pygmaea* by ethyl
1318 methanesulphonate, **Mutation Research**, v.104, p.147-152, 1982.
1319
- 1320 HUSSAIN, B. et al. Fish eco-genotoxicology: Comet and micronucleus assay in fish
1321 erythrocytes as in situ biomarker of freshwater pollution. **Saudi Journal of Biological
1322 Sciences**, v.25, p.393–398, 2018.
1323
- 1324 INSTITUTO ÁGUA E TERRA (IAT). Encarte II: Análise Regional da EEG. Brasil. 2006.
1325 Disponível em: <[https://www.iat.pr.gov.br/Pagina/Plano-de-Manejo-Estacao-Ecologica-
1326 do-Guaraguacu](https://www.iat.pr.gov.br/Pagina/Plano-de-Manejo-Estacao-Ecologica-do-Guaraguacu)>. Accessed on November 12, 2022.
1327
- 1328 JARAPALA, S.R., KANDLAKUNTA, B., THINGNGANING, L. Evaluation of trace
1329 metal content by ICP-MS using closed vessel microwave digestion in freshwater fish.
1330 **Journal of Environmental and Public Health**, 2014.
1331
- 1332 JAVED, M.; USMANI, N. Accumulation of heavy metals and human health risk
1333 assessment via the consumption of freshwater fish *Mastacembelus armatus* inhabiting,
1334 thermal power plant effluent loaded canal. **Springerplus**, v. 5, 2016.
1335
- 1336 JESUS, I.S. et al. Analysis of metal contamination and bioindicator potential of predatory
1337 fish species along Contas River basin in northeastern Brazil. **Bulletin of Environmental
1338 Contamination and Toxicology**, v.92, p.551-556, 2014.
1339
- 1340 JIANG, Z.Y., HUNT, J.V., WOLFF, S.P. Ferrous ion oxidation in the presence of xylenol
1341 orange for detection of lipid hydroperoxide in low density lipoprotein. **Analytical
1342 Biochemistry**, v.202, n.2, p.384-389, 1992.
1343
- 1344 KAMARUZZAMAN, B.Y. et al. Heavy Metal Accumulation in Commercially Important
1345 Fishes of South West Malaysian Coast. **Research Journal of Environmental Sciences**,
1346 2011.
1347

- 1348 KEEN, J. H.; HABIG, W. H.; JAKOBY, W. B. Mechanism for the several activities of the
1349 glutathione S-transferases. **Journal of Biological Chemistry**, v.251, n.20, p.6183-6188,
1350 1976.
- 1351
- 1352 KHOSHNOOD, Z. Effects of Environmental Pollution on Fish: A Short Review.
1353 **Transylvanian Review of Systematical and Ecological Research**, v.19, n.1, p.49-60,
1354 2017.
- 1355
- 1356 KROON, F.; STRETEN, C.; HARRIES, S. A protocol for identifying suitable biomarkers
1357 to assess fish health: A systematic review. **PloS one**, v. 12, n. 4, p. e0174762, 2017.
- 1358
- 1359 KRUPINA, K., GOGINASHVILI, A., CLEVELAND, D.W. Causes and consequences of
1360 micronuclei. **Current Opinion in Cell Biology**, 2021.
- 1361
- 1362 KUMAR, N. et al. Cellular stress and histopathological tools used as biomarkers in
1363 *Oreochromis mossambicus* for assessing metal contamination. **Environmental**
1364 **Toxicology and Pharmacology**, v.49, p.137-147, 2017.
- 1365
- 1366 KUMAR, N. et al. Metal determination and biochemical status of marine fishes facilitate
1367 the biomonitoring of marine pollution. **Marine Pollution Bulletin**, v.170, 2021.
- 1368
- 1369 LEÃO-BUCHIR, J. et al. BDE-99 (2,2-,4,4-,5 - pentain polybrominated diphenyl ether)
1370 induces toxic effects in *Oreochromis niloticus* after sub-chronic and oral exposure.
1371 **Environmental Toxicology and Pharmacology**, v.97, 104034, 2023.
- 1372
- 1373 LEHUN, A.L. et al. Genotoxic effects of urban pollution in the Iguaçu River on two fish
1374 populations. **Journal of Environmental Science and Health, Part A. Toxic/hazardous**
1375 **substances and environmental engineering**, v.56, p.984–991, 2021
- 1376
- 1377 LEITE, L. A. R. et al. Bioaccumulation and Health Risk Assessment of Trace Metal
1378 Contamination in the Musculature of the Trahira Fish (*Hoplias Malabaricus*) from Two
1379 Neotropical Rivers in Southeastern Brazil. **SSRN Electronic Journal**, 16 fev. 2021.
- 1380

- 1381 LI, L.; YANG, X. The essential element manganese, oxidative stress, and metabolic
1382 diseases: Links and interactions. **Oxidative Medicine and Cellular Longevity**, 2018.
- 1383
- 1384 LIEBEL, S., TOMOTAKE, M.E.M., OLIVEIRA-RIBEIRO, C.A. Fish histopathology as
1385 biomarker to evaluate water quality. **Ecotoxicology and Environmental Contamination**,
1386 v.8, p.9–15, 2013.
- 1387
- 1388 LOZANO, I. E. et al. Comparison of scale and otolith age readings for trahira, *Hoplias*
1389 *malabaricus* (Bloch, 1794), from Paraná River, Argentina. **Journal of Applied**
1390 **Ichthyology**, v. 30, n. 1, p. 130–134, fev. 2013.
- 1391
- 1392 MALIK, D.S. et al. A review on impact of water pollution on freshwater fish species and
1393 their aquatic environment. In: **Advances in Environmental Pollution Management:**
1394 **Wastewater Impacts and Treatment Technologies. Agro Environ Media -**
1395 **Agriculture and Environmental Science Academy**, Haridwar, India, pp. 10–28, 2020.
- 1396
- 1397 MALLATT, J. Fish gill structural changes induced by toxicants and other irritants: A
1398 statistical review. **Canadian Journal of Fisheries and Aquatic Sciences**, v.42, p.630–
1399 648, 1985.
- 1400
- 1401 MARINOVIĆ, Z. et al. Gill histopathology as a biomarker for discriminating seasonal
1402 variations in water quality. **Applied Sciences**, v.11, 2021.
- 1403
- 1404 MELA, M. et al. Effects of dietary methylmercury on liver and kidney histology in the
1405 neotropical fish *Hoplias malabaricus*. **Ecotoxicology and Environmental Safety**, v.68,
1406 n.3, p.426–435, 2007.
- 1407
- 1408 MELA, M. et al. Mercury distribution in target organs and biochemical responses after
1409 subchronic and trophic exposure to Neotropical fish *Hoplias malabaricus*. **Fish**
1410 **Physiology and Biochemistry**, v.40, p.245–256, 2014.
- 1411
- 1412 MELA, M. et al. Risks of waterborne copper exposure to a cultivated freshwater
1413 Neotropical catfish (*Rhamdia quelen*). **Ecotoxicology and Environmental Safety**, v. 88,
1414 p. 108–116, 1 fev. 2013.

- 1415 MONTENEGRO, A. K. A. et al. Piscivory by *Hoplias aff. malabaricus* (Bloch, 1794): A
1416 question of prey availability? **Acta Limnologica Brasiliensia**, v. 25, n. 1, p. 68–78, 2013.
1417
- 1418 MONTES, C. et al. Evaluation of metal contamination effects in piranhas through
1419 biomonitoring and multi biomarkers approach. **Heliyon**, v.6, 2020.
1420
- 1421 MOZANZADEH, M. T. et al. The effect of salinity on growth performance, digestive and
1422 antioxidant enzymes, humoral immunity and stress indices in two euryhaline fish species:
1423 Yellowfin seabream (*Acanthopagrus latus*) and Asian seabass (*Lates
1424 calcarifer*). **Aquaculture**, v.534, 736329, 2021.
1425
- 1426 MYERS, N. et al. Biodiversity hotspots for conservation priorities. **Nature**, v.403, p.853–
1427 858, 2000.
1428
- 1429 NORDBERG, M.; NORDBERG, G. F. Metallothionein and Cadmium Toxicology—
1430 Historical Review and Commentary. **Biomolecules**, v. 12, n.360, 2022.
1431
- 1432 OBIAKOR, M. O.; OKONKWO, J. C.; EZEONYEJAKU, C. D. Eco-genotoxicology:
1433 Micronucleus Assay in Fish Erythrocytes as in situ Aquatic Pollution Biomarker: a
1434 Review. **Journal of Animal Science Advances** , v. 2, n. 1, p. 123–133, 2012.
1435
- 1436 OLIVEIRA, F. G. et al. Toxicological effects of anthropogenic activities in *Geophagus
1437 brasiliensis* from a coastal river of southern Brazil: A biomarker approach. **Science of the
1438 Total Environment**, v. 667, p. 371–383, 1 jun. 2019.
1439
- 1440 OLIVEIRA, H.H.Q. et al. Gill Histopathological Biomarkers in Fish Exposed to Trace
1441 Metals in the Todos os Santos Bay, Brazil. **Biological Trace Element Research**, v.200,
1442 p.3388–3399, 2022.
1443
- 1444 PASTORINO, P. et al. Ecology of oxidative stress in the Danube barbel (*Barbus
1445 balcanicus*) from a winegrowing district: Effects of water parameters, trace and rare earth
1446 elements on biochemical biomarkers. **Science of the Total Environment**, v. 772, 10 jun.
1447 2021.
1448

- 1449 PAULA, A. A.; RISSO, W. E.; MARTINEZ, C. B. DOS R. Effects of copper on an
1450 omnivorous (*Astyanax altiparanae*) and a carnivorous fish (*Hoplias malabaricus*): A
1451 comparative approach. **Aquatic Toxicology**, v. 237, 1 ago. 2021.
1452
- 1453 PAULINO, M.G. et al. Biotransformations, Antioxidant System Responses, and
1454 Histopathological Indexes in the Liver of Fish Exposed to Cyanobacterial Extract.
1455 **Environmental Toxicology and Chemistry**, v.39, p.1041–1051, 2020.
1456
- 1457 PEREIRA, N. J. et al. Biomarcadores histológicos em brânquias de peixes na avaliação da
1458 contaminação ambiental do Rio Mearim, Nordeste brasileiro. **Brazilian Journal of**
1459 **Development**, v. 6, n. 9, p. 68063–68079, 2020.
1460
- 1461 QU, R. et al. Metal accumulation and oxidative stress biomarkers in liver of freshwater
1462 fish *Carassius auratus* following in vivo exposure to waterborne zinc under different pH
1463 values. **Aquatic Toxicology**, v.150, p.9–16, 2014.
1464
- 1465 RABITTO, I.S. et al. Effects of dietary Pb(II) and tributyltin on neotropical fish, *Hoplias*
1466 *malabaricus*: Histopathological and biochemical findings. **Ecotoxicology and**
1467 **Environmental Safety**, v.60, p.147–156, 2005.
1468
- 1469 R CORE TEAM. R: A Language and Environment for Statistical Computing. R
1470 Foundation for Statistical Computing, Vienna, Austria, 2017. Available in:<
1471 <https://www.R-project.org/>>.
1472
- 1473 REIS, C.S. et al. Avaliação da Atividade Antrópica no Rio Guaraguaçu (Pontal do Paraná,
1474 Paraná). **Engenharia Sanitária e Ambiental**, v.20, n.3, p. 389-394, 2015.
1475
- 1476 REIS, T. et al. *Hoplias* aff. *malabaricus* Bloch, 1794 (Characiformes: Erythrinidae)
1477 parasites. **Arquivos do Instituto Biológico**, v. 84, n. 1–5, 1 fev. 2017.
1478
- 1479 RODRIGUEZ-CEA, A.; AYLLON, F.; GARCIA-VAZQUEZ, E. Micronucleus test in
1480 freshwater fish species: An evaluation of its sensitivity for application in field surveys.
1481 **Ecotoxicology and Environmental Safety**, v.56, p.442–448, 2003.

- 1482 SAAD ABDELKARIM, M. Biomonitoring and bioassessment of running water quality in
1483 developing countries: A case study from Egypt. **Egyptian Journal of Aquatic Research**,
1484 2020.
- 1485
- 1486 SALEH, Y.S.; MARIE, M.A.S. Use of *Arius thalassinus* fish in a pollution biomonitoring
1487 study, applying combined oxidative stress, hematology, biochemical and histopathological
1488 biomarkers: A baseline field study. **Marine Pollution Bulletin**, v.106, p.308–322, 2016.
- 1489
- 1490 SALGADO, L.D. et al. Integrated assessment of sediment contaminant levels and
1491 biological responses in sentinel fish species *Atherinella brasiliensis* from a sub-tropical
1492 estuary in south Atlantic. **Chemosphere**, v.219, p.15–27, 2019.
- 1493
- 1494 SALGADO, L. D. et al. Sediment contamination and toxic effects on Violet Goby fish
1495 (*Gobioides broussonnetii* - Gobiidae) from a marine protected area in South Atlantic.
1496 **Environmental Research**, v. 195, 1 abr. 2021.
- 1497
- 1498 SANTANA, M.S. et al. Diffuse sources of contamination in freshwater fish: Detecting
1499 effects through active biomonitoring and multi-biomarker approaches. **Ecotoxicology and**
1500 **Environmental Safety**, v.149, p.173–181, 2018.
- 1501
- 1502 SATO, R. Y.; COSTA, A. P. L.; PADIAL, A. A. The invasive tropical tanner grass
1503 decreases diversity of the native aquatic macrophyte community at two scales in a
1504 subtropical tidal river. **Acta Botanica Brasilica**, v. 35, n. 1, p. 140–150, 2021.
- 1505
- 1506 SAVASSI, L.A. et al. Heavy metal contamination in a highly consumed Brazilian fish:
1507 immunohistochemical and histopathological assessments. **Environmental Monitoring**
1508 **and Assessment**, v.192, 2020.
- 1509
- 1510 SEDLAK, J.; LINDSAY, R. H. Estimation of total, protein-bound, and nonprotein
1511 sulfhydryl groups in tissue with Ellman's reagent. **Analytical Biochemistry**, v.25, p.192-
1512 205, 1968.
- 1513
- 1514 SFAKIANAKIS, D.G. et al. Effect of heavy metals on fish larvae deformities: a review.
1515 **Environmental Research**, v.137, p.246–255, 2015.

- 1516 SILVA DE ASSIS, H.C. Der Einsatz von Biomarkern zur summarischen Erfassung vom
1517 Gewässerverschmutzungen. PhD thesis, Berlin Technical University, Berlin, Germany,
1518 1998.
- 1519
- 1520 SILVA, V. et al. Conhecendo os principais solos do Litoral do Paraná : abordagem para
1521 educadores do ensino fundamental e médio. Sociedade Brasileira de Ciência do Solo,
1522 Núcleo Estadual do Paraná– Matinhos (PR) : UFPR, 32 p. ISBN 978-85-86504-10-5,
1523 2013.
- 1524
- 1525 SINGO, J. M.; ARAÚJO-RAMOS, A. T.; ROCHA, J. R. C. Physical-Chemical
1526 Characterization of Peri River, Pontal do Paraná, PR, Brazil. **International Journal of**
1527 **Advanced Engineering Research and Science**, v.7, n.5, p.314-323, 2020.
- 1528
- 1529 SKIDMORE J.F. Toxicity of Zinc Compounds to Aquatic Animals, With Special
1530 Reference to Fish. **The Quarterly Review of Biology**, v.39, p.227–248, 1964.
- 1531
- 1532 SOUZA, G.L.C. et al. PhysicalChemical Parameters Evaluation of Pery River Waters in
1533 Pontal do Paraná, PR. **IOSR Journal of Environmental Science, Toxicology and Food**
1534 **Technology**, v.13, n.6, p. 69-78, 2019.
- 1535
- 1536 SOUZA, R.M. et al. Occurrence, impacts and general aspects of pesticides in surface
1537 water: A review. **Process Safety and Environmental Protection**, v.135, p.22–37, 2020.
- 1538
- 1539 TESSER, T.T., DA ROCHA, C.M., CASTRO, D. Metal contamination in omnivores,
1540 carnivores and detritivores fish along the Tramandaí River Basin, RS, Brazil.
1541 **Environmental Nanotechnology, Monitoring & Management**, v.16, 2021.
- 1542
- 1543 U.S. EPA. “Method 3520C (SW-846): Continuous Liquid-Liquid Extraction,” Revision 3.
1544 1996a. Disponível em: <<https://www.epa.gov/hw-sw846/sw-846-test-method-3520c-continuous-liquid-liquid-extraction>>. Accessed on October 28, 2022.
- 1545
- 1546
- 1547
- 1548

- 1549 U.S. EPA. “Method 3052 (SW-846): Microwave Assisted Acid Digestion of Siliceous and
1550 Organically Based Matrices”. 1996b. Disponível em: <[https://www.epa.gov/hw-
1551 sw846/sw-846-test-method-3052-microwave-assisted-acid-digestion-siliceous-and-
1552 organically-based](https://www.epa.gov/hw-sw846/sw-846-test-method-3052-microwave-assisted-acid-digestion-siliceous-and-organically-based)>. Accessed on January 17, 2023.
1553
- 1554 U.S. EPA. “Method 8270E (SW-846): Semivolatile Organic Compounds by Gas
1555 Chromatography/ Mass Spectrometry (GC/MS),” Washington, DC, 2014. Disponível em:
1556 <[https://www.epa.gov/esam/epa-method-8270e-sw-846-semivolatile-organic-compounds-
1557 gas-chromatographymass-spectrometry-gc](https://www.epa.gov/esam/epa-method-8270e-sw-846-semivolatile-organic-compounds-gas-chromatographymass-spectrometry-gc)>. Accessed on November 08, 2022.
1558
- 1559 VAN DER OOST, R.; BEYER, J.; VERMEULEN, N.P. E. Fish bioaccumulation and
1560 biomarkers in environmental risk assessment: a review. **Environmental Toxicology and
1561 Pharmacology**, v. 13, n. 2, p. 57-149, 2003.
1562
- 1563 VIANA, H. C. et al. Aggregation of hepatic melanomacrophage centers in *S. herzbergii*
1564 (Pisces, Ariidae) as indicators of environmental change and well-being. **Arquivo
1565 Brasileiro de Medicina Veterinária e Zootecnia**, v.73, n.4, p.868–876, 2021.
1566
- 1567 VIARENGO, A. et al. A simple spectrophotometric method for metallothionein
1568 evaluation in marine organisms: an application to Mediterranean and Antarctic molluscs.
1569 **Marine Environmental Research**, v.44, p.69–84, 1997.
1570
- 1571 VITULE, J.R.S.; UMBRIA, S.C.; ARANHA, J.M.R. Introduction of the African catfish
1572 *Clarias gariepinus* (BURCHELL, 1822) into Southern Brazil. **Biological Invasions**, v. 8,
1573 p.677–681, 2006.
1574
- 1575 VOIGT, C.L. et al. Bioconcentration and bioaccumulation of metal in freshwater
1576 Neotropical fish *Geophagus brasiliensis*. **Environmental Science and Pollution
1577 Research**, v.22, p.8242–8252, 2015.
1578
- 1579 WU, T. et al. Use of conductivity to indicate long-term changes in pollution processes in
1580 Lake Taihu, a large shallow lake. **Environmental Science and Pollution Research
1581 International**, v. 27, n.17, p.21376–21385, 2020.
1582

- 1583 ZARONI, M. J., SANTOS, H. G. Empresa Brasileira de Pesquisa Agropecuária
1584 (Embrapa): Espodossolos. 2021. Disponível em: [https://www.embrapa.br/agencia-de-](https://www.embrapa.br/agencia-de-informacao-tecnologica/tematicas/solos-tropicais/sibcs/chave-do-sibcs/espodossolos)
1585 [informacao-tecnologica/tematicas/solos-tropicais/sibcs/chave-do-sibcs/espodossolos](https://www.embrapa.br/agencia-de-informacao-tecnologica/tematicas/solos-tropicais/sibcs/chave-do-sibcs/espodossolos).
1586 Acessado em 2 de maio de 2023.
- 1587
- 1588 ZEE. Zoneamento Ecológico Econômico do Litoral do Paraná. Decreto Estadual nº 4.996
1589 de 05 de setembro de 2016. Curitiba. 2016.
- 1590
- 1591
- 1592
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1616 3.10. SUPPLEMENTARY MATERIAL

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1618 Supplementary material 1. Measurements of length and weight, in addition to the sex and gonadal state of the
 1619 twenty-seven fish captured along the Guaraguaçu River divided into three sectors.

Individual	Sample Sector	Total length (cm)	Total weight (g)	Sex	Gonadal State
1	1	35.4	480	Male	Imature
2	1	35	545	Male	Mature
3	1	52.5	1755	Male	Imature
4	1	38.1	630	Male	Mature
5	1	43.7	1070	Male	Mature
6	1	38.5	615	Female	Mature
7	1	32.6	390	Male	Mature
8	1	35	490	Female	Mature
9	1	46.5	1310	Male	Mature
10	1	33.8	375	Female	Mature
11	1	36.7	525	Female	Mature
12	2	36.8	535	Female	Mature
13	2	41.5	770	Female	Imature
14	2	49.4	1640	Male	Mature
15	2	55.7	2490	Female	Mature
16	2	52.4	1880	Male	Mature
17	2	37.7	515	Female	Imature
18	2	31.5	295	Male	Imature
19	2	56.3	2280	Male	Mature
20	2	56.27	2415	Female	Mature
21	2	35.4	502	Male	Imature
22	2	55.6	2045	Male	Mature
23	3	36.8	540	Male	Imature
24	3	37.1	625	Male	Mature
25	3	46.2	1270	Female	Imature
26	3	45.7	1140	Female	Mature
27	3	32.6	410	Female	Imature

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1627 Supplementary material 2. Pesticides analyzed in water in the three sectors.

Analysis	Sector 1	Sector 2	Sector 3	LQ	Reference	Date analysis
Alachlor	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Aldrin + Dieldrin	< 0,03 µg/L	< 0,03 µg/L	< 0,03 µg/L	0,03	EPA 3510C:1996, 8270D:2014	04/04/2022
Atrazine	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Bentazone	< 100,0 µg/L	< 100,0 µg/L	< 100,0 µg/L	100,0	EPA 3510C:1996, 8270D:2014	04/04/2022
Chlordane	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Endosulfan	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Endrin	< 0,03 µg/L	< 0,03 µg/L	< 0,03 µg/L	0,03	EPA 3510C:1996, 8270D:2014	04/04/2022
Heptachlor + Heptachlor epoxide	< 0,01 µg/L	< 0,01 µg/L	< 0,01 µg/L	0,01	EPA 3510C:1996, 8270D:2014	04/04/2022
Hexachlorobenzene	< 0,005 µg/L	< 0,005 µg/L	< 0,005 µg/L	0,005	EPA 3510C:1996, 8270D:2014	04/04/2022
Lindane	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Metolachlor	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Methoxychlor	< 0,03 µg/L	< 0,03 µg/L	< 0,03 µg/L	0,03	EPA 3510C:1996, 8270D:2014	04/04/2022
Pendimethalin	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Pentachlorophenol	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Propanyl	< 0,1 µg/L	< 0,1 µg/L	< 0,1 µg/L	0,1	EPA 3510C:1996, 8270D:2014	04/04/2022
Simazine	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Trifluralin	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Malathion	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Parathion	< 0,04 µg/L	< 0,04 µg/L	< 0,04 µg/L	0,04	EPA 3510C:1996, 8270D:2014	04/04/2022
Permethrin	< 0,05 µg/L	< 0,05 µg/L	< 0,05 µg/L	0,05	EPA 3510C:1996, 8270D:2014	04/04/2022
Azinphos	< 0,01000	< 0,01000	< 0,01000	0,01000	EPA 3510C:1996, 8270D:2014	04/04/2022

1628 Legend: LQ- Limit of Quantification; EPA- Environmental Protection Agency

1629 Supplementary material 3. Analysis of trace elements in water using a Flame Atomic Absorption
1630 Spectrophotometer (FAAS)

Sector	Mn	Cd	Pb	Cu	Cr	Ni
Limits of detection (mg/L)	0.002	0.003	0.011	0.005	0.006	0.008
Limit of quantification(mg/L)	0.005	0.008	0.034	0.016	0.017	0.024
1	0.0106	<LQ	UN	<LQ	<LQ	UN
2	0.0175	<LQ	UN	<LQ	<LQ	UN
3	0.0108	<LQ	UN	<LQ	<LQ	UN

1631 Legend: LQ- Limit of Quantification; UN- Undetectable

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1634 Supplementary material 4. Analysis of trace elements in water using Inductively Coupled Plasma Optical
1635 Emission Spectrometry (ICP - OES)

Sector	Al	As	Fe	Zn
Limit of quantification(mg/L)	0.01	0.02	0.01	0.01
1	1.73	UN	5.46	UN
2	2.51	UN	12.41	UN
3	2.68	UN	8.94	UN

1636 Legend: UN- Undetectable

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1640 Supplementary material 5. Analysis of mercury in water using Inductively Coupled Plasma Optical Emission
1641 Spectrometry (ICP - OES)

Sector	Hg
Limit of quantification ($\mu\text{g/L}$)	0.2
1	<LQ
2	<LQ
3	<LQ

1642 Legend: LQ- Limit of Quantification

1643 Supplementary material 6. Table of concentration (mg/Kg) of trace elements in *Hoplias malabaricus* muscle in
 1644 the three sampling sectors.

	Trace elements									
	Al	As	Cr	Cu	Mn	Fe	Zn	Cd	Ni	Pb
LQ (mg/L) *	0.01	0.02	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.02
Brazilian Legislation (mg/Kg) **	-	1	0.1	30	-	-	50	0.05	5	0.3
Statistical***	p-value = 0.19 (> 0.05)									
Trace elements (mg/Kg)										
Sector	Al	As	Cr	Cu	Mn	Fe	Zn	Cd	Ni	Pb
S1	3.871	ND	ND	ND	4.608	34.585	83.939	ND	ND	ND
S1	14.675	ND	ND	ND	8.830	35.904	68.525	ND	ND	ND
S1	8.159	ND	ND	ND	ND	34.666	75.676	ND	ND	ND
S1	ND	ND	ND	ND	6.646	22.852	103.558	ND	ND	ND
S1	ND	ND	ND	ND	5.349	39.987	36.713	ND	ND	ND
S2	ND	ND	ND	ND	ND	15.502	30.947	ND	ND	ND
S2	ND	ND	ND	ND	ND	38.911	51.339	ND	ND	ND
S2	2.014	ND	3.469	ND	14.572	75.367	194.459	ND	ND	ND
S2	ND	ND	2.281	ND	ND	46.845	34.749	ND	ND	ND
S2	ND	ND	ND	ND	5.164	84.382	64.138	ND	ND	ND
S3	2.727	ND	5.551	2.671	2.417	80.452	51.458	ND	ND	ND
S3	3.592	ND	ND	2.415	ND	44.252	56.152	ND	ND	ND
S3	5.362	ND	ND	ND	ND	60.098	30.586	ND	ND	ND
S3	2.432	5.569	ND	ND	ND	48.370	21.206	ND	ND	ND
S3	4.482	ND	ND	3.435	ND	52.089	54.259	ND	ND	ND

1645 Legend: * LQ: ICP-OES Limit of Quantification ; ** DECREE 55.871 of 03/26/1965, Modifies DECREE No.
 1646 50.040, 01/24/1961, referring to regulatory norms for the use of food additives / ANVISA: RDC No. 42,
 1647 08/29/2013, Provides for the Technical Regulation MERCOSUR on Maximum Limits of Inorganic
 1648 Contaminants in Food; ***p-value > 0.05 means no significant difference between sectors; ND - Not
 1649 Determined.

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1656 Supplementary material 7. Loadings table representing the correlation of each original variable (trace elements)
1657 with the x (PCA1) and y (PCA2) axes. Zn and Fe are the two elements that have the greatest relationship with
1658 the PCA1 and PCA2 axes, respectively.

Trace elements	PCA1	PCA2
Al (mg/Kg)	0	0
As (mg/Kg)	0	0
Cr (mg/Kg)	0	0
Cu (mg/Kg)	0	0
Mn (mg/Kg)	0	0
Fe (mg/Kg)	0.134	-0.989
Zn (mg/Kg)	0.987	0.135

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1680 Supplementary Material 8. Mean values and standard errors (Mean \pm SE) of the biochemical biomarkers
 1681 responses.
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Tissue	Biomarker	Sectors		
		S1	S2	S3
Brain	AChE	24.0 \pm 2.66 ^a	10.0 \pm 1.73 ^b	22.2 \pm 3.51 ^a
Muscle	AChE	17.20 \pm 1.31 ^{ab}	21.9 \pm 1.72 ^a	12.4 \pm 1.44 ^b
Gill	GST	*4.05 \pm 0.06 ^a	*4.14 \pm 0.07 ^a	*4.53 \pm 0.11 ^b
Liver	GST	49.30 \pm 3.24 ^a	63.3 \pm 4.69 ^b	51.3 \pm 4.16 ^{ab}
Kidney	GST	*1.72 \pm 0.25 ^a	*2.67 \pm 0.05 ^b	*3.35 \pm 0.11 ^b
Gill	GSH	1.05 \pm 0.17 ^a	1.48 \pm 0.17 ^a	1.39 \pm 0.27 ^a
Liver	GSH	*0.28 \pm 0.29 ^a	*1.17 \pm 0.36 ^b	*1.62 \pm 0.44 ^b
Kidney	GSH	0.48 \pm 0.06 ^a	0.71 \pm 0.10 ^a	0.67 \pm 0.23 ^a
Gill	SOD	140 \pm 9.09 ^a	186 \pm 10.30 ^b	214 \pm 14.50 ^b
Liver	SOD	*4.98 \pm 0.07 ^a	*5.66 \pm 0.11 ^b	*5.64 \pm 0.08 ^b
Kidney	SOD	*5.23 \pm 0.09 ^a	*4.92 \pm 0.09 ^a	*5.05 \pm 0.12 ^a
Gill	CAT	3.40 \pm 0.49 ^a	1.19 \pm 0.49 ^b	0.89 \pm 0.31 ^b
Liver	CAT	0.70 \pm 0.09 ^a	0.68 \pm 0.06 ^a	1.01 \pm 0.22 ^a
Kidney	CAT	0.74 \pm 0.09 ^a	1.02 \pm 0.07 ^a	0.86 \pm 0.26 ^a
Gill	GPx	10.80 \pm 1.17 ^a	15.60 \pm 1.24 ^b	20.30 \pm 2.52 ^b
Liver	GPx	20.50 \pm 1.34 ^a	30.20 \pm 1.99 ^b	27.70 \pm 3.68 ^{ab}
Kidney	GPx	8.33 \pm 0.99 ^a	16.30 \pm 1.40 ^b	23.5 \pm 3.72 ^b
Gill	LPO	*1.86 \pm 0.05 ^a	*2.37 \pm 0.03 ^b	*2.61 \pm 0.19 ^b
Liver	LPO	*2.58 \pm 0.06 ^a	*3.18 \pm 0.10 ^b	*3.45 \pm 0.16 ^b
Kidney	LPO	17.2 \pm 1.33 ^a	18.5 \pm 1.07 ^a	21.80 \pm 2.49 ^a
Liver	MET	8.84 \pm 0.50 ^a	10.3 \pm 0.33 ^a	9.65 \pm 2.36 ^a

1683 Legend: **AChE**- acetylcholinesterase; **GST**- Glutathione S-transferase; **GSH**- Reduced Glutathione; **SOD**-
 1684 Superoxide dismutase; **CAT**- Catalase; **GPx**-Glutathione peroxidase; **LPO**- Lipoperoxidation; **MET**-
 1685 Metallothionein; **S1**- Sector 1; **S2**- Sector 2; **S3**- Sector 3; (*) Show the use of log transformation to achieve the
 1686 assumptions of homogeneity and normality of the data; Different letters mean difference between sectors.

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4. CONSIDERAÇÕES FINAIS

Apesar do Rio Guaraguaçu possuir uma importância social provendo o abastecimento de água para os municípios de Pontal do Paraná, Matinhos e Paranaguá (Elste et al., 2019), econômica voltada para a pesca de subsistência (Reis et al., 2015) e ecossistêmica, a partir da análise dos parâmetros físico-químicos da água foi demonstrado uma clara e aparente diferença quando separado entre os três setores descritos, principalmente ao observar a condutividade e transparência dos setores. Observa-se que a condutividade no setor 2 se encontra maior que no setor 3, setor que sofre com a influência de marés por estar mais próximo da foz do rio, e conseqüentemente seria o setor com maior condutividade e salinidade do Rio Guaraguaçu (Staszczak; Rocha, 2018). Tal fato, pode ser explicado principalmente pela influência do canal retificado do rio Pery, que possui um aterro sanitário além da presença de lançamento de esgoto doméstico, podendo estar influenciando e apresentando um aumento da condutividade pelo excesso da concentração de elementos, como fósforo e nitrogênio, e seus íons considerados naturalmente limitantes do crescimento biótico (Reis et al., 2015; Souza et al., 2019; Singo; Araújo-Ramos; Rocha, 2020).

Nas análises químicas da água, não foi encontrado uma contaminação por agrotóxicos no Rio Guaraguaçu, pela metodologia utilizada. O elemento traço manganês foi quantificado, mas abaixo do limite estipulado por lei. Ferro e alumínio, dois elementos essenciais, apresentaram concentrações maiores do que o estipulado por lei e sua presença pode estar relacionada às características geomorfológicas e pedológicas da região. Entretanto, assim como os agrotóxicos, os elementos traço podem bioacumular no tecido animal e ocasionar sérios problemas para a saúde do organismo. A ausência de agrotóxicos em concentrações significativas pode estar relacionada com a agricultura que historicamente em sua maioria é de origem familiar/subsistência, possuindo como principais cultivos a banana, mandioca, o feijão, o milho e o arroz (ZEE, 2016).

Apesar da maioria dos elementos traços não estarem presentes na água de acordo com os métodos utilizados, no músculo das traíras, o elemento Cr foi encontrado em dois organismos do setor 2 e um do setor 3, os setores caracterizados por possuírem diferentes graus de impacto, o que pode indicar a contaminação da região por determinadas substâncias. No geral, os únicos elementos que apareceram em todos os organismos de todos os setores foram Fe que não possui limite estabelecido, e Zn que possuía limite, mas a legislação que o estabelecia foi revogada. A presença de tais elementos em todas as amostras pode estar relacionada ao tipo de solo presente (IAT, 2006) ou sedimento na região do rio Guaraguaçu,

já que estatisticamente não há diferença significativa entre os três setores para todos os elementos traço analisados. No entanto, vale destacar a importância de uma atualização na legislação brasileira ao estipular limites de mais elementos traço em alimentos, água e sedimento devido principalmente o risco de em concentrações exacerbadas, mesmo elementos essenciais, causarem problemas e riscos à saúde dos peixes e conseqüentemente dos seres humanos.

Ao analisar as respostas das atividades de biomarcadores bioquímicos, verificamos que existe uma diferença entre os setores amostrados. Evidencia-se assim, que principalmente no setor 2, que possui extrema influência antrópica, possui um aumento do sistema antioxidante e de biotransformação, além de aumento de lipoperoxidação comparado com o setor 1 caracterizado por ser conservado. O setor 2 também apresentou uma inibição de acetilcolinesterase cerebral, comparado aos outros dois setores. A inibição da atividade de AChE é amplamente conhecida como biomarcador de exposição para pesticidas organofosforados e carbamatos (Fajardo; Ocampo, 2018). No entanto, diversos contaminante também tem sido classificados como inibidores da atividade de AChE, incluindo metais e outras classes de poluentes orgânicos ambientais (Fu et al., 2018). Esses resultados corroboraram com a hipótese estipulada pelo presente trabalho. Com isso, verificou-se também a importância da espécie *H. malabaricus* como excelente modelo biológico para utilização em biomonitoramento, já que foi capaz de demonstrar diferentes resultados para diferentes biomarcadores.

Os resultados de histopatologia, reforçaram o encontrado na análise dos biomarcadores bioquímicos. Lesões teciduais da brânquia e fígado, foram mais evidentes em peixes do setor 2, considerado mais antropizado, além de peixes do setor 3 que mesmo com menos impactos, ainda apresentou danos teciduais nos organismos. Dentre as lesões encontradas em brânquias podem se destacar principalmente no setor 2 a presença de aneurisma e hiperplasia com fusão parcial da lamela secundária, dobras das extremidades das lamelas secundárias, hiperplasia com fusão total das lamelas secundárias e aumento das células de muco. No setor 3, as alterações mais comumente encontradas foram lifting epitelial e hiperplasia com fusão parcial. Ao analisar o fígado, entre as alterações encontradas em peixes do setor 2, pode-se citar a presença de centro de melanomacrófagos, congestão sanguínea nos sinusóides, vacuolização dos hepatócitos e necrose. Já para o setor 3, destaca-se a presença de morte celular, ou necrose.

Ao se analisar a presença de micronúcleo, todos os três setores indicaram mutagenicidade nos organismos. Além disso, alterações nucleares também foram observadas

como “blebbed”, “notched”, “lobed”, “binucleus” e “vacuolated”, entretanto não houve diferença estatística significativa entre os três setores analisados do Rio Guaraguaçu. Os produtos oriundos e utilizados pelas atividades antrópicas são as principais causas de alterações morfológicas no núcleo de células de peixes (Canedo et al., 2021), como agrotóxicos (Oliveira et al., 2019), lançamento de esgoto (Lehun et al., 2021), elementos traço (Francisco et al., 2019; ALI et al., 2020) e hidrocarbonetos policíclicos aromáticos (HPAs) (Benincá et al., 2011).

Logo, vale salientar a importância do presente trabalho, ao destacar seu pioneirismo sendo um dos primeiros, se não o primeiro, trabalho na análise da qualidade da água do rio Guaraguaçu que utiliza o biomonitoramento com peixes nativos para análise de biomarcadores. A utilização do biomonitoramento junto do monitoramento tradicional se torna uma ferramenta eficaz, ao conseguir evidenciar os problemas encontrados em um dos bens e recursos mais preciosos de qualquer ecossistema, os seus organismos. Com os resultados aqui alcançados, busca-se incentivar futuros trabalhos no importante rio Guaraguaçu criando um alerta para a necessidade de uma melhora na qualidade de vida de todos os residentes com grau de vulnerabilidade social que precisam desse rio para subsistência, abastecimento e sofrem com as consequências da contaminação desse rio, além de criar um direcionamento para criação de ações governamentais capazes de controlar, diminuir e até mesmo encerrar com atividades antrópicas capazes de interferir na qualidade da água desse importante rio da costa paranaense.

Entretanto, salienta-se que sejam necessários outros trabalhos para análise de possíveis contaminantes em sedimentos. Uma análise mais profunda da possibilidade da presença de outros xenobióticos como HPAs e poluentes orgânicos persistentes (POPs) também é de extrema importância, além da expansão das análises para mais de uma espécie de peixe ou até mesmo para mais de uma espécie de animal em diferentes níveis tróficos, com a possibilidade de análise em períodos distintos do ano ao avaliar os efeitos da sazonalidade no Rio Guaraguaçu, para que assim se possa ter um direcionamento maior para quais xenobióticos podem estar influenciando a saúde dos organismos da região do Guaraguaçu.

REFERÊNCIAS

ABREU-MOTA, M. A. et al. Sedimentary biomarkers along a contamination gradient in a human-impacted sub-estuary in Southern Brazil: A multi-parameter approach based on spatial and seasonal variability. **Chemosphere** v.103, p.156–163, 2014.

AEBI, H. Catalase in vitro. **Methods in Enzymology**, v.105, p.121-126, 1984.

AL-SABTI, K. et al. Chromium-induced Micronuclei in Fish. **Journal of Applied Toxicology**, v. 13, n. 5, p. 333–336, 1994.

AL-SABTI, K.; METCALFE, C. D. Fish micronuclei for assessing genotoxicity in water. **Mutation Research**, v. 343, p. 121–135, 1995.

ALI, D. et al. Fish as bio indicators to determine the effects of pollution in river by using the micronucleus and alkaline single cell gel electrophoresis assay. **Journal of King Saud University - Science**, v. 32, p. 2880–2885, 2020.

ALI, F. K.; EL-SHEHAWI, A. M.; SEEHY, M. A. Micronucleus test in fish genome: A sensitive monitor for aquatic pollution. **African Journal of Biotechnology**, v. 7, n. 5, p. 606–612, 2008.

AMIARD, J. C. et al. Metallothioneins in aquatic invertebrates: Their role in metal detoxification and their use as biomarkers. **Aquatic Toxicology**, v. 76, n. 2, p. 160–202, 10 fev. 2006.

ANGELI, J.L.F. et al. Statistical assessment of background levels for metal contamination from a subtropical estuarine system in the SW Atlantic (Paranaguá Estuarine System, Brazil). **Journal of Sedimentary Environments**, v.5, p.137–150, 2020.

ARAÚJO, E. S.; VITULE, J. R. S.; PADIAL, A. A. A. Checklist of aquatic macrophytes of the Guaraguaçu river basin reveals a target for conservation in the Atlantic rainforest. **Acta Scientiarum. Biological Sciences**, v. 43, n.1, e50542, 2021.

ARIF, M.; KUMAR, R.; PARVEEN, S. Reduction in Water Pollution in Yamuna River Due to Lockdown Under COVID-19 Pandemic. ChemRxiv. Cambridge: Cambridge Open Engage; This content is a preprint and has not been peer-reviewed, 2020.

ARMAS, E. D. et al. DIAGNÓSTICO ESPAÇO-TEMPORAL DA OCORRÊNCIA DE HERBICIDAS NAS ÁGUAS SUPERFICIAIS E SEDIMENTOS DO RIO CORUMBATAÍ E PRINCIPAIS AFLUENTES. **Química Nova**, v. 30, n. 5, p. 1119–1127, 2007.

ATLI, G.; CANLI, M. Responses of metallothionein and reduced glutathione in a freshwater fish *Oreochromis niloticus* following metal exposures. **Environmental Toxicology and Pharmacology**, v. 25, n. 1, p. 33–38, jan. 2008.

AYHAN, N.; YAMAN, M. Evaluation of Iron and Zinc Contents of Some Fish Species. **Biological Trace Element Research**, v. 200, p.1376–1382, 2022.

BALBONI, L.; COLAUTTI, D. C.; BAIGÚN, C. R. M. Biology of growth of *Hoplias aff. malabaricus* (Bloch, 1794) in a shallow pampean lake (Argentina). **Neotropical Ichthyology**, v. 9, n. 2, p. 437–444, 2011.

BALLESTEROS, M.L. et al. Multi-biomarker responses in fish (*Jenynsia multidentata*) to assess the impact of pollution in rivers with mixtures of environmental contaminants. **Science of The Total Environment**, v.595, p. 711- 722, 2017.

BARBIERI, G. Dinâmica da reprodução e crescimento de *Hoplias malabaricus* (Bloch, 1794) (Osteichthyes, Erythrinidae) da Represa do Monjolinho, São Carlos/SP. **Revista Brasileira de Zoologia**, v. 6, n. 2, p. 225–233, 1989.

BARBOSA, K. B. F. et al. Estresse oxidativo: conceito, implicações e fatores modulatórios. **Revista de Nutrição**, v. 23, n. 4, p. 629–643, 2010.

BARHOUMI, S. et al. Spatial and seasonal variability of some biomarkers in *Salaria basilisca* (Pisces: Blennidae): Implication for biomonitoring in Tunisian coasts. **Ecological Indicators**, v.14, p.222–228, 2012.

BENINCÁ, C. et al. Chronic genetic damages in *Geophagus brasiliensis* exposed to anthropic impact in Estuarine Lakes at Santa Catarina coast – southern of Brazil. **Environmental Monitoring and Assessment**, v.184, p.2045–2056, 2011.

BERNET, D. et al. Histopathology in fish: proposal for a protocol to assess aquatic pollution. **Journal of Fish Diseases**, v. 22, n. 1, p. 25–34, 1999.

BORA, L.S., THOMAZ, S.M., PADIAL, A.A. Evidence of rapid evolution of an invasive *poaceae* in response to salinity. **Aquatic Sciences**, v. 82, n.76, 2020.

BRADFORD, M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. **Analytical Biochemistry**, v. 72, n. 1, p. 248- 254, 1976.

BRASIL. Decreto nº. 55871 de 26 de março de 1965. Modifica o Decreto nº 50.040, de 24 de janeiro de 1961, referente a normas reguladoras do emprego de aditivos para alimentos, alterado pelo Decreto nº 691, de 13 de março de 1962. Diário Oficial da União, Poder Executivo, Brasília, 1965.

BRASIL. Lei nº 7.802, de 11 de julho de 1989. Dispõe sobre a pesquisa, a experimentação, a produção, a embalagem e rotulagem, o transporte, o armazenamento, a comercialização, a propaganda comercial, a utilização, a importação, a exportação, o destino final dos resíduos e embalagens, o registro, a classificação, o controle, a inspeção e a fiscalização de agrotóxicos, seus componentes e afins, e dá outras providências. Diário Oficial da União, Brasília, DF 11/07/1989.

BRASIL. Lei nº 9.433, de 8 de janeiro de 1997. Institui a Política Nacional de Recursos Hídricos, cria o Sistema Nacional de Gerenciamento de Recursos Hídricos, regulamenta o inciso XIX do art. 21 da constituição federal, e altera o art. 1º da lei nº 8.001, de 13 de março de 1990, que modificou a lei nº 7.990, de 28 de dezembro de 1989. Diário oficial da República Federativa do Brasil, Brasília, DF 09/01/1997, P. 470.

BRASIL. Ministério da Saúde. Agência Nacional de Vigilância Sanitária. Resolução - RDC n. 42 de 29 de agosto de 2013. Dispõe sobre o Regulamento Técnico MERCOSUL sobre Limites Máximos de Contaminantes Inorgânicos em Alimentos. Diário Oficial da República Federativa do Brasil, Brasília, 168, 33, Seção 1, 2013.

BURKINA, V.; ZLABEK, V.; ZAMARATSKAIA, G. Effects of pharmaceuticals present in aquatic environment on Phase I metabolism in fish. **Environmental Toxicology and Pharmacology**, v.40, p.430–444, 2015.

CALADO, S.L.M. et al. Biochemical and genotoxicity assessment of a polluted urban river using the native fish *Astyanax altiparanae* Garutti & Britski (Teleostei, Characidae). **Ecotoxicology and Environmental Contamination**, v. 14, n.1, p. 73-77, 2019.

CANEDO, A. et al. Micronucleus test and nuclear abnormality assay in zebrafish (*Danio rerio*): Past, present, and future trends. **Environmental Pollution**, v.290, 2021.

CARRASCO, K. R., TILBURY, K. L.; MYERS, M. S. Assessment of the piscine micronucleus test as an in situ biological indicator of chemical contaminant effects. **Canadian Journal of Fisheries and Aquatic Sciences**, v. 47, p. 2123-2136, 1990.

CARVALHO, B. M. et al. Length-weight relationships of native and non-native fishes in a subtropical coastal river of the Atlantic Rain Forest. **Acta Limnologica Brasiliensia**, v. 34, n. 5, 2022.

CARVALHO, L. N.; FERNANDES, C. H. V.; MOREIRA, V. S. S. Alimentação de *Hoplias malabaricus* (Bloch, 1794) (Osteichthyes, Erythrinidae) no rio Vermelho, Pantanal Sul Mato-Grossense. **Revista brasileira Zoociências**, v. 4, n. 2, p. 227–236, 2002.

CASTRO, J. S. et al. Biomarcadores histopatológicos na espécie *Hoplias malabaricus* (Pisces, Osteichthyes, Erythrinidae) em uma Unidade de Conservação de São Luís (MA). **Arquivo Brasileiro de Medicina Veterinária e Zootecnia**, v. 66, n. 6, p. 1687–1694, 2014.

CAVALCANTI, L. P. A. N. et al. Intoxicação por Organofosforados: Tratamento e Metodologias Analíticas Empregadas na Avaliação da Reativação e Inibição da Acetilcolinesterase. **Revista Virtual de Química**, v. 8, n. 3, p. 739–766, 2016.

CAVALLINI, N.G. Contaminação ambiental na bacia do rio Guaraguaçu: determinação quantitativa de contaminantes inorgânicos e diagnóstico a partir de bioindicador. Matinhos-PR, 2018. 156 p. Dissertação (Mestrado em Desenvolvimento Territorial Sustentável) - Curso de Pós Graduação em Desenvolvimento Territorial Sustentável (PPGDTS), Universidade Federal do Paraná- Setor Litoral, 2018.

CAVALLINI, N.G. et al. Determination of Lead (Pb) in stools of *Lontra longicaudis* (Olfers, 1818) by flame atomic absorption spectrometry (FAAS). **Eclética Química Journal**, v.43, p.70-78, 2018.

CAVALLINI, N. G.; REIS, R. A.; TIEPOLO, L. M. O SILENCIOSO GRITO QUÍMICO: RISCOS E AMEAÇAS NO RIO GUARAGUAÇU SOB A PERSPECTIVA ECOSISTÊMICA / THE QUIET CHEMICAL SCREAM: RISKS AND IMMINENT THREATS IN THE GUARAGUAÇU RIVER UNDER THE ECOSYSTEM SYSTEM. **Brazilian Journal of Development**, v. 6, n. 9, p. 66540–66553, 2020.

CHAUDHRY, F.N.; MALIK, M.F. Factors Affecting Water Pollution: A Review. **Journal of Ecosystem and Ecography**, v.07, 2017.

CHOWDHURY, S.; SAIKIA, S. K. Oxidative Stress in Fish: A Review. **Journal of Scientific Research**, v.12, n.1, p.145–160, 2020.

CHU-KOO, F. W.; PÉREZ, A. M. D. BIOLOGÍA Y CULTIVO DEL FASACO *Hoplias malabaricus* Bloch 1794 (CHARACIFORMES: ERYTHRINIDAE). **Folia Amazónica**, v. 16, n. 1–2, p. 11–21, 2007.

CONAMA. Resolução CONAMA 357/2005, de 17 de março de 2005. Dispõem sobre a classificação dos corpos de água e diretrizes ambientais para seu enquadramento, bem como estabelece as condições e padrões de lançamento de efluentes, e dá outras providências. Brasília, Ministério do Meio ambiente, 2005.

CONAMA. Resolução CONAMA 430/2011, de 13 de maio de 2011. Dispõem sobre as condições e padrões de lançamento de efluentes, complementa e altera a resolução nº357, de 17 de março de 2005, do Conselho Nacional do Meio Ambiente– CONAMA. Brasília, Ministério do Meio Ambiente, 2011.

CONTENTE, R.F.; STEFANONI, M.F.; SPACH, H.L. Fish assemblage structure in an estuary of the Atlantic Forest biodiversity hotspot (southern Brazil). **Ichthyological Research**, v. 58, p.38-50, 2011.

COTRAN, R. S. et al. Pathologic basis of disease. Philadelphia, PA: **Elsevier Saunders**, 2005.

COSTA, J. R. M. A. et al. Enzymatic inhibition and morphological changes in *Hoplias malabaricus* from dietary exposure to lead(II) or methylmercury. **Ecotoxicology and Environmental Safety**, v. 67, n. 1, p. 82–88, maio 2007.

CRUZ-ESQUIVEL, Á.; MARRUGO-NEGRETE, J. Methylmercury concentrations in *Prochilodus magdalenae* (Teleostei: Curimatidae) and *Hoplias malabaricus* (Teleostei: Erythrinidae) in the lower Cauca-Magdalena river system, northern Colombia. **Acta Biologica Colombiana**, v. 27, n. 1, p. 28–35, 1 jan. 2022.

DALZUCHIO, T. et al. The use of biomarkers to assess the health of aquatic ecosystems in Brazil: a review. **International Aquatic Research**, v.8, n.4, p. 283-298, 2016.

DOMANICO, A.; DELFINO, R.; FREYRE, L. EDAD Y CRECIMIENTO DE HOPLIAS MALABARICUS MALABARICUS(BLOCH,1794)(TELEOSTEI,ERYTRINIDAE)EN LA LAGUNA DE LOBOS(ARGENTINA). **IHERINGIA, Série Zoologia**, v. 74, p. 141–149, 1993.

DUDGEON, D. Multiple threats imperil freshwater biodiversity in the Anthropocene. **Current Biology**, v. 29, p. 942–995, 2019.

DUFFUS, J.H. “Heavy Metals”-A Meaningless Term? (IUPAC Technical Report). **Pure and Applied Chemistry**. v.74, p.793–807, 2002.

EL-KHAYAT, H. M. M. et al. Assessment of metallothionein expression in *Biomphalaria alexandrina* snails and *Oreochromis niloticus* Fish as a biomarker for water pollution with heavy metals. **Egyptian Journal of Aquatic Biology & Fisheries**, v. 24, n. 2, p. 209–223, 2020.

ELLMAN, G.L. et al. A new and rapid colorimetric determination of acetylcholinesterase activity. **Biochemical Pharmacology**, v.7, p.88– 95, 1961.

ELSTE, G.A.S. et al. A contaminação do rio Guaraguaçu (Litoral do Paraná): limites e riscos ao desenvolvimento territorial regional. **Guaju, Matinhos**, v.5, n.2, p. 54-70, jul./dez. 2019.

ESCALANTE-ROJAS, M.C. et al. Integrated use of biomarkers to evaluate the reproductive physiology of *Astyanax fasciatus* and *Hoplias malabaricus* males (Teleostei: Characiformes) in polluted reservoirs. **Ecotoxicology and Environmental Safety**, v.208, 2021.

ESTEVEZ, F.A. Fundamentos de Limnologia. 2o edição, Interciência, 1998.

ESTEVEZ, F.A. Fundamentos de Limnologia. 3º edição, Interciência, 2011.

EUROPEAN COMMUNITY (EC). Commission Regulation of the European Community (EC) No. 1881/2006, dated December 19, 2006, setting maximum levels for certain contaminants in foodstuffs. EUR-Lex (L364), p. 5-24, 2006.

EVANS, T.G.; KÜLTZ, D. The cellular stress response in fish exposed to salinity fluctuations. **Journal of Experimental Zoology Part A: Ecological and Integrative Physiology**, v.333, n.6, p. 421–435, 2020.

FABRIN, T. M. C. et al. Performance of biomarkers metallothionein and ethoxyresorufin O-deethylase in aquatic environments: A meta-analytic approach. **Chemosphere**, v. 205, p. 339–349, 1 ago. 2018.

FAJARDO, L.J., OCAMPO, P.P. Inhibition of acetylcholinesterase activities in whitegoby, *Glossogobius giuris* from the East Bay of Laguna Lake, Philippines. **International Journal of Agricultural Technology**, v.14, n.7, p.1181- 1192, 2018.

FERNANDES, I. F. et al. Ecotoxicological evaluation of water from the Sorocaba River using an integrated analysis of biochemical and morphological biomarkers in bullfrog tadpoles, *Lithobates catesbeianus* (Shaw, 1802). **Chemosphere**, v. 275, 2021.

FLORES-LOPES, F.; THOMAZ, A.T. Histopathologic alterations observed in fish gills as a tool in environmental monitoring. **Brazilian Journal of Biology**, 2011.

FOOD AND AGRICULTURE ORGANIZATION (FAO). Compilation of legal limits for hazardous substances in fish and fishery products. **FAO Fisheries Circular**, v.464, p.5–100, 1983.

FRANCISCO, C.D.M. et al. Genotoxicity assessment of polluted urban streams using a native fish *Astyanax altiparanae*. **Journal of Toxicology and Environmental Health, Part A**, v. 82, p. 514–523, 2019.

FREITAS, L.C. et al. Histological biomarkers and biometric data on trahira *Hoplias malabaricus* (Pisces, Characiformes, Erythrinidae): a bioindicator species in the Mearim river, Brazilian Amazon. **Brazilian Journal of Biology**, v. 82, e263047, 2022.

FRIBERG, N. et al. Biomonitoring of human impacts in freshwater ecosystems: the good, the bad and the ugly. **Advances in Ecological Research**, v. 44, p. 1–68, 2011.

FU, H. et al. Acetylcholinesterase Is a Potential Biomarker for a Broad Spectrum of Organic Environmental Pollutants. *Environ. Sci. Technol.*, v.52, n.15, p.8065–8074, 2018.

GALVANESE, E. F. et al. Community stability and seasonal biotic homogenisation emphasize the effect of the invasive tropical tanner grass on macrophytes from a highly dynamic neotropical tidal river. **Aquatic Sciences**, v. 84, n. 2, 1 abr. 2022.

GAO, R. et al. Mechanism of pyrogallol autoxidation and determination of superoxide dismutase enzyme activity. **Bioelectrochemistry and Bioenergetics**, v.45, n.1, p.41-45, 1998.

GARAI, P. et al. Effect of Heavy Metals on Fishes: Toxicity and Bioaccumulation. **The Journal of Clinical Toxicology** , v.11, 2021.

GARCÍA-CAPARRÓS, P. et al. Oxidative Stress and Antioxidant Metabolism under Adverse Environmental Conditions: a Review. **Botanical Review**, v. 87, p. 421–466, 2021.

GAZOLA-SILVA, F. F. et al. Chthonerpeton viviparum Parker & Wettstein, 1929(Amphibia, Gymnophiona, Typhlonectinae) in Paraná state, Braziland the first record of predation of this species by *Hoplias malabaricus* (Bloch, 1794) (Actinopterygii, Erythrinidae). **Pan-American Journal of Aquatic Sciences** , v.2, n.3, p. 261-262, 2007.

GEMSEMER, R.W. et al. Evaluating the effects of pH, hardness, and dissolved organic carbon on the toxicity of aluminum to freshwater aquatic organisms under circumneutral conditions. **Environmental Toxicology and Chemistry**, v. 37, p. 49-60, 2018.

GEORGIEVA, E. et al. A Review on Multi-Biomarkers in Fish for the Assessment of Aquatic Ecosystem Contamination with Organic Pollutants. **Ecologia Balkanica**, v.13, p.321–330, 2021.

GOMES, A. D. et al. The role of ovarian steroids in reproductive plasticity in *Hoplias malabaricus* (Teleostei: Characiformes: Erythrinidae) in tropical reservoirs with different degrees of pollution. **General and Comparative Endocrinology**, v. 222, p. 1–10, 1 out. 2015.

GONZALES, G.; CRESPO, S.; BRUSLE, J. Histocytological study of the liver of the cabrilla sea bass, *Serranus cabrilla* (Teleostei, Serranidae), an available model for marine fish experimental studies. **Journal of Fish Biology**, v.43, p.363-73, 1993.

GRASSI, D.J. et al. Cytogenetic characterization of *Hoplias malabaricus* (Bloch, 1794) from the Ctlamochita River (Córdoba, Argentina): First evidence for southernmost populations of this species complex and comments on its biogeography. **Comparative Cytogenetics**, v. 11, p.15–28, 2017.

GUTTERIDGE, J. M. C. Invited review free radicals in disease processes: A compilation of cause and consequence. **Free Radical Research Communications**, v. 19, n. 3, p. 141–158, 1993.

HÄDER, D.P. et al. Anthropogenic pollution of aquatic ecosystems: emerging problems with global implications. **Science of the Total Environment**. v. 713, 2020.

HAFEMAN, D. G.; SUNDE, R.A.; HOEKSTRA, W.C. Effect of dietary selenium on erythrocyte and liver glutathione peroxidase in the rat. **Journal of Nutrition**, v.104, n.4, p.580-587, 1974.

HASHIMI, M.H.; HASHIMI, R.; RYAN, Q. Toxic Effects of Pesticides on Humans, Plants, Animals, Pollinators and Beneficial Organisms. **Asian Plant Research Journal**, v. 5, p. 37–47, 2020.

HASSANINEZHAD, L. et al. Assessment of gill pathological responses in the tropical fish yellowfin seabream of Persian Gulf under mercury exposure. **Toxicology Reports**, v.1, p.621-628, 2014.

HAYASHI, M. The micronucleus test-most widely used in vivo genotoxicity test. **Genes and Environment**, 2016.

HEDAYATI, A. Fish Biomarkers, Suitable Tools For Water Quality Monitoring. **International Journal of Veterinary and Animal Research**, v. 1, n. 3, p. 63–69, 2018.

HOOFTMAN, R.N.; RAAT, W.K. Induction of nuclear anomalies (micronuclei) in peripheral blood erythrocytes of Eastern mudminnow *Umbra pygmaea* by ethyl methanesulphonate, **Mutation Research**, v.104, p.147-152, 1982.

HUBER, P. C.; ALMEIDA, W. P. GLUTATIONA E ENZIMAS RELACIONADAS: PAPEL BIOLÓGICO E IMPORTÂNCIA EM PROCESSOS PATOLÓGICOS. **Química Nova**, v. 31, n. 5, p. 1170–1179, 2008.

HUSSAIN, B. et al. Fish eco-genotoxicology: Comet and micronucleus assay in fish erythrocytes as in situ biomarker of freshwater pollution. **Saudi Journal of Biological Sciences**, v.25, p.393–398, 2018.

INSTITUTO ÁGUA E TERRA (IAT). Plano de Manejo: Estação Ecológica do Guaraguaçu. 2006. Disponível em: <<https://www.iat.pr.gov.br/Pagina/Plano-de-Manejo-Estacao-Ecologica-do-Guaraguacu>>. Acesso em 13 de setembro de 2022.

JARAPALA, S.R., KANDLAKUNTA, B., THINGNGANING, L. Evaluation of trace metal content by ICP-MS using closed vessel microwave digestion in freshwater fish. **Journal of Environmental and Public Health**, 2014.

JAVED, M.; USMANI, N. Accumulation of heavy metals and human health risk assessment via the consumption of freshwater fish *Mastacembelus armatus* inhabiting, thermal power plant effluent loaded canal. **Springerplus**, v. 5, 2016.

JESUS, I.S. et al. Analysis of metal contamination and bioindicator potential of predatory fish species along Contas River basin in northeastern Brazil. **Bulletin of Environmental Contamination and Toxicology**, v.92, p.551-556, 2014.

JIANG, Z.Y., HUNT, J.V., WOLFF, S.P. Ferrous ion oxidation in the presence of xylenol orange for detection of lipid hydroperoxide in low density lipoprotein. **Analytical Biochemistry**, v.202, n.2, p.384-389, 1992.

KAMARUZZAMAN, B.Y. et al. Heavy Metal Accumulation in Commercially Important Fishes of South West Malaysian Coast. **Research Journal of Environmental Sciences**, 2011.

KEEN, J. H.; HABIG, W. H.; JAKOBY, W. B. Mechanism for the several activities of the glutathione S-transferases. **Journal of Biological Chemistry**, v.251, n.20, p.6183-6188, 1976.

KHOSHNOOD, Z. Effects of Environmental Pollution on Fish: A Short Review. **Transylvanian Review of Systematical and Ecological Research**, v.19, n.1, p.49-60, 2017.

KLAASSEN, C. D.; LIU, J.; DIWAN, B. A. Metallothionein protection of cadmium toxicity. **Toxicology and Applied Pharmacology**, v. 238, n. 3, p. 215–220, 1 ago. 2009.

KLAASSEN, C. D.; WATKINS III, J. B. Fundamentos em Toxicologia de Casarett e Doull. 2^o edição, Porto Alegre, Artmed, 2012.

KROON, F.; STRETEN, C.; HARRIES, S. A protocol for identifying suitable biomarkers to assess fish health: A systematic review. **PloS one**, v. 12, n. 4, p. e0174762, 2017.

KRUPINA, K., GOGINASHVILI, A., CLEVELAND, D.W. Causes and consequences of micronuclei. **Current Opinion in Cell Biology**, 2021.

KUMAR, N. et al. Cellular stress and histopathological tools used as biomarkers in *Oreochromis mossambicus* for assessing metal contamination. **Environmental Toxicology and Pharmacology**, v.49, p.137-147, 2017.

KUMAR, N. et al. Metal determination and biochemical status of marine fishes facilitate the biomonitoring of marine pollution. **Marine Pollution Bulletin**, v.170, 2021.

LANA, P.C. et al. The subtropical estuarine complex of Paranaguá Bay, Brazil. In: Seeliger U, Lacerda LD, Kjerfve BJ (eds) Coastal marine ecosystems of Latin America. **Springer**, Berlin, p. 131–145, 2001.

LEÃO-BUCHIR, J. et al. BDE-99 (2,2-,4,4-,5 - pentain polybrominated diphenyl ether) induces toxic effects in *Oreochromis niloticus* after sub-chronic and oral exposure. **Environmental Toxicology and Pharmacology**, v.97, 104034, 2023.

LEHUN, A.L. et al. Genotoxic effects of urban pollution in the Iguaçu River on two fish populations. **Journal of Environmental Science and Health, Part A. Toxic/hazardous substances and environmental engineering**, v.56, p.984–991, 2021

LEITE, L. A. R. et al. Bioaccumulation and Health Risk Assessment of Trace Metal Contamination in the Musculature of the Trahira Fish (*Hoplias Malabaricus*) from Two Neotropical Rivers in Southeastern Brazil. **SSRN Electronic Journal**, 16 fev. 2021.

LI, L.; YANG, X. The essential element manganese, oxidative stress, and metabolic diseases: Links and interactions. **Oxidative Medicine and Cellular Longevity**, 2018.

LIEBEL, S., TOMOTAKE, M.E.M., OLIVEIRA-RIBEIRO, C.A. Fish histopathology as biomarker to evaluate water quality. **Ecotoxicology and Environmental Contamination**, v.8, p.9–15, 2013.

LIMA, M. C. B. et al. Biologia reprodutiva do peixe traíra, *Hoplias malabaricus* (Bloch, 1794) (Characiformes: Erythrinidae) no açude Marechal Dutra, Rio Grande do Norte, Brasil. **Biota Amazônia**, v. 7, n. 2, p. 21–25, 19 jun. 2017.

LÓPEZ-DOVAL, J.; BARATA, C.; DÍEZ, S. El uso de organismos como indicadores de la contaminación y evaluación del riesgo sobre el ecosistema acuático en el Embalse de Flix (Catalunya, ne de España). Em: POMPEO, M.; MOSCHINI-CARLOS, V.; NISHIMURA, P.Y.; DA SILVA, S.C.; LOPEZ-DOVAL, J.C. Ecologia de reservatórios e interfaces., Capítulo 1: organismos como indicadores de la contaminación. Instituto de Biociências da Universidade de São Paulo, São Paulo, 2015. p. 1-31.

LOZANO, I. E. et al. Comparison of scale and otolith age readings for trahira, *Hoplias malabaricus* (Bloch, 1794), from Paraná River, Argentina. **Journal of Applied Ichthyology**, v. 30, n. 1, p. 130–134, fev. 2013.

MAGALHÃES, D. P.; FERRÃO FILHO, A. S. Ecotoxicologia como ferramenta no biomonitoramento de ecossistemas aquáticos. **Oecologia Brasiliensis**, v. 12, n. 3, p. 355-381, 2008.

MALIK, D.S. et al. A review on impact of water pollution on freshwater fish species and their aquatic environment. In: **Advances in Environmental Pollution Management: Wastewater Impacts and Treatment Technologies**. Agro Environ Media - Agriculture and Environmental Science Academy, Haridwar, India, pp. 10–28, 2020.

MALLATT, J. Fish gill structural changes induced by toxicants and other irritants: A statistical review. **Canadian Journal of Fisheries and Aquatic Sciences**, v.42, p.630–648, 1985.

MANNERVIK, B. Glutathione Peroxidase. **Methods in Enzymology**, v. 113, p. 490–495, 1985.

MARINOVIĆ, Z. et al. Gill histopathology as a biomarker for discriminating seasonal variations in water quality. **Applied Sciences**, v.11, 2021.

MARQUES, D. K. S.; GURGEL, H. DE C. B.; LUCENA, I. Época de reprodução de *Hoplias malabaricus* Bloch, 1794 (Osteichthyes, Erythrinidae) da barragem do rio Gramame, Alhandra, Paraíba, Brasil. **Revista Brasileira de Zootecias**, v. 3, n. 1, p. 61–67, jun. 2001.

MARTINS, L.; PAZ, A. V.; BRENTANO, D. M. Avaliação da Geração de Micronúcleo em Junvenis de *Centropomus parallelus* (Robalo-Peva) Expostos a Diferentes Concentrações Salinas 1. **Revista Técnico-Científico do IFSC**, v. 2, n. 1, p. 13–16, 2010.

MCCORDS, J. M.; FRIDOVICH, I. Superoxide Dismutase an enzymic function for ERYTHROCUPREIN (HEMOCUPREIN)*. **Journal of Biological Chemistry**, v. 244, n. 22, p. 6049–6065, 1969.

MEISTER, A.; ANDERSON, M. E. GLUTATHIONE. **Annual Review of Biochemistry**, v. 52, p. 711–760, 1983.

MELA, M. et al. Effects of dietary methylmercury on liver and kidney histology in the neotropical fish *Hoplias malabaricus*. **Ecotoxicology and Environmental Safety**, v.68, n.3, p.426–435, 2007.

MELA, M. et al. Risks of waterborne copper exposure to a cultivated freshwater Neotropical catfish (*Rhamdia quelen*). **Ecotoxicology and Environmental Safety**, v. 88, p. 108–116, 1 fev. 2013.

MELA, M. et al. Mercury distribution in target organs and biochemical responses after subchronic and trophic exposure to Neotropical fish *Hoplias malabaricus*. **Fish Physiology and Biochemistry**, v.40, p.245–256, 2014.

MELO, R. M. C. et al. Comparative morphology of the reproductive system of seven species of ostariophysan fishes from the upper Das Velhas River, Brazil. **Journal of Morphology**, v. 278, n. 2, p. 170–181, 1 fev. 2017.

MILLS, G. C. HEMOGLOBIN CATABOLISM: I. GLUTATHIONE PEROXIDASE, AN ERYTHROCYTE ENZYME WHICH PROTECTS HEMOGLOBIN FROM OXIDATIVE BREAKDOWN*. **Journal of Biological Chemistry**, v. 229, n. 1, p. 189–197, 1957.

MONTEIRO, D. A.; RANTIN, F. T.; KALININ, A. L. Dietary intake of inorganic mercury: Bioaccumulation and oxidative stress parameters in the neotropical fish *Hoplias malabaricus*. **Ecotoxicology**, v. 22, n. 3, p. 446–456, abr. 2013.

MONTENEGRO, A. K. A. et al. Piscivory by *Hoplias aff. malabaricus* (Bloch, 1794): A question of prey availability? **Acta Limnologica Brasiliensia**, v. 25, n. 1, p. 68–78, 2013.

MONTES, C. et al. Evaluation of metal contamination effects in piranhas through biomonitoring and multi biomarkers approach. **Heliyon**, v.6, 2020.

MORAES, D. S. L.; JORDÃO, B. Q. Degradação de recursos hídricos e seus efeitos sobre a saúde humana. **Revista Saúde Pública**, v. 36. n. 3, p. 370-374, 2002.

MORAES, M. F. P. G.; BARBOLA, I. DE F. Hábito alimentar e morfologia do tubo digestivo de *Hoplias malabaricus* (Osteichthyes, Erythrinidae) da Lagoa Dourada, Ponta Grossa, Paraná, Brasil. **Acta Biológica Paranaense**, v. 24, n. (1-4), p. 1–23, 1995.

MOZANZADEH, M. T. et al. The effect of salinity on growth performance, digestive and antioxidant enzymes, humoral immunity and stress indices in two euryhaline fish species: Yellowfin seabream (*Acanthopagrus latus*) and Asian seabass (*Lates calcarifer*). **Aquaculture**, v.534, 736329, 2021.

MYERS, N. et al. Biodiversity hotspots for conservation priorities. **Nature**, v.403, p.853–858, 2000.

NORDBERG, M.; NORDBERG, G. F. Metallothionein and Cadmium Toxicology—Historical Review and Commentary. **Biomolecules**, v. 12, n.360, 2022.

OBIAKOR, M. O.; OKONKWO, J. C.; EZEONYEJIAKU, C. D. Eco-genotoxicology: Micronucleus Assay in Fish Erythrocytes as In situ Aquatic Pollution Biomarker: a Review. **Journal of Animal Science Advances** , v. 2, n. 1, p. 123–133, 2012.

OCCHI, T. V. T. **Biological Invasions and its Effects on Biodiversity**. Curitiba-PR, 2020. 59 p. Tese (Doutorado em Ecologia e Conservação)- Curso de Pós- Graduação em Ecologia e Conservação (PPGECO), Universidade Federal do Paraná- Setor de Ciências Biológicas, 2020.

OLIVEIRA, F. G. et al. Toxicological effects of anthropogenic activities in *Geophagus brasiliensis* from a coastal river of southern Brazil: A biomarker approach. **Science of the Total Environment**, v. 667, p. 371–383, 1 jun. 2019.

OLIVEIRA, H. W.; VALDES, S. A. C. FREQUÊNCIA DE MICRONÚCLEOS EM TILÁPIAS *OREOCHROMIS NILOTICUS* (PERCIFORMES, CICHLIDAE) DE PISCICULTURAS NO MUNICÍPIO DE MATUTINA (MG). **Revista do COMEIA**, v. 1, n. 1, p. 41–50, 2019.

OLIVEIRA, H.H.Q. et al. Gill Histopathological Biomarkers in Fish Exposed to Trace Metals in the Todos os Santos Bay, Brazil. **Biological Trace Element Research**, v.200, p.3388–3399, 2022.

PALHARES, D.; GRISOLIA, C. K. Comparison between the micronucleus frequencies of kidney and gill erythrocytes in tilapia fish, following mitomycin C treatment. **Genetics and Molecular Biology**, v. 25, n. 3, p. 281–284, 2002.

PANTALEÃO, S. DE M. et al. The piscine micronucleus test to assess the impact of pollution on the Japarutuba River in Brazil. **Environmental and Molecular Mutagenesis**, v. 47, n. 3, p. 219–224, abr. 2006.

PASTORINO, P. et al. Ecology of oxidative stress in the Danube barbel (*Barbus balcanicus*) from a winegrowing district: Effects of water parameters, trace and rare earth elements on biochemical biomarkers. **Science of the Total Environment**, v. 772, 10 jun. 2021.

PAULA, A. A.; RISSO, W. E.; MARTINEZ, C. B. DOS R. Effects of copper on an omnivorous (*Astyanax altiparanae*) and a carnivorous fish (*Hoplias malabaricus*): A comparative approach. **Aquatic Toxicology**, v. 237, 1 ago. 2021.

PAULINO, M.G. et al. Biotransformations, Antioxidant System Responses, and Histopathological Indexes in the Liver of Fish Exposed to Cyanobacterial Extract. **Environmental Toxicology and Chemistry**, v.39, p.1041–1051, 2020.

PEREIRA, N. J. et al. Biomarcadores histológicos em brânquias de peixes na avaliação da contaminação ambiental do Rio Mearim, Nordeste brasileiro. **Brazilian Journal of Development**, v. 6, n. 9, p. 68063–68079, 2020.

PEREIRA, R.S. Identificação e Caracterização das Fontes de Poluição em Sistemas Hídricos. **Revista Eletrônica de Recursos Hídricos**, v. 1 (1), p. 23-40, 2004.

PESSOA, E. K. R. et al. Morfologia Comparativa do Trato Digestório dos Peixes *Hoplias malabaricus* e *Hypostomus puarum* do Açude Marechal Dutra, Rio Grande do Norte, Brasil. **Biota Amazônia**, v. 3, n. 1, p. 48–57, 30 jun. 2013.

QU, R. et al. Metal accumulation and oxidative stress biomarkers in liver of freshwater fish *Carassius auratus* following in vivo exposure to waterborne zinc under different pH values. **Aquatic Toxicology**, v.150, p.9–16, 2014.

RABITTO, I.S. et al. Effects of dietary Pb(II) and tributyltin on neotropical fish, *Hoplias malabaricus*: Histopathological and biochemical findings. **Ecotoxicology and Environmental Safety**, v.60, p.147–156, 2005.

RATHI, B. S.; KUMAR, P. S.; VO, D.N. Critical review on hazardous pollutants in water environment: Occurrence, monitoring, fate, removal technologies and risk assessment. **Science of The Total Environment**, v. 797, 2021.

R CORE TEAM. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria, 2017. Disponível em:< <https://www.R-project.org/>>.

REIS, C.S. et al. Avaliação da Atividade Antrópica no Rio Guaraguaçu (Pontal do Paraná, Paraná). **Engenharia Sanitária e Ambiental**, v.20, n.3, p. 389-394, 2015.

REIS, T. et al. *Hoplias* aff. *malabaricus* Bloch, 1794 (Characiformes: Erythrinidae) parasites. **Arquivos do Instituto Biológico**, v. 84, n. 1–5, 1 fev. 2017.

RESH, V. H. Which group is best? Attributes of different biological assemblages used in freshwater biomonitoring programs. **Environmental Monitoring and Assessment**, v. 138, p. 131– 138. 2008.

RODRIGUEZ-CEA, A.; AYLLON, F.; GARCIA-VAZQUEZ, E. Micronucleus test in freshwater fish species: An evaluation of its sensitivity for application in field surveys. **Ecotoxicology and Environmental Safety**, v.56, p.442–448, 2003.

ROVER JÚNIOR, L. et al. SISTEMA ANTIOXIDANTE ENVOLVENDO O CICLO METABÓLICO DA GLUTATIONA ASSOCIADO A MÉTODOS ELETROANALÍTICOS NA AVALIAÇÃO DO ESTRESSE OXIDATIVO. **Química Nova**, v. 24, n. 1, p. 112–119, 2001.

SAAD ABDELKARIM, M. Biomonitoring and bioassessment of running water quality in developing countries: A case study from Egypt. **Egyptian Journal of Aquatic Research**, 2020.

SALEH, Y.S.; MARIE, M.A.S. Use of *Arius thalassinus* fish in a pollution biomonitoring study, applying combined oxidative stress, hematology, biochemical and histopathological biomarkers: A baseline field study. **Marine Pollution Bulletin**, v.106, p.308–322, 2016.

SALGADO, L.D. et al. Integrated assessment of sediment contaminant levels and biological responses in sentinel fish species *Atherinella brasiliensis* from a sub-tropical estuary in south Atlantic. **Chemosphere**, v.219, p.15–27, 2019.

SALGADO, L. D. et al. Sediment contamination and toxic effects on Violet Goby fish (*Gobioides broussonnetii* - Gobiidae) from a marine protected area in South Atlantic. **Environmental Research**, v. 195, 1 abr. 2021.

SANTANA, M.S. et al. Diffuse sources of contamination in freshwater fish: Detecting effects through active biomonitoring and multi-biomarker approaches. **Ecotoxicology and Environmental Safety**, v.149, p.173–181, 2018.

SATO, R. Y.; COSTA, A. P. L.; PADIAL, A. A. The invasive tropical tanner grass decreases diversity of the native aquatic macrophyte community at two scales in a subtropical tidal river. **Acta Botanica Brasilica**, v. 35, n. 1, p. 140–150, 2021.

SAVASSI, L.A. et al. Heavy metal contamination in a highly consumed Brazilian fish: immunohistochemical and histopathological assessments. **Environmental Monitoring and Assessment**, v.192, 2020.

SEDLAK, J.; LINDSAY, R. H. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. **Analytical Biochemistry**, v.25, p.192-205, 1968.

SFAKIANAKIS, D.G. et al. Effect of heavy metals on fish larvae deformities: a review. **Environmental Research**, v.137, p.246–255, 2015.

SHARMA, A. et al. Worldwide pesticide usage and its impacts on ecosystem. **SN Applied Sciences**, 2019.

SILVA DE ASSIS, H.C. Der Einsatz von Biomarkern zur summarischen Erfassung vom Gewässerverschmutzungen. PhD thesis, Berlin Technical University, Berlin, Germany, 1998.

SILVA DE ASSIS, H. C. et al. Hematologic and hepatic responses of the freshwater fish *Hoplias malabaricus* after saxitoxin exposure. **Toxicon**, v. 66, p. 25–30, maio 2013.

SILVA, D. C. V. R.; POMPÊO, M.; PAIVA, T. C. B. A Ecotoxicologia no contexto atual no Brasil. *Ecologia de reservatórios e interfaces*. São Paulo: Instituto de Biociências, p. 460, 2015.

SILVA, F.F.G. Composição e distribuição da ictiofauna do rio Guaraguaçu (Paranaguá, Paraná-BR) e biologia alimentar de três espécies. Curitiba-PR, 2008. 100 p. Dissertação (Mestrado Ciências Biológicas, área de concentração Zoologia) - Curso de Pós Graduação em Ciências Biológicas - Zoologia, Setor de Ciências Biológicas, Universidade Federal do Paraná., 2008.

SILVA, V. et al. Conhecendo os principais solos do Litoral do Paraná : abordagem para educadores do ensino fundamental e médio. Sociedade Brasileira de Ciência do Solo, Núcleo Estadual do Paraná– Matinhos (PR) : UFPR, 32 p. ISBN 978-85-86504-10-5, 2013.

SINGO, J. M.; ARAÚJO-RAMOS, A. T.; ROCHA, J. R. C. Physical-Chemical Characterization of Peri River, Pontal do Paraná, PR, Brazil. **International Journal of Advanced Engineering Research and Science**, v.7, n.5, p.314-323, 2020.

SKIDMORE J.F. Toxicity of Zinc Compounds to Aquatic Animals, With Special Reference to Fish. **The Quarterly Review of Biology**, v.39, p.227–248, 1964.

SOUZA-BASTOS, L. R. et al. Evaluation of the water quality of the upper reaches of the main Southern Brazil river (Iguaçu River) through in situ exposure of the native siluriform *Rhamdia quelen* in cages. **Environmental Pollution**, v. 231, p. 1245-1255, 2017.

SOUZA, G.L.C. et al. PhysicalChemical Parameters Evaluation of Pery River Waters in Pontal do Paraná, PR. **IOSR Journal of Environmental Science, Toxicology and Food Technology**, v.13, n.6, p. 69-78, 2019.

SOUZA, R.M. et al. Occurrence, impacts and general aspects of pesticides in surface water: A review. **Process Safety and Environmental Protection**, v.135, p.22–37, 2020.

STASZCZAK, I.; ROCHA, J. R. C. Avaliação da influência das marés nos parâmetros físico-químicos presentes nas águas do rio Guaraguaçu, PR. **PERIÓDICO TCHÊ QUÍMICA**, Vol. 15, N. 30. Porto Alegre, RS. 2018. ISSN 2179-0302.

TESSER, T.T., DA ROCHA, C.M., CASTRO, D. Metal contamination in omnivores, carnivores and detritivores fish along the Tramandaí River Basin, RS, Brazil. **Environmental Nanotechnology, Monitoring & Management**, v.16, 2021.

THIRUMOORTHY, N. et al. A Review of Metallothionein Isoforms and their Role in Pathophysiology. **World Journal of Surgical Oncology**, v. 9, n. 54, 2011.

TORRES, G.V. Dinâmica da ocupação da terra e transformação da paisagem da bacia hidrográfica do rio Guaraguaçu, litoral do Paraná. Matinhos-PR, 2019. 91 p. Dissertação (Mestrado em Desenvolvimento Territorial Sustentável) - Curso de Pós Graduação em Desenvolvimento Territorial Sustentável (PPGDTS), Universidade Federal do Paraná- Setor Litoral, 2019.

TUNDISI, J. G.; TUNDISI, T. M. **Recursos hídricos no século XXI**. São Paulo: Oficina de Textos, 2011.

U.S. EPA. “Method 3520C (SW-846): Continuous Liquid-Liquid Extraction,” Revision 3. 1996a. Disponível em: <<https://www.epa.gov/hw-sw846/sw-846-test-method-3520c-continuous-liquid-liquid-extraction>>. Accessed on October 28, 2022.

U.S. EPA. “Method 3052 (SW-846): Microwave Assisted Acid Digestion of Siliceous and Organically Based Matrices”. 1996b. Disponível em: <<https://www.epa.gov/hw-sw846/sw-846-test-method-3052-microwave-assisted-acid-digestion-siliceous-and-organically-based>>. Accessed on January 17, 2023.

U.S. EPA. “Method 8270E (SW-846): Semivolatile Organic Compounds by Gas Chromatography/ Mass Spectrometry (GC/MS),” Washington, DC, 2014. Disponível em: <<https://www.epa.gov/esam/epa-method-8270e-sw-846-semivolatile-organic-compounds-gas-chromatographymass-spectrometry-gc>>. Accessed on November 08, 2022.

U.S. GEOLOGICAL SURVEY. The Water in You: Water and the Human Body. Estados Unidos. 2019. Disponível em: < https://www.usgs.gov/special-topic/water-science-school/science/water-you-water-and-human-body?qt-science_center_objects=0#qt-science_center_objects >. Acessado em: 20 de junho de 2021.

VAN DER OOST, R.; BEYER, J.; VERMEULEN, N.P. E. Fish bioaccumulation and biomarkers in environmental risk assessment: a review. **Environmental Toxicology and Pharmacology**, v. 13, n. 2, p. 57-149, 2003.

VIANA, H. C. et al. Aggregation of hepatic melanomacrophage centers in *S. herzbergii* (Pisces, Ariidae) as indicators of environmental change and well-being. **Arquivo Brasileiro de Medicina Veterinária e Zootecnia**, v.73, n.4, p.868–876, 2021.

VIARENGO, A. et al. A simple spectrophotometric method for metallothionein evaluation in marine organisms: an application to Mediterranean and Antarctic molluscs. **Marine Environmental Research**, v.44, p.69–84, 1997.

VITULE, J.R.S.; UMBRIA, S.C.; ARANHA, J.M.R. Introduction of the African catfish *Clarias gariepinus* (BURCHELL, 1822) into Southern Brazil. **Biological Invasions**, v. 8, p.677–681, 2006.

VITULE, J.R.S. Distribuição, abundância e estrutura populacional de peixes introduzidos no rio Guaraguaçu, Paranaguá, Paraná. Curitiba-PR, 2008. 162 p. Tese (Doutorado em Ciências Biológicas área de concentração – Zoologia) - Curso de Pós-Graduação em Ciências Biológicas – Zoologia, Setor de Ciências Biológicas da Universidade Federal do Paraná, 2008.

VOIGT, C.L. et al. Bioconcentration and bioaccumulation of metal in freshwater Neotropical fish *Geophagus brasiliensis*. **Environmental Science and Pollution Research**, v.22, p.8242–8252, 2015.

WINSTON, G.W.; DI GIULIO, R. T. Prooxidant and antioxidant mechanisms in aquatic organisms. **Aquatic Toxicology**, v. 19, n. 2, p. 137-161, 1991.

WU, T. et al. Use of conductivity to indicate long-term changes in pollution processes in Lake Taihu, a large shallow lake. **Environmental Science and Pollution Research International**, v. 27, n.17, p.21376–21385, 2020.

YADAV, K. K.; TRIVEDI, S. P. Sublethal exposure of heavy metals induces micronuclei in fish, *Channa punctata*. **Chemosphere**, v. 77, n. 11, p. 1495–1500, 2009.

ZARONI, M. J., SANTOS, H. G. Empresa Brasileira de Pesquisa Agropecuária (Embrapa): Espodossolos. 2021. Disponível em: <https://www.embrapa.br/agencia-de-informacao-tecnologica/tematicas/solos-tropicais/sibcs/chave-do-sibcs/espodossolos>. Acessado em 2 de maio de 2023.

ZEE. Zoneamento Ecológico Econômico do Litoral do Paraná. Decreto Estadual nº 4.996 de 05 de setembro de 2016. Curitiba. 2016.